

Review

A gene catalogue of the amphioxus nervous system

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Received: 2006.02.14; Accepted: 2006.05.21; Published: 2006.05.22

The elaboration of extremely complex nervous systems is a major success of evolution. However, at the dawn of the post-genomic era, few data have helped yet to unravel how a nervous system develops and evolves to complexity. On the evolutionary road to vertebrates, amphioxus occupies a key position to tackle this exciting issue. Its "simple" nervous system basically consists of a dorsal nerve cord and a diffuse net of peripheral neurons, which contrasts greatly with the complexity of vertebrate nervous systems. Notwithstanding, increasing data on gene expression has faced up this simplicity by revealing a mounting level of cryptic complexity, with unexpected levels of neuronal diversity, organisation and regionalisation of the central and peripheral nervous systems. Furthermore, recent gene expression data also point to the high neurogenic potential of the epidermis of amphioxus, suggestive of a skin-brain track for the evolution of the vertebrate nervous system. Here I attempt to catalogue and synthesise current gene expression data in the amphioxus nervous system. From this global point of view, I suggest scenarios for the evolutionary origin of complex features in the vertebrate nervous system, with special emphasis on the evolutionary origin of placodes and neural crest, and postulate a pre-patterned migratory pathway of cells, which, in the epidermis, may represent an intermediate state towards the deployment of one of the most striking innovative features of vertebrates: the neural crest and its derivatives.

Keywords: Non-neural ectoderm, neurogenic potential, cryptic complexity, neural crest, placodes.

1. Introduction

The evolution of the nervous system in the Animal Kingdom is currently under major debate in Evolutionary Developmental Biology and Comparative Embryology [1]. In this context, the evolutionary path that leads to the morphological complexity of the nervous system passes through the invertebrate/vertebrate transition. At the beginning of the XXth century, the cephalochordate amphioxus occupied the most prominent role in illuminating, at the morphological level, such a transition (see [2] for an historical perspective). During the last decade, increasing information on the molecular aspects of amphioxus development has revealed a number of previously hidden features. One prominent example is the molecular regionalisation of the amphioxus nervous system, which is greatly hindered by its anatomical simplicity. The central nervous system of an adult amphioxus is a mere tubular nerve cord lying dorsally over the notochord. The nerve cord and the notochord cover together the entire length of the animal, except in the anterior region, where only the notochord reaches the rostral tip of the body (Fig. 1). This exclusive rostral expansion of the notochord is a hallmark of the taxon Cephalochordata (corda - for corda dorsalis, original name of the notochord - in the head). The only externally visible landmarks of the amphioxus nerve cord are discrete enlargements at both the anterior and posterior ends, regarded as the

anterior vesicle and the caudal ampulla, respectively. Transiently during embryonic development, in young larvae, the anterior swelling is commonly known as cerebral vesicle and is much more pronounced than in adults. Apart from the serially repeated exit points of the dorsal nerves, the nerve cord of amphioxus is devoid of rhombomeres or other external morphological indications of segmentation. The outgoing dorsal nerves, unsupplied with ganglia, contribute to the peripheral nervous system by innervating most of the organs of the body, shaping an irregular nerve net, where a number of peripheral neurons are also accommodated [3]. In contrast with the anatomically non-regionalised central nervous system and the apparently disorganised peripheral nervous system, the discrete expression patterns of distinct genes and the specific distribution of certain proteins outline a completely different chronicle for the nervous system of amphioxus. Most of the neural genes characterised to date reveal gapped expression domains within the developing central nervous system, and restricted expression to certain cell subpopulations within the embryonic epidermis. Retinoic acid treatments greatly affect these gene expression domains [4], suggesting a well-controlled pre-patterning that defines distinct territories, not only within the neural ectoderm but also within the epidermis of amphioxus embryos.

After two centuries of heated debate, amphioxus is still an epicentre for the controversial evolutionary

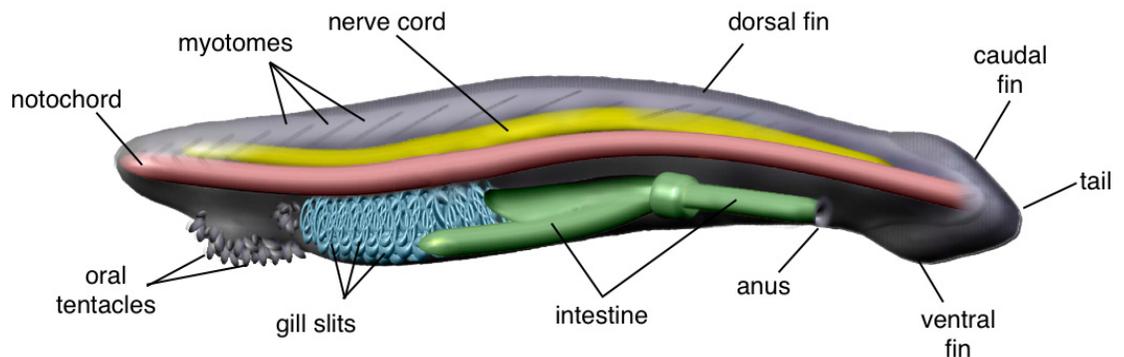
origins of vertebrates. Whatever its actual position, as the sister group of vertebrates, as the earliest chordate, or as (less likely) the earliest deuterostome [5], its privileged phylogenetic position sets amphioxus in the appropriate place to cast light on some of the main transitions that occurred during animal evolution: the origin of deuterostomes, the origin of Chordates, or the origin of vertebrates, all of them correlated with novel evolutionary features, and some of them linked to the elaboration of the most complex nervous systems. The relative phylogenetic position of cephalochordates and urochordates has very recently received a lot of attention [6]. Both cephalochordates and urochordates are invertebrate chordates that share with vertebrates certain anatomical features: a notochord, pharyngeal slits and a dorsal hollow nerve cord. However, these pre-vertebrate characteristics are lost in tunicates after metamorphosis, while they are maintained for life in cephalochordates. Furthermore, cephalochordates but not urochordates possess additional vertebrate-like characters, such as a post-anal tail and segmentally arranged muscles (myomeres) (Fig. 1). Both clades include the only invertebrates able to form a dorsal nerve cord through neurulation. However, unlike in urochordates, in amphioxus the nervous system seems to be developmentally active beyond metamorphosis. This is supported by modern anatomy [3], which shows that cytoarchitectural reorganisations should occur within the nerve cord of amphioxus until reaching a definitive internal structure in the adult.

