

**Supplement Figure 1. *uPA attenuated inflammatory osteoclastogenesis through***

***plasmin/PAR-1.***

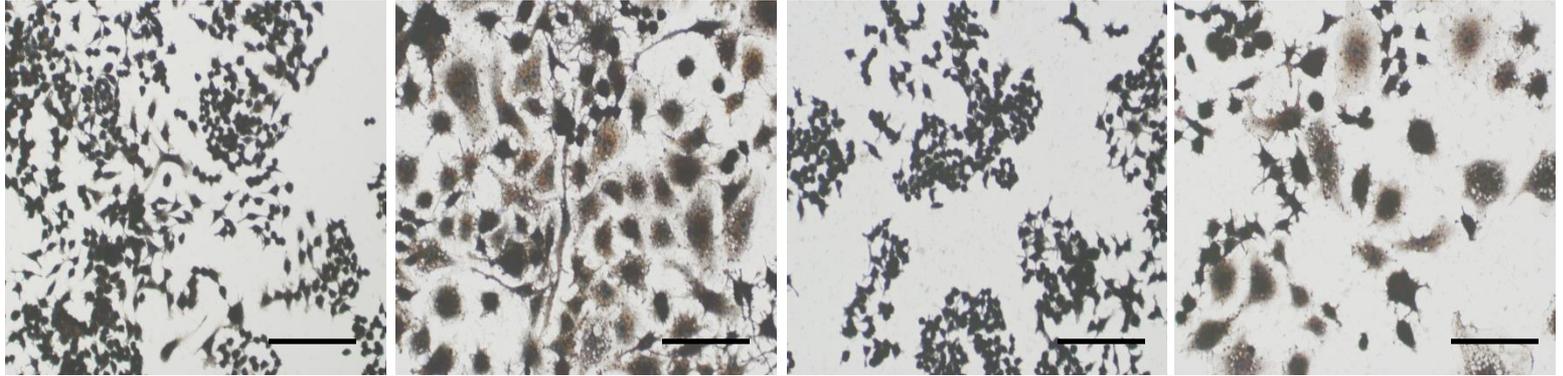
(A) RAW264.7 cells were cultured for 3 days in the absence or presence of LPS (1  $\mu\text{g/ml}$ ) or uPA (10 nM) as indicated. Then, TRAP-staining was performed to detect mature OCs. (B) Mature OCs were identified as multinucleated TRAP-positive cells (n=4). (C) The expression of NFATc1 in RAW264.7 cells was evaluated by Western blot analysis. The histogram on the bottom panel shows quantitative representations of NFATc1 obtained from densitometry analysis after normalization to the levels of GAPDH expression (n=3). (D) Bone marrow-derived cells from the wild-type mice were cultured for 3 days in the absence or presence of LPS (1  $\mu\text{g/ml}$ ), M-CSF (100 ng/ml) or uPA (10 nM). Mature OCs were identified as multinucleated TRAP-positive cells (n=3). (E) RAW264.7 cells were cultured for 3 days in the absence or presence of LPS (1  $\mu\text{g/ml}$ ), uPA (10 nM),  $\alpha$ 2AP (2 nM), or plasmin (1  $\mu\text{g/ml}$ ) as indicated. Then, TRAP-staining was performed to detect mature OCs. (F) Mature OCs were identified as multinucleated TRAP-positive cells (n=5). (G) The expression of NFATc1 in RAW264.7 cells was evaluated by Western blot analysis. The histogram on the bottom panel shows quantitative representations of NFATc1 obtained from densitometry analysis after normalization to the levels of GAPDH expression (n=3). (H) Bone marrow-derived cells from the wild-type mice were cultured for 3 days in the absence or presence of LPS (1  $\mu\text{g/ml}$ ), M-CSF (100 ng/ml), uPA (10 nM),  $\alpha$ 2AP (2 nM), or plasmin (1  $\mu\text{g/ml}$ ) as indicated. Mature OCs were identified as multinucleated TRAP-positive cells (n=3). (I) RAW264.7 cells were cultured for 3 days in the absence or presence of LPS (1  $\mu\text{g/ml}$ ), uPA (10 nM), or PAR-1 antagonist SCH 79797 (1 ng/ml) as indicated. Then, TRAP-staining was performed to detect mature OCs. (J) Mature OCs were identified as multinucleated TRAP-positive cells (n=4). (K) The expression of NFATc1 in RAW264.7 cells was evaluated by Western blot analysis. The histogram on the bottom panel shows quantitative representations of NFATc1 obtained from densitometry analysis after normalization to the levels of GAPDH expression (n=3). (L) Bone marrow-derived cells from the wild-type mice were cultured for 3 days in the absence or presence of LPS (1  $\mu\text{g/ml}$ ), M-CSF (100 ng/ml), uPA (10 nM), or PAR-1 antagonist SCH 79797 (1 ng/ml) as indicated. Mature OCs were identified as multinucleated TRAP-positive cells (n=3). (M) RAW264.7 cells were cultured for 3 days in the absence or presence of LPS (1  $\mu\text{g/ml}$ ) or PAR-1 agonist TFLLRN-NH<sub>2</sub> (1  $\mu\text{g/ml}$ ) as indicated. Then, TRAP-staining was performed to detect mature OCs. (N) Mature OCs were identified as multinucleated TRAP-positive cells (n=5).