The present communion between modern morphology and the use of molecular biology tools is revealing an unexpected variety of cell types, territories and boundaries along the antero-posterior axis of the nerve cord and on the epidermis of amphioxus. Here I attempt to summarize distinct patterns of gene expression, giving an overall view of the formation of the central and peripheral nervous systems of amphioxus and suggesting the possible origin of two characteristic neural features in vertebrates: the placodes and the neural crest.

consequence of the animal-vegetal polarity of the oocyte, induced by the sperm entry point during fertilisation. At the onset of gastrulation, the vegetal pole of the blastula invaginates, leaving a wide-open blastopore, which subsequently closes. As soon as invagination begins, *AmphiWnt1* turns on around the blastoporal opening in conjunction with *AmBra*, followed by the expression of *AmphiWnt8* and a down-regulation of β -catenin in the invaginating mesendoderm [7,8]. This territorial area homing the expression of these genes is soon segregated into the blastoporal lips, which remain on the posterior side of the embryo and will contribute to the tail bud later in development [9].

Other amphioxus Wnt genes are also expressed around the blastopore in an overlapping pattern to that of *AmphiNotch* and *AmBra* [7, 10, 11]. This indicates that both the Wnt and Notch pathways may act as a posterior signalling centre involved in the specification and maintenance of the posterior identity. Whether the Notch and Wnt signalling systems are involved in the specification of the dorso-ventral axis remains unclear. However, it has been suggested that Wnt signals may be involved in restricting expression of genes such as *AmphiSox1/2/3*, *AmphiDRAL* and *AmphiDll* to the ectoderm [7]. *AmphiSox1/2/3* and *AmphiNeurogenin* are the earliest known markers for the presumptive neuroectoderm in amphioxus [12]. The expression of both largely overlaps at the early gastrula, on the outer dorsal epiblast (ectoderm), thus indicating the existence of a dorso-ventral axis already at this stage. Opposite *AmphiSox1/2/3* and *AmphiNeurogenin*, *AmphiDRAL* is expressed in the non-neural ectoderm, suggesting that the neural/non-neural boundary might become delimited early in development [13]. Throughout gastrulation, some of the genes involved in neural patterning are expressed in the dorsal blastoporal lip, which has been regarded as the homolog of the amphibian Spemann's organiser [14], crucially involved in neural induction. In vertebrates, the early *nodal* expression is involved in the establishment of the Spemann's organiser and

Figure 1. Morphological features of an adult amphioxus. Lateral view showing the internal anatomy and part of the overlying segmented muscle blocks (myomeres).



2. Early gene expression

In amphioxus, the embryonic anterior-posterior axis is determined early (by the time of gastrulation), as a

induces the expression of organiser-specific genes such as *gooseoid*. In amphioxus, *AmphiNodal's*

strongest expression is in the inner hypoblast (mesendoderm) of the dorsal blastoporal lip [15]. Therefore, *AmphiNodal* might well induce the expression of *gooseoid* in the dorsal blastoporal lip, where it overlaps with other organiser-related genes such as *AmHNF3*, *AmBra* and *AmphiFoxD*, but not *AmphiOtx* [11, 16-19]. This incomplete overlapping expression occurs for a short period of time and, in comparison with vertebrates, it is slightly delayed in development. Thus, it is unclear to what extent the dorsal blastoporal lip functions, if it does, as a dorsal-ventral organiser, though it may have some properties related to the Spemann's organiser, such as influencing the formation of the neural plate and the underlying chordamesoderm.

As gastrulation proceeds, some genes begin to be restrictively expressed in certain regions of the neuroectoderm, which may be an anticipation of the cryptic regionalisation of the neural tube and the epidermis observed in succeeding stages. *Islet* and *AmphiPax6* are first solely expressed in the anterior neuroectoderm, whereas *AmphiEvxA* is expressed just in the posterior neuroectoderm and *AmphiMsx* in the middle part [20-23]. The homogeneous expression of *AmphiSox1/2/3* and *AmphiNeurogenin* in the dorsal epiblast is gradually restricted only to the neural ectoderm [12]. *AmphiZic* and *AmphiVent*, which were earlier widely expressed dorsally, become restricted to the lateral regions of the nascent neural plate [14, 24].

Conversely, other genes are expanded to the neuroectoderm, as shown by *AmphiBrn1/2/4*, *AmphiPax3/7* and the various *AmphiHairy* genes. As the gastrula elongates to reach the earliest neurula stage, the expression of *AmphiBrn1/2/4*, initially located in the dorsal epiblast, gradually spans from the posterior to the anterior neural plate [25]. The initial expression of *AmphiPax3/7* in the dorsal presumptive axial and paraxial mesoderm is additionally extended to the dorsal ectoderm, where it also appears at the lateral edges of the presumptive neural plate [26]. Likewise, the *AmphiHairy* genes also label the forming neural plate, in this case accompanied by a complementary striped expression in the presumptive somitic mesoderm [27]. This raises the recurrent question on the effect that mesoderm segmentation might exert on the molecular regionalisation of the neural tube in amphioxus. Although not segmental, the mesodermal expression of *Id* and *AmphiDmbx* could be tentatively related to this issue, since in vertebrates they have a well-defined location within the developing central nervous system [28, 29]. Furthermore, the apposed ectodermal-mesodermal expression also observed for the amphioxus *snail* gene, which is detected in the dorsal presumptive somitic mesoderm and the overlying edges of the forming neural plate, and several other genes (e.g. *AmphiZic*, *AmphiVent*) also suggest a collaborative role between the two germ layers [30].

3. Patterning of the neural tube

At the end of gastrulation, as the neural plate starts to form, the amphioxus embryo becomes more vertebrate-like. However, in contrast with vertebrates, neural folds do not form during amphioxus neurulation. Instead, the non-neural ectoderm detaches from the edges of the open neural plate, overgrowing laterally over the neural plate and fusing in the dorsal midline. As the non-neural ectoderm covers the neural plate, the lateral edges of the latter curl up dorsally and fuse in the dorsal midline, underneath the non-neural ectoderm, to form the neural tube. Once the neural tube is formed, it stills remains anteriorly open through the neuropore. Subsequently, as the neurula elongates, the posterior neural tube and the adjacent notochord and somites stem from the tail bud, which probably still maintains some of the organiser properties mentioned above. Throughout neurulation, the various gene expression patterns are highly dynamic within the central nervous system (Table 1) and reveal discrete molecular boundaries, most of them hidden at a morphological level.

The central nervous system of amphioxus neurulae, in its advanced stages, consists of a dorsal neural cord lacking constrictions along its rostrocaudal axis. The only regional difference lies at the anterior end, only regional difference lies at the anterior end where the neural canal is slightly dilated into a cerebral vesicle, anterior to the level of somite 1 and up to the anterior part of somite 2. Most of the cerebral vesicle is dominated by the expression of *AmphiOtx*, yet with a gap of expression halfway along its anterior-posterior axis. The anterior domain of *AmphiOtx* covers a region that includes the infundibular organ. This organ is the source of the Reissner's fiber, which fills the central neural canal, and is probably the homologue of the subcommisural organ of the vertebrate diencephalon [16]. In this region, *AmphiBF-1* labels a subset of *AmphiOtx*-positive cells [31]. Similarly, *AmphiDll*-positive cells are also embedded in the rostral domain of *AmphiOtx*, but located more anteriorly than *AmphiBF-1*, in the preneuroporal region [32]. *AmphiD1* is also expressed in the cerebral vesicle at the level of the presumptive infundibular organ, while *AmphiTH* is expressed at the level of the presumptive lamellar organ, which in turn is covered by the caudal expression of *AmphiOtx* [33,34]. These sharp expressions probably reflect a complex internal morphology within the cerebral vesicle; this is also observed for *AmphiPax6*, which is expressed in a few cells of the developing frontal eye [20]. The posterior expression of *AmphiOtx* overlaps with that of *AmphiDll*, *AmphiPax6*, *AmphiSim* and the anteriormost expression of *AmphiFoxB*. The delimited expression of *AmphiSim* in this area is especially interesting, since it is a marker of the posterior diencephalon and midbrain in vertebrates. However, amphioxus orthologues of other vertebrate midbrain and MHB (Midbrain-Hindbrain Boundary) markers,

such as *AmphiEvx*, *AmphiEn*, *AmphiPax2/5/8* and *AmphiWnt1*, have failed to provide evidence for a true midbrain and MHB homolog in amphioxus [29,35]. Both *AmphiBf-1* and *AmphiDll* are orthologue genes to those marking the vertebrate telencephalon and, in the case of *AmphiDll*, other parts of the vertebrate diencephalon as well. Together with the expression of *AmphiOtx*, the orthologues of which are markers of the vertebrate telencephalon, diencephalon and midbrain, and the territorial domain occupied by *AmphiSim*, combined gene expression suggests that the amphioxus cerebral vesicle is largely homologous to the vertebrate forebrain (telencephalon plus diencephalon), although certain homology with the midbrain cannot be completely ruled out.

In the hypothetical absence of a midbrain homolog in amphioxus, the territory posterior to the cerebral vesicle, caudal to *AmphiOtx* expression at the level of somite 2, has been widely seen as a homolog of the craniate hindbrain. Following the expression of *AmphiOtx*, amphioxus hindbrain is subjugated to the expression of different *Hox* genes, partially overlapped along the anterior-posterior axis. Spatially, the first *Hox* gene expressed in the anterior hindbrain is *AmphiHox1* (somites 2-4), followed by *AmphiHox3* (somites 4-8) and *AmphiHox4* (somites 6-8) [36]. *AmphiFoxB* is expressed segmentally as well, concomitantly with *Hox* genes, though it shows a prominent gap of expression at the level of somite 5, where the pigment spot will form and, intriguingly, where the expression of other segmental genes is also disrupted (e.g. *Mnx*, *Islet*) [35].

Other genes broadly expressed within the neural tube of amphioxus are: *AmphiSox1/2/3*, *AmphiNetrin*, *AmphiF-spondin*, the different *Hairy* genes and *AmphiBrn1/2/4*. All of them label certain fractions of the hindbrain and some of them include some spots of expression within the cerebral vesicle. While *AmphiSox1/2/3* is excluded along the midline and the anterior part of the neural tube, *AmphiNetrin* shows a fairly complementary pattern, being expressed just along the midline [12,37]. Notwithstanding, *AmphiNetrin* expression in the floor plate lasts longer than *AmphiSox1/2/3* expression does and always overlies its own expression in the dorsal notochord, even below the cerebral vesicle (from which it is excluded), until the larval stages, when *AmphiNetrin* is lost at both anterior and posterior ends of the notochord. The long-lasting expression of *AmphiNetrin* in the floor plate suggests a role of the amphioxus floor plate in the control of axon path finding, possibly guiding newborn neurons within the nerve cord [37]. *AmphiF-spondin* expression by the end of neurulation largely overlaps the expression of *AmphiNetrin*, though covering broader areas within the neural tube, including the cerebral vesicle, where it shows a gap of expression just on the ventral side [38]. Similar to *AmphiSox1/2/3*, *AmphiBrn1/2/4* is excluded from the midline, but is additionally expressed in the cerebral vesicle with the same gap of expression as the one observed for the *AmphiF-spondin* gene [25].