(O) The expression of NFATc1 in RAW264.7 cells was evaluated by Western blot analysis. The histogram on the bottom panel shows quantitative representations of NFATc1 obtained from densitometry analysis after normalization to the levels of GAPDH expression (n=3). (P) Bone marrow-derived cells from the wild-type mice were cultured for 3 days in the absence or presence of LPS (1  $\mu$ g/ml), M-CSF (100 ng/ml), or PAR-1 agonist TFLLRN-NH<sub>2</sub> (1  $\mu$ g/ml) as indicated. Mature OCs were identified as multinucleated TRAP-positive cells (n=3). The data represent the mean  $\pm$  SEM. \*,  $P < 0.01$ ; \*\*,  $P < 0.05$ ; NS, not significant. Scale bar = 200  $\mu$ m.

**Supplement Figure 2. *The AMPK pathway activated by uPA attenuated inflammatory osteoclastogenesis.***

(A) RAW264.7 cells were cultured for 3 days in the absence or presence of LPS (1  $\mu$ g/ml) or AICAR (100  $\mu$ M) as indicated. Then, TRAP-staining was performed to detect mature OCs. (B) Mature OCs were identified as multinucleated TRAP-positive cells (n=4). (C) The expression of NFATc1 in RAW264.7 cells was evaluated by Western blot analysis. The histogram on the bottom panel shows quantitative representations of NFATc1 obtained from densitometry analysis after normalization to the levels of GAPDH expression (n=3). (D) Bone marrow-derived cells from the wild-type mice were cultured for 3 days in the absence or presence of LPS (1  $\mu$ g/ml), M-CSF (100 ng/ml), or AICAR (100  $\mu$ M) as indicated. Mature OCs were identified as multinucleated TRAP-positive cells (n=3). (E) RAW264.7 cells were stimulated with 10 nM uPA for the indicated periods. Phosphorylation of AMPK and AMPK were evaluated by a Western blot analysis. The histogram on the bottom panel shows quantitative representations of phospho-AMPK obtained from densitometry analysis after normalization to the levels of AMPK expression. (F) RAW264.7 cells were cultured for 3 days in the absence or presence of LPS (1  $\mu$ g/ml), uPA (10 nM), or compound C (0.1 or 0.5  $\mu$ M) as indicated. Then, TRAP-staining was performed to detect mature OCs. (G) Mature OCs were identified as multinucleated TRAP-positive cells (n=4). (H) The expression of NFATc1 in RAW264.7 cells was evaluated by Western blot analysis. The histogram on the bottom panel shows quantitative representations of NFATc1 obtained from densitometry analysis after normalization to the levels of GAPDH expression (n=3). (I) Bone marrow-derived cells from the wild-type mice were cultured for 3 days in the absence or presence of LPS (1  $\mu$ g/ml), M-CSF (100 ng/ml), or compound C (0.1  $\mu$ M) as indicated. Mature OCs were identified as multinucleated TRAP-positive cells (n=3). (J) RAW264.7 cells were pretreated with 2 nM  $\alpha$ 2AP for 30 minutes and then stimulated with 10 nM uPA for 60 minutes. Phosphorylation of AMPK and AMPK were

evaluated by a Western blot analysis. The histogram on the bottom panel shows quantitative representations of phospho-AMPK obtained from densitometry analysis after normalization to the levels of AMPK expression. (K) RAW264.7 cells were stimulated with 1  $\mu\text{g}/\text{ml}$  plasmin for the indicated periods. Phosphorylation of AMPK and AMPK were evaluated by a Western blot analysis. The histogram on the bottom panel shows quantitative representations of phospho-AMPK obtained from densitometry analysis after normalization to the levels of AMPK expression. (L) RAW264.7 cells were stimulated with 1  $\mu\text{g}/\text{ml}$  PAR-1 agonist TFLLRN-NH<sub>2</sub> for the indicated periods. Phosphorylation of AMPK and total amount of AMPK were evaluated by a Western blot analysis. The histogram on the bottom panel shows quantitative representations of phospho-AMPK obtained from densitometry analysis after normalization to the levels of AMPK expression. (M) RAW264.7 cells were pretreated with 1  $\mu\text{g}/\text{ml}$  PAR-1 antagonist SCH 79797 for 60 minutes and then stimulated with 10 nM uPA for 60 minutes. Phosphorylation of AMPK and total amount of AMPK were evaluated by a Western blot analysis. The histogram on the bottom panel shows quantitative representations of phospho-AMPK obtained from densitometry analysis after normalization to the levels of AMPK expression. The data represent the mean  $\pm$  SEM. \*,  $P < 0.01$ ; \*\*,  $P < 0.05$ ; NS, not significant. Scale bar = 200  $\mu\text{m}$ .