Throughout the neural tube, amphioxus *Hairy* genes are expressed in complementary patterns. *AmphiHairyB* is expressed in the anterior neural tube, spatially followed by *AmphiHairyC* and *AmphiHairyD*, being *AmphiHairyA* the most restricted one to the posterior neural tube. Independently, all of them show gapped expression and overlap at certain levels, just in some cases filling the gaps of one another. As mentioned previously, in general the expression of the different *AmphiHairy* genes in the neural tube is accompanied by a striped expression in the somitic mesoderm, but in the case of *AmphiHairyC* and *AmphiHairyD*, it is further escorted by their expression in the notochord [27]. The apposed expression of the notochord and the neural plate is also observable for *AmphiNetrin*, *AmHNF3* and *AmphiHh*, at neurula stages [17,37,39]. Though the expression of these three genes is undetectable in the ventral cerebral vesicle, it is clearly seen throughout the rest of the neural tube and along the whole length of the notochord, including the portion underlying the cerebral vesicle. In vertebrates, the notochord is a transient embryonic structure that acts as an organiser inducing the floor plate and cooperating in the establishment of the dorso-ventral axis of the neural tube and somites. Although in amphioxus this structure is permanent, based on comparable gene expression patterns, it may well have properties similar to those of the vertebrate notochord, yet with a longer period of action that could extend beyond metamorphosis. In this regard, however, the role of the anterior notochord is obscured by the fact that genes such as *AmphiNetrin*, *AmHNF3* or *AmphiHh* seem to be unable to induce floor plate fates in the ventral cerebral vesicle, which suggests that the cells lying ventrally in the cerebral vesicle may not be competent to respond to their signals [37].

4. Iterative gene expression patterns

The cryptic internal segmentation of the amphioxus neural tube has been visualized by iterative gene expression patterns. As illustrative examples, *AmphiKrox*, *shox*, *islet* and *AmphiMnx* share a one somite-wide periodicity of expression throughout the neural tube. All of them are expressed in the *Hox*-delineated hindbrain regions, though not all their patterns are in the same phase, indicating that different sets of cells take part in the iteration. Both *shox* and *islet* transcripts are located at the level of somite boundaries, while *AmphiKrox* transcripts are centralised at the level of somites and *AmphiMnx* ones are close to the posterior border of the somites [39-41]. The repetitive pattern appears more synchronised adjacent to the first four somites and is altered at the level of somite number 5, as occurs with the segmental expression of *AmphiFoxB*.

Similarly, *AmphiCoe* and *AmphiElav* are also iteratively expressed along the nerve cord during neurulation [42-44]. The *AmphiCoe*-expressing cells within the nerve cord show a periodicity matching the adjacent somites, in a pattern similar to those of *AmphiFoxB* and *AmphiNeurogenin*, at the same

developmental stages [12,35]. Unlike *AmphiKrox*, *shox* and *AmphiMnx* expression, the *AmphiCoe* transcripts are not solely confined to the ventral part of the nerve cord, since they are also present in dorsal cells lying in comparable positions to those expressing *islet*. Interestingly, the dorsal expression of both *islet* and *AmphiCoe* only occurs beyond the level of somite 5, where the dorsal cells expressing them are displaced anteriorly to the accompanying expressing cells in the ventrolateral floor of the nerve cord [41]. *AmphiElav* expression throughout neurulation is concomitant with that of the abovementioned iteratively expressed genes. However, though serially arranged, *AmphiElav*-expressing cells do not clearly match somite boundaries and are located on both the dorsal and ventral sides of the nerve cord, at all levels posterior to the cerebral vesicle [44].

In most cases, the iterative expression of these genes is maintained up to the larval stages, being down-regulated, but not completely missing, at about three days post-fertilisation. Similar expression patterns have been described for other genes, such as *AmphiNk2-1*, *AmphiNk2-2*, *AmphiFringe* or *AmphiMsx*, although their transcripts are not detected in the neural tube that far in development [21, 45-47]. The four genes are segmentally expressed in the neural tube at mid-neurula stages, just after mesoderm segmentation has commenced, but fade away from these locations by the end of neurulation. Again, this transient expression may correlate mesoderm segmentation with the development of internal metameric neural structures. However, other genes, such as *AmphiDach* and *AmphiERR*, show an iterative pattern of expression, most obviously at larval stages [48,49]. In both cases, the series of positive cells have a caudal limit of expression at the level of somites 5-6. It would seem that the only genes iteratively expressed beyond the level of somite 5 up to the latest stages of development analysed are *AmphiCoe*, *AmphiElav* and *AmphiNeurogenin*, but always excluding the most posterior part of the neural tube [12,43,44]. Since these genes are early markers for neuronal fates, this might suggest that posterior neuronal fates are delayed in development until later larval stages, at which their expression has not yet been analysed.

The combinatorial iterative expression of all these genes, at different times and in distinct subsets of cells, suggests that the amphioxus nerve cord is internally organised in a variety of neurons, segmentally arranged in a dorso-ventral and an antero-posterior order, which might affect their relative connectivity and consequently their particular functions. Although it is difficult to correlate the internal morphology of the embryonic and adult nerve cord, the differentiation events associated with the expression of the above-mentioned genes might only reflect the preliminary organisation of the anterior part of the adult nerve cord, which according to Wicht and Lacalli spans up to the level of myomere number 11 [3]. Given that the intermediate and posterior parts of the adult nerve cord show a slightly distinct

internal organisation, these regions could be morphogenetically delineated later in development. Similar early genetic programs could be acting at advanced larval stages and even beyond metamorphosis, when the embryo is still posteriorly elongating and dramatic morphological changes occur along the body, including the formation of certain structures that will be subsequently innervated by the descending branches of the dorsal nerves. However, this possibility is still to be explored, since the metamorphic and post-metamorphic larvae of amphioxus have rarely been used for gene expression studies.

5. Neural expression in the non-neural ectoderm

In recent years, the epidermis of amphioxus has received particular attention [50], as a number of genes are specifically expressed by distinct epidermal cell populations during amphioxus embryogenesis, which indicates that distinct genetic programmes are switched on in particular cells and territories of epidermal tissue, previously thought to be uniform. Some of these genes, such as *AmphiCoe*, *AmphiElav* and *AmphiTrk*, spread spottily over the epidermal surface of the embryo [43,44,51]. While *AmphiCoe* and *AmphiElav* are also expressed in the central nervous system, by the time they do in epidermal cells, *AmphiTrk* expression is solely confined to the non-neural ectoderm during neurulation, yet is restrictively expressed in the central nervous system in adult amphioxus. Within the neuroectoderm, all three of these genes are expressed in developing neuronal precursors, or putative mature neurons in the case of *AmphiTrk*. Noticeably, the respective orthologue genes in vertebrates are widely expressed in neuronal precursors during embryogenesis. All *AmphiCoe*, *AmphiElav* and *AmphiTrk* genes are expressed by scattered individual epidermal cells distributed over the ventrolateral flanks of neurulae. These cells show a distinctive elongated morphology that clearly sets them apart from surrounding epidermal cells. This dissimilarity is most obvious as neurulation proceeds. The number of cells expressing *AmphiElav* is apparently higher than the number of cells expressing *AmphiCoe* or *AmphiTrk*, which suggests that the embryonic epidermis might contain sub-populations of neuronal cells, which might depend on the combinatorial or differential expression of certain genes, such as *AmphiCoe* or *AmphiTrk*, to abandon their epidermal condition. Thus, the scattered epidermal expression of genes involved in neural specification or differentiation suggests a neurogenic potential for the embryonic epidermis of amphioxus, which would be able to generate neuronal cells from ordinary ectodermal cells, giving rise, among other cells perhaps, to the primary sensory neurons observed by Scanning Electron Microscopy (SEM) in the epidermis of late neurulae [43].

Conversely, other genes show a far more limited axial expression in the epidermis than those mentioned above. In particular, the anterior epidermis appears to concentrate much of the epidermal gene

expression studied to date: *AmphiNeurogenin*, *AmphiPax6*, *Islet*, *AmphiTrk*, *AmphiMsx*, *AmphiOtx*, *AmphiBMP2/4*, *AmphiWnt11*, *AmphiFoxQ2* and *AmphiTob* are all expressed in this region, though in a partially overlapping manner [12,16,20,21,41,51,52-55]. This could correlate with the concurrence of a variety of specialised sensory cell types morphologically characterised in the anterior part of late larval and adult amphioxus specimens [3]. In this respect, the rostral expression of *AmphiMsx* and *AmphiTrk* at early larval stages has been related to the development of the Corpuscles of de Quatrefages, a multi-cellular organ composed of supporting cells and peripheral neurons that remain enclosed within a subepidermal capsule of connective tissue in the rostrum [21,51]. At least some of the peripheral neurons of the Corpuscles of de Quatrefages are primary neurons that send axonal processes to the central nervous system *via* the rostral nerves, but they are morphologically different from those primary neurons observed by SEM at late neurula stages. The latter probably represent the early steps of differentiation of the Type-I receptors described in late larval and adult amphioxus. The most common receptor cell types are called Type I and Type II receptors. While Type I receptors are primary sensory neurons, Type II receptors are secondary sensory neurons devoid of axons. Both types of sensory neurons are widely distributed over the entire epidermis but appear densely grouped in the rostrum, buccal cirri and tail [3]. The oral region of late larvae is also rich in other neuronal cell types exclusively located in this area, such as the oral spines and those of the synaptic complex of the oral nerve [56]. Although the origins of all these peripheral neurons are not clear, there is no evidence indicating anything other than a local origin. Therefore, the assorted neural-related gene expression in these areas could be associated with distinct steps of neural commitment and differentiation of concrete ectodermal cells.

The overlapping expression area of both *AmphiNeurogenin* and *AmphiPax6* in the ectoderm has been homologised with the vertebrate olfactory epithelia, which expresses the respective orthologous genes [12,20]. The vertebrate *Pax-6* gene is the most specific maker of the olfactory epithelia, but is also expressed by the lens placode. *Msx1* and *Msx2* are also expressed in a few other placodes. Conversely, *Sox2*, *Sox3*, *Neurogenin1* and *Neurogenin2*, *Delta-1* and *MyT-1* (*NeuroD*) are expressed in most of the vertebrate placodes [57]. Few specific placodal genes have been studied to date in amphioxus. Among these, except for *AmphiSox1/2/3*, both *AmphiPax6* and *AmphiNeurogenin* are broadly expressed in the rostral ectoderm of early larvae, where *AmphiMsx* is also expressed, yet covering a narrower area on each side of the larvae. Since amphioxus is devoid of placodes, it is very tempting to correlate these gene expression data with the presence of a putative rudimentary placode-like region, which could be defined as a relatively broad ectodermal area of the rostrum able to give rise to distinct sensorial organs and neuronal subpopulations,

and located only in the anterior part of the embryonic ectoderm.

Apart from the scattered expression of certain neural genes (*AmphiTrk*, *AmphiCoe* and *AmphiElav*), the embryonic epidermis of amphioxus expresses a few other genes almost ubiquitously. This is the case of *AmphiEvxB*, *AmphiDral*, *AmphiDil* and *AP-2*, which are widely expressed in the non-neural ectoderm throughout neurulation [13,32,58,59]. These unexpected expression patterns make interpretation of the function of these genes in amphioxus embryogenesis very difficult. Furthermore, *AmphiEvxB* is a highly derived gene, fruit of a specific cephalochordate duplication and a general, cephalochordate-specific role, in epidermal functions has been suggested for this gene [58]. However, the overall epidermal expression of the genes characterised to date in amphioxus might lead to an alternative hypothesis. If the embryonic epidermis of amphioxus is endowed with the neurogenic potential of generating assorted neuronal cell types, *AmphiDil* and *AP-2* could participate in the epidermal *versus* non-epidermal sorting of the different cells in the embryonic skin of amphioxus. Given that both genes are homogeneously expressed in this tissue, the decision between epidermal and neural would not rely on the expression of these genes, but on their expression together with other genes such as *AmphiElav* or other uncharacterised proneural genes.