LPS (1 μg/ml)	-	+	-	+
uPA (10 nM)	-	-	+	+

Fig. S1A

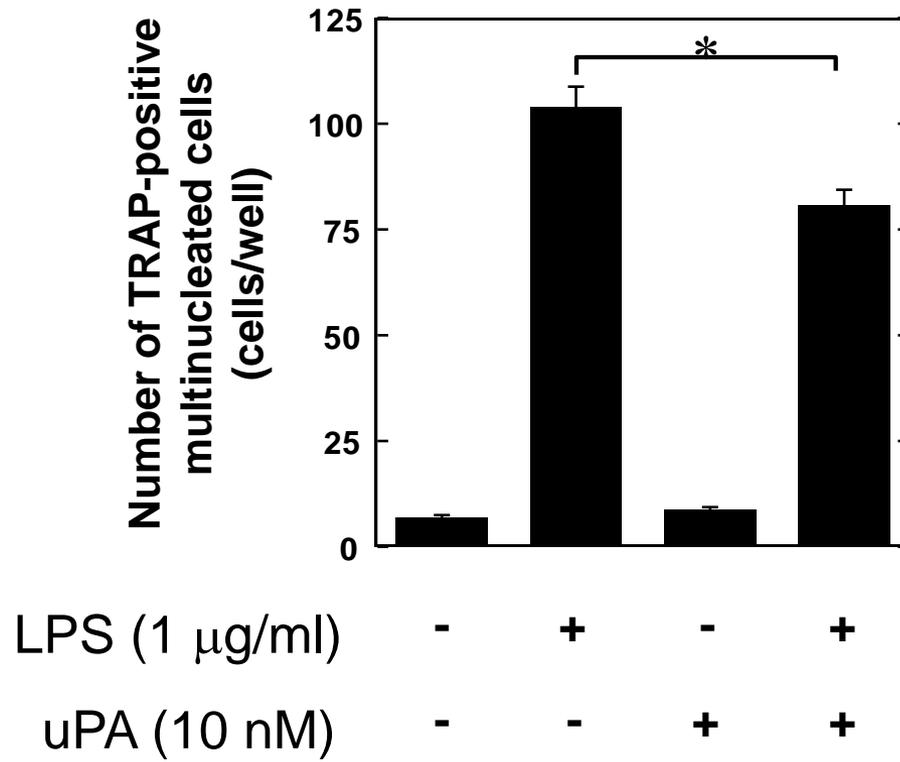


Fig. S1B