6. Caveats on putative neural crest precursors

The neural crest is a vertebrate-specific embryonic cell population defined by its origins at the neural plate border, migratory capacity, pluripotency and characteristic gene expression profile. Although the genetic machinery of neural crest appears to be at least partially conserved in both urochordates and cephalochordates [60], there is no compelling evidence, to date, for any invertebrate embryonic cell population that has all the properties of the vertebrate neural crest. Even so, some migratory populations have been described in both urochordate and cephalochordate embryos. In both cases, the migratory cells were identified through Dil cell tracing *in vivo*. While these cells express *HNK-1* and *Zic* gene markers in the ascidian *Ectainascidia turbinata*, no gene expression profile could be determined for the migratory cells in amphioxus [51,61]. Nevertheless, as noted in the previous section, amphioxus gene expression data demonstrate that particular epidermal cells have a distinctive gene expression profile, which would match the distinctive behaviour of these cells that migrate in a ventro-dorsal direction during neurulation. Although this is only data correlation, the genetic program of these individually migrating cells is likely to be different from the surrounding epidermal cells lacking these properties.

Recent studies on BrdU incorporation throughout amphioxus development demonstrate that the embryonic epidermis of the neurulae is devoid of cell proliferation [62]. These results, together with the absence of apoptosis at the same stages and in tissue

[63], indicate that the migrated epidermal cells in late neurulae did not relocate by movements caused by the elimination or addition of nearby cells. The absence of epidermal mitosis is consistent with the ciliation of most cells in the embryonic skin of amphioxus neurula. As mitosis and ciliation seem to be mutually excluding in most eukaryotic cells, proliferation and differentiation also seem to be incompatible [62]. This is consistent with some epidermal cells being able to differentiate during neurulation, when mitosis arrests occur. As in vertebrates, *AmphiDRAL* may be involved in the cell division shutdown, since its downregulation in the epidermis only happens when proliferation is resumed in this tissue, at advanced larval stages [13,61]. Under these circumstances, some cells might be able to activate certain differentiation genetic programs that would lead into the early differentiation of the primary neurons observed at late neurula stages [43]. However, such a spatio-temporal genetic program is currently unknown, though it is possible to establish certain associations between gene expression and epidermal subpopulations.

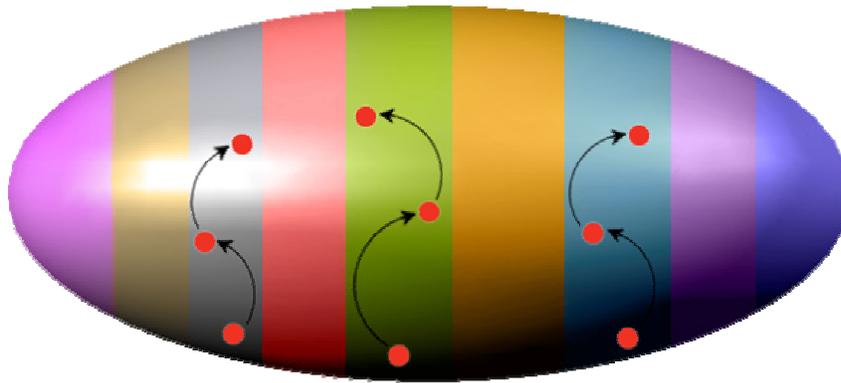
At present, it seems clear that distinct genetic programs are operating in the embryonic skin, distinguishing individual and possibly migrating cells from the rest of the epidermis. As previously mentioned, the most prominent epidermal subpopulations are those labelled by *AmphiCoe*, *AmphiElav* and *AmphiTrk* expression. These are followed by the much smaller subpopulations expressing *islet* and *AmphiERR*, which, unlike those expressing *AmphiCoe*, *AmphiElav* and *AmphiTrk*, appear only in the trunk region [4, 49]. As a common denominator, all these genes are involved in the specification of neuronal precursors or in neuronal differentiation, thereby strengthening the local neurogenic potential of the amphioxus embryonic epidermis. In tandem with this, one of the most striking features of *AmphiTrk* and *AmphiElav* expression is the dorsalisation of the signal as neurulation proceeds, which follows the same direction at the same stages of development of those migrating individual cells observed by DiI tracing [51]. Furthermore, the detailed position of *AmphiTrk*- and *AmphiElav*-positive cells locates them within the epidermis or just beneath it, suggesting that if these cells are migrating they might be doing it either through the epidermis or through the subepidermal plexus. On taking this independent evidence together with the expression of the vertebrate *Trk* and *Hu/Elav* orthologue genes in the neural crest and/or its derivatives, it is very tempting to establish parallelism between these cells and the vertebrate neural crest. Therefore, if these cells (*AmphiTrk*- and *AmphiElav*-positive cells) individually detached from the epidermal layer and entered the sub-epidermal plexus, until reaching the definitive (and unknown) location at which they would finally differentiate into particular neuronal types, they would conform some of the properties of vertebrate neural crest cells.

The particular migration of vertebrate neural crest cells is also characterised by the precise definition of their individual routes, which are crucially dependent on an antero-posterior Hox code. Interestingly, nested Hox expression in scattered epidermal cells has been observed in the embryonic epidermis of amphioxus [4] by the time that migration begins and the earliest neural gene expression is already occurring. Furthermore, under retinoic acid treatment the location of those individual epidermal cells that express Hox genes is altered, as is that of the *AmphiCoe*, *islet* and *AmphiERR*-positive cells and, importantly, that of the primary neurons observed at late neurula stages [4]. These results suggest that the embryonic skin of amphioxus is probably pre-patterned following an antero-posterior Hox code, which in conjunction with other genes would divide the epidermis into distinct territories with particular gene expression profiles. Under this hypothesis, individual cells would migrate vertically through defined routes, which would molecularly, but not morphologically, be delimited into epidermal meridians (Fig. 2). Another particularity of vertebrate neural crest cells is their pluripotency. As mentioned previously, amphioxus skin possesses a moderate diversity of peripheral sensory neurons, probably born locally from normal epidermal cells through different expression profiles [50]. Collation of all the data set out here suggests that the individually migrating cells in amphioxus epidermis may well match cells with a particular gene expression profile that are responsible for the final differentiation into distinct types of peripheral sensory neurons, though these have much less pluripotency than in vertebrates.

In summary, based on: i) the presence of distinct genetic programs in individual embryonic epidermal cells; ii) the ability to migrate as individual cells; iii) the ability to detach, presumably before migration; iv) correlation with the antero-posterior boundaries of Hox expression domains; and v) the partial pluripotency of these cells, I suggest that amphioxus possesses an embryonic cell population that exhibits most of the properties of vertebrate neural crest cells. Thus, it is tempting to speculate that they might represent an intermediate evolutionary step that ended up in the deployment of vertebrate neural crest cells. The main caveats of this thesis are that these cells do not originate from the edges of the neural tube and that migration has the opposite orientation (ventro-dorsal). However these two caveats can be reconciled if, in an evolutionary sense, the amphioxus nerve neural system represents an intermediate state, from a diffuse neural epidermal net all around the body, like that of hemichordates [64], to full concentration on the dorsal side of the body, where the nerve tube cohabits in vertebrates with the embryonic germ of the neural crest and placodes.

Figure 2. Hypothetical model for neural crest-like cell migration in amphioxus. The expression of characteristic neural genes in scattered individual cells in the epidermis indicates that neuronal precursors are generated from the

normal epidermal cells following a particular genetic program switched on during neurulation. These unknown genetic programs should endow these particular cells with additional distinctive properties, which might include the acquisition of individual migratory behaviour. The migration and terminal differentiation of those neuronal precursors, in particular locations, could depend on the pre-patterning of the neurula epidermis. This model summarises and gives an oversimplified view of the epidermal expression of different genes that seem to participate in the epidermal patterning of the amphioxus neurula. As in the neural tube, *Hox* genes are expressed in a nested manner and, together with the expression of *AmphiPax6*, *AmphiEn* and *AmphiCdx*, divide the entire epidermis into meridians of differential expression. All these genes are expressed in a scattered manner, but are delimited along the anteroposterior axis as indicated by colour code. In the case of the *Hox* genes, the meridians were established by the combinatorial expression of *AmphiHox* 1, 3, 4 and 6. Under this hypothetical model, those individual cells (red dots) mentioned above, would migrate vertically (arrows) through defined routes, as delimited by each meridian of combinatorial expression.



7. Conclusion

Accumulating data on gene expression during amphioxus embryogenesis challenges the “classic” morphological simplicity of the amphioxus nervous system. Segmentally-expressed, iteratively-expressed, “spottily”-expressed genes, neural and non-neural ectodermal specificity, cell migration in the ectoderm and cryptic complexity are all providing us with new insights into the neural genetic programs of this, our old beast, half-way along the path of construction of complex vertebrate nervous systems. In the near post-genomic future, more and more data will be arriving and, hopefully, experimental embryology (by means of genetic manipulation of living embryos) will help

clarify the numerous question marks on the evolution of the nervous system, for which the friendly little amphioxus may have some answers.

Acknowledgements

The author wishes to thank an anonymous referee for very helpful comments and Robin Rycroft for checking the English. The author's research was funded by grant BFU2005-00252 (Spanish Ministry of Education and Science) and by the European Community's Human Potential Programme HPRN-CT-2002-00263/BMC 'Neurogenome'.

Conflict of interest

The author declares that no conflict of interest exists.

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Tables

Table 1. Summary of gene expression in the nervous system and non-neural ectoderm

Gene	Neural ectoderm expression	Non-neural ectoderm expression	Orientative functions in vertebrates	References
AmHNF-3-1	Ventral midline of the neural tube (Floor plate)	Negative	Neural induction and notochord specification (organiser functions)	[17]
AmHNF-3-2	Negative	Negative		
AmphiBMP2/4	Negative	Anterodorsal patch of rostral epidermis	Dorsoventral axis patterning	[52]
AmphiBrn1/2/4	Neural plate (excluding ventral midline) and cerebral vesicle	Negative	Neurogenesis	[25]
AmphiCdx	Posterior neural tube	Posterior ectoderm	Axial patterning	[65]
AmphiCoe	Bilateral series of dorsal and ventral cells within the neural tube and posterior cerebral vesicle	Ventrolateral scattered individual epidermal cells	Neuronal differentiation	[43]
AmphiD1	Particular regions of the cerebral vesicle	Negative	Neurotransmission	[33]
AmphiDach	Particular regions of the cerebral vesicle and scattered cells within the neural tube	Negative	Central nervous system development	[48]
AmphiDll	Anterior neural plate borders, roof plate and ventrolateral cells in the vicinity of the pigment spot	Spread all over the epidermis	Eye and limb development Ectoderm regionalisation, dorsoventral axis establishment and forebrain specification	[32]