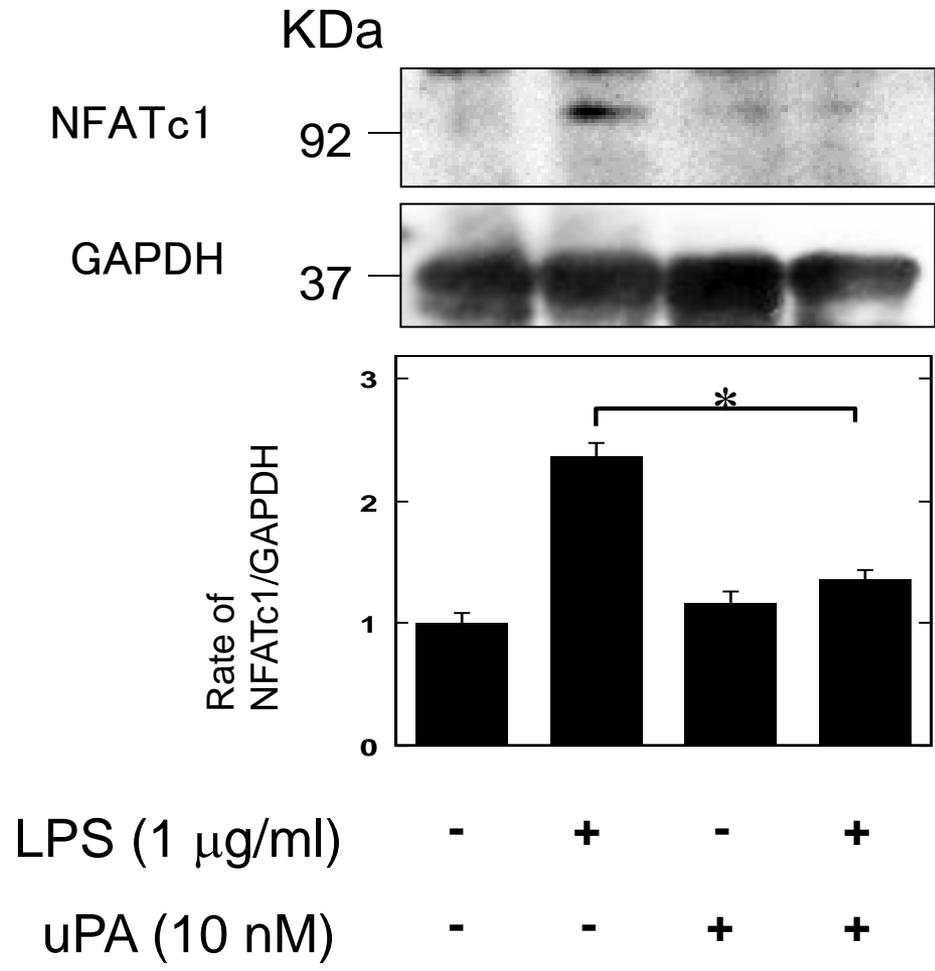


Fig. S1C

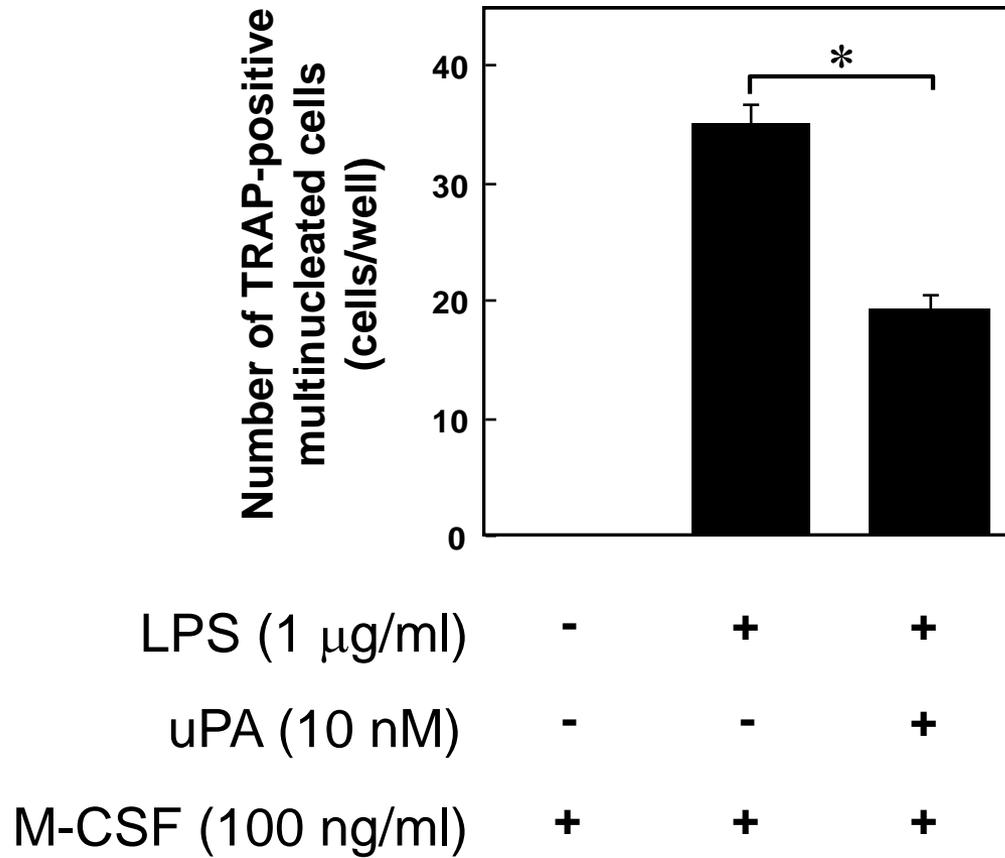
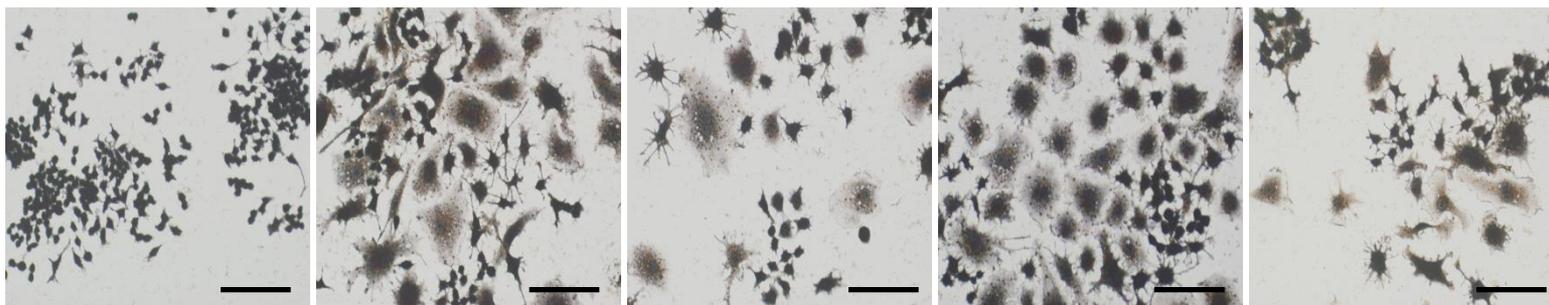