Gene	Neural ectoderm expression	Non-neural ectoderm expression	Orientative functions in vertebrates	References
AmphiDRAL	Negative	Homogeneously expressed all over the epidermis	Differentiation	[13]
AmphiElav	Bilateral series of dorsal and ventral cells within the neural tube and posterior the cerebral vesicle	Ventrolateral scattered individual epidermal cells	Neurogenesis and neuronal differentiation	[42] [44]
AmphiEn	Posterior cerebral vesicle and two clusters of cells within the neural tube at the trunk level	Ventrolateral band of scattered epidermal cells at the level of the second somite	Rostrocaudal segmentation and neurogenesis Specification and maintenance of boundaries	[66]
AmphiERR	Bilateral series of ventral cells within the neural tube and the presumptive frontal eye	Ventrolateral scattered individual epidermal cells at the trunk level	Neuronal differentiation	[49]
AmphiEvxA	Two bilateral clusters of cells within the neural tube at the trunk level	Posterior ectoderm	Gastrulation, neurogenesis, appendage development and tailbud formation	[23] [58]
AmphiEvxB	Negative	Homogeneously expressed all over the epidermis		
AmphiFoxB	Iterative series of cells in the neural plate and two patches of cells within the cerebral vesicle	Negative	Neural segmentation	[34]
AmphiFoxD	Anterior neural plate and ventral cerebral vesicle	Negative	Neural crest development	[19]
AmphiFoxQ2	Negative	Rostral ectoderm		[54]
AmphiFringe	Iterative series of cells in the anterior neural plate and in the cerebral vesicle	Negative	Specification and maintenance of boundaries	[47]
AmphiF-spondin	Cerebral vesicle and neural tube (most of the central nervous system)	Negative	Neural specific extracellular matrix	[37]
AmphiGli	Neural plate	Unknown	Motor neuron induction	[24]
AmphiGsx	Cerebral vesicle	Negative	Axial patterning	[65]
AmphiHairyA	Posterior neural tube	Negative	Neurogenesis and somitogenesis	[27]
AmphiHairyB	Bilateral gapped expression in the neural tube	Negative		
AmphiHairyC	Bilateral gapped expression in the neural tube	Negative		
AmphiHairyD	Bilateral gapped expression in the neural tube	Negative		
AmphiHh	Ventral midline of the neural tube (Floor plate)	Negative	Axial patterning	[38]
AmphiHox1	Segmented expression in the neural tube (S2-S4)	Ventrolateral scattered individual epidermal cells at the trunk level	Axial patterning	[4] [35]
AmphiHox3	Segmented expression in the neural tube (S4-S8)	Ventrolateral scattered individual epidermal cells at the trunk-caudal level		
AmphiHox4	Segmented expression in the neural tube (S6-S8)	Ventrolateral scattered individual epidermal cells at the caudal level		
AmphiKrox	Bilateral iterative series of cells in the neural plate and cerebral vesicle	Negative	Specification and maintenance of segmental identity	[39] [41]
AmphiMnx	Bilateral iterative series of cells in the neural plate	Negative	Motor neuron differentiation	[40]
AmphiMsx	Dorsolateral series of cells in the neural tube	Bilateral patches of rostral epidermal cells	Nervous system development Neural crest and placodes development	[21]
AmphiNetrin	Ventral midline of the neural tube (Floor plate)	Negative	Axonal path finding	[36]
AmphiNeurogenin	Neural plate (excluding ventral midline) and dorsal neural tube later in development	Scattered cells of the rostral epidermis	Neural specification and differentiation	[12]
AmphiNk2-1	Ventral midline of neural tube (Floor plate) and ventral cerebral vesicle	Negative	Development of the central nervous system and thyroid gland	[46]
AmphiNk2-2	Ventrolateral cells within the cerebral vesicle	Negative	Nervous system regionalisation	[45]
AmphiNotch	Posterior and anterior neural plate Dorsal cells of the cerebral vesicle	Negative	Cell fate decision in the neuroectoderm and formation of boundaries	[10]
AmphiOtx	Cerebral vesicle	Scattered cells of the rostradorsal epidermis	Anterior head formation	[16]
AmphiPax2/5/8	Neural tube and anterodorsal cerebral vesicle	Negative	Midbrain-hindbrain boundary formation	[67]
AmphiPax3/7	Anterior neural plate and dorsal third of cerebral vesicle	Dorsal ectoderm	Dorsoventral regionalisation of the central nervous system, neural crest and placodes development, and	[26]

Gene	Neural ectoderm expression	Non-neural ectoderm expression	Orientative functions in vertebrates	References
AmphiPax6	Anterior neural tube and cerebral vesicle	Anterior epidermis	myogenesis Eye development and cell type specification	[20]
AmphiSim	Posterior cerebral vesicle	Negative	Neural segmentation	[34]
AmphiSox1/2/3	Dorsal neural tube	Negative	Neural specification and differentiation	[12]
AmphiTH	Particular regions of the cerebral vesicle	Negative	Neurotransmission	[33]
AmphiTob	Scattered cells along the neural tube and cerebral vesicle	Anterior epidermis	Proliferation arrest	[55]
AmphiTrk	Ventral and dorsal neural tube (adults)	Ventrolateral scattered individual epidermal cells	Neuronal survival and differentiation	[51]
AmphiVent	Neural plate borders	Negative	Mesoderm patterning	[14]
AmphiWnt11	Presumptive neuroectoderm	Dorsal, anteroventral and posterior ectoderm	Axial patterning	[53]
AmphiWnt3	Neural plate borders and cerebral vesicle	Posterior ectoderm		[9]
AmphiWnt4	Neural tube and posterior cerebral vesicle	Negative		[68]
AmphiWnt5	Ventral cerebral vesicle	Negative		[9]
AmphiWnt6	Neural plate borders and posterior neural tube	Negative		
AmphiWnt7b	Neural tube	Negative		[68]
AmphiWnt8	Patch of cells within the ventral cerebral vesicle	Negative		
AmphiXlox	Transient cluster of cells in the neural tube at the level of somite 5 (pigment spot)	Negative	Axial patterning	[65]
AmphiZic	Neural tube and cerebral vesicle	Negative	Neural crest induction	[24]
AP-2	Cerebral vesicle	Spread all over the epidermis	Neural crest development	[59]
Bblhx3	Bilateral iterative series of cells within the neural tube	Negative	Axial patterning and neuronal differentiation	[70]
BF-1	Anteroventral cerebral vesicle	Negative	Telencephalon development	[31]
Id	Anterior neuroectoderm	Negative	Neural crest development	[28]
Islet	Bilateral iterative series of cells within the neural tube	Ventrolateral band of scattered epidermal cells at the trunk level and rostral epidermis	Neuronal patterning and differentiation	[4] [22] [41]
rab GDI	Anterior neural tube and cerebral vesicle	Negative	Regulation of vesicle-mediated cellular transport	[71]
Shox	Bilateral iterative series of cells within the neural tube	Negative	Growth control and neural specification	[41]
Snail	Ventrolateral neural tube	Dorsal ectoderm	Neural crest development	[30]