Fig. S1D



LPS (1 $\mu\text{g/ml}$ )	-	+	+	+	+
uPA (10 nM)	-	-	+	+	-
$\alpha 2\text{AP}$ (2 nM)	-	-	-	+	-
Plasmin (1 $\mu\text{g/ml}$ )	-	-	-	-	+

Fig. S1E

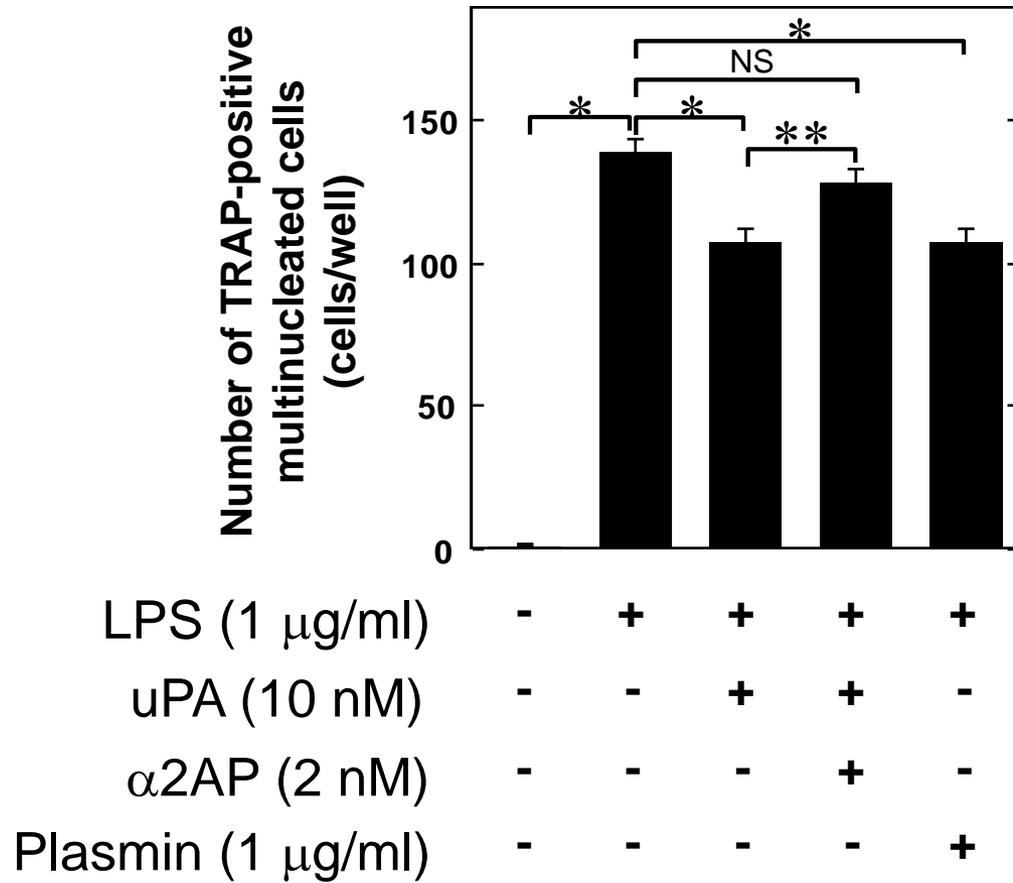


Fig. S1F

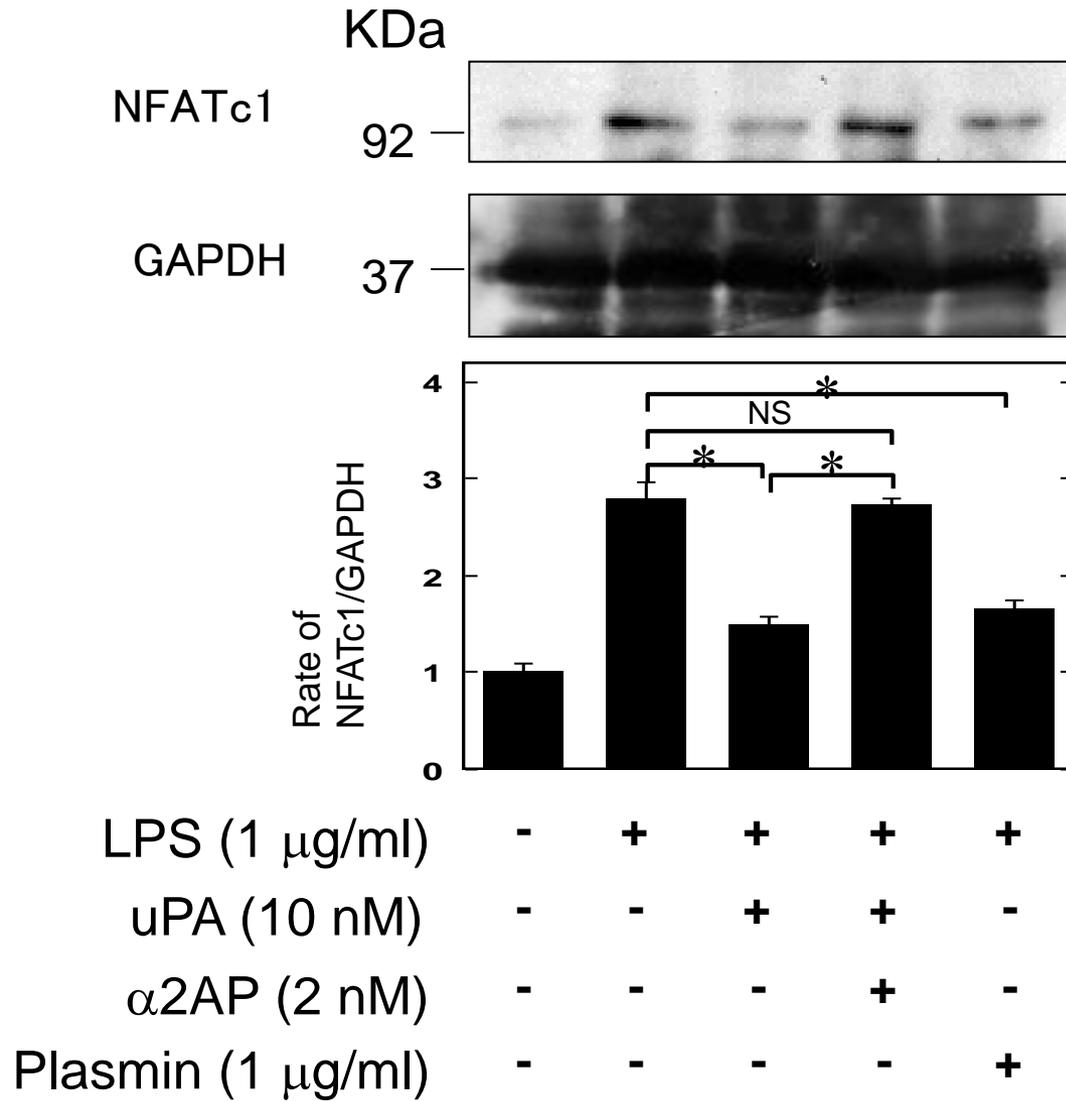
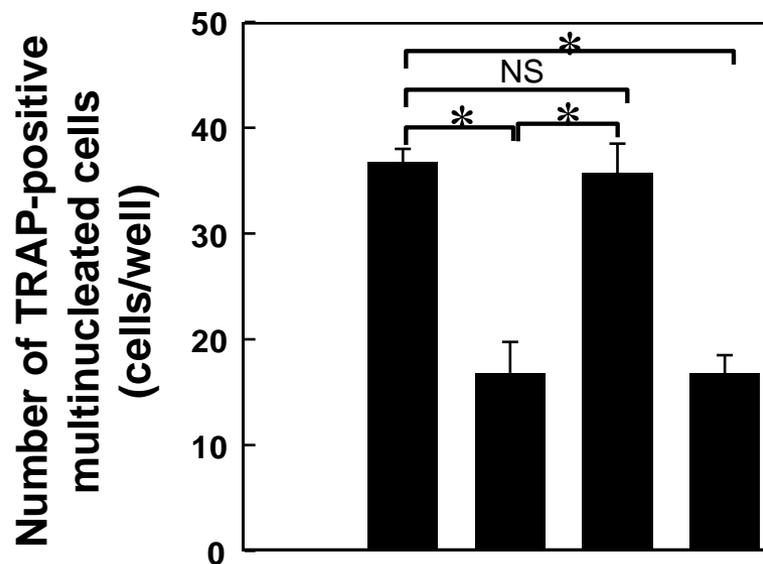
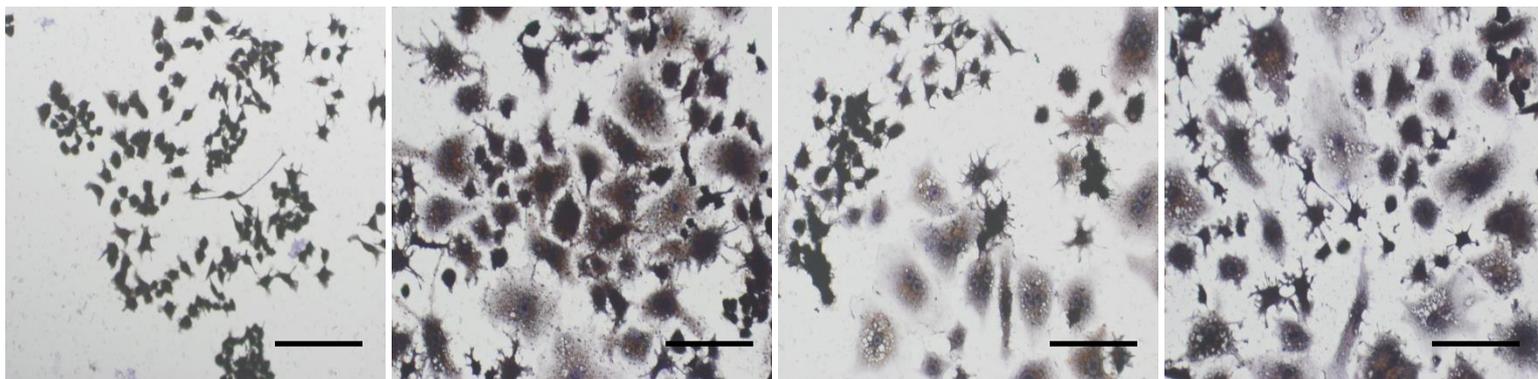


Fig. S1G



LPS (1 $\mu$ g/ml)	-	+	+	+	+
uPA (10 nM)	-	-	+	+	-
$\alpha$ 2AP (2 nM)	-	-	-	+	-
Plasmin (1 $\mu$ g/ml)	-	-	-	-	+
M-CSF (100 ng/ml)	+	+	+	+	+

Fig. S1H



LPS (1  $\mu\text{g/ml}$ ) -

+

+

+

uPA (10 nM) -

-

+

+

SCH 79797 (1 ng/ml) -

-

-

+

Fig. S1I

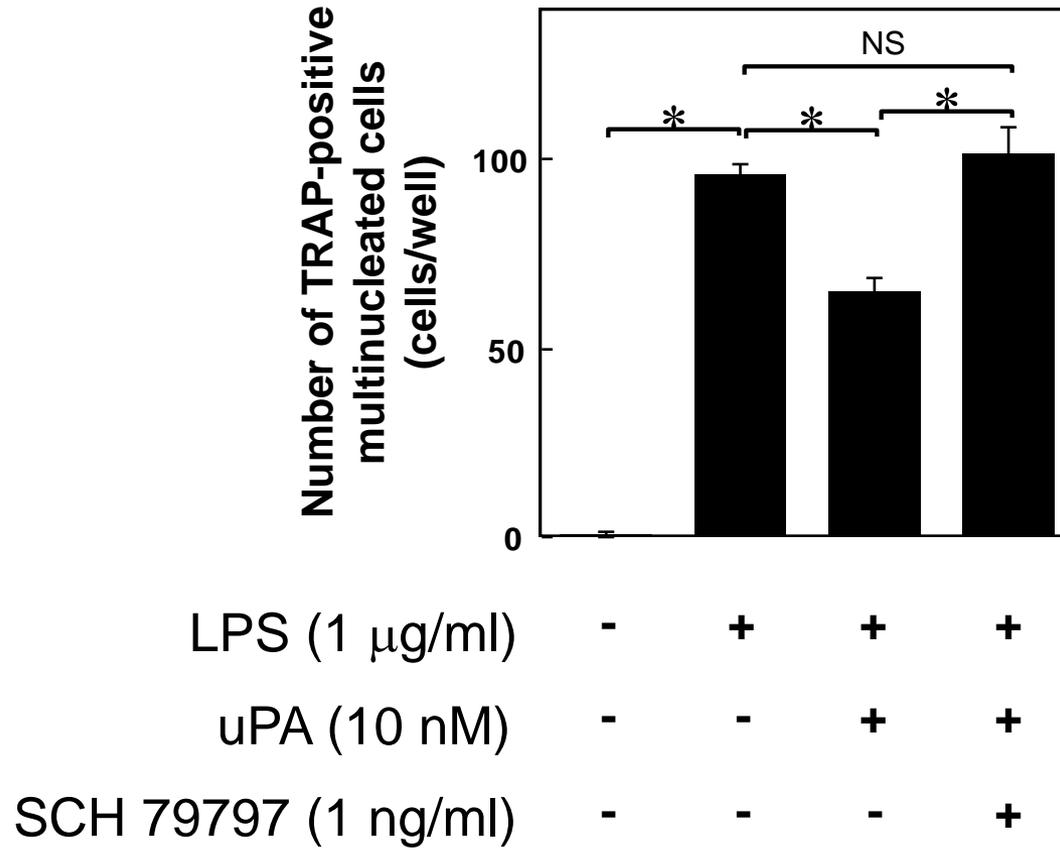


Fig. S1J

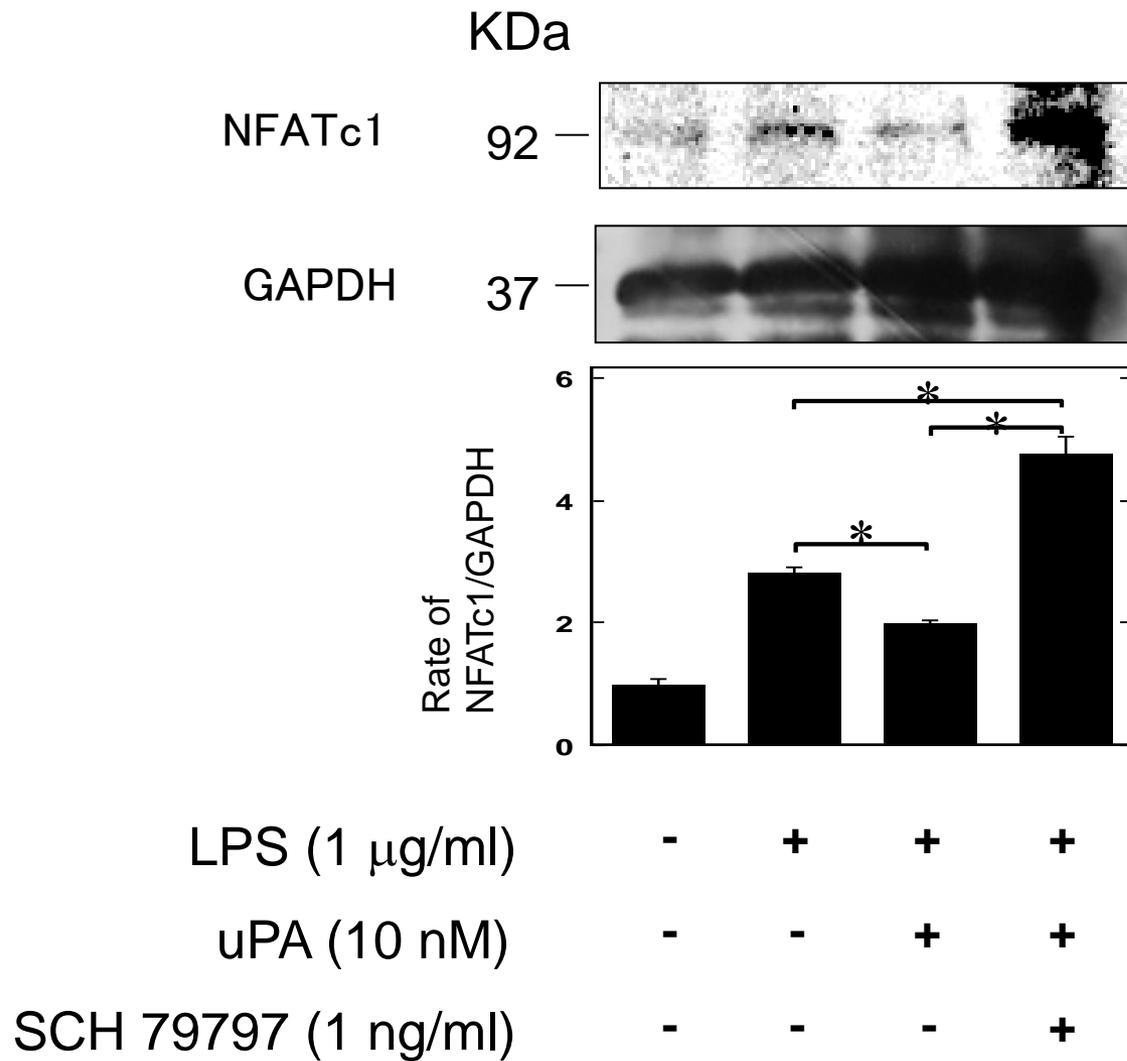


Fig. S1K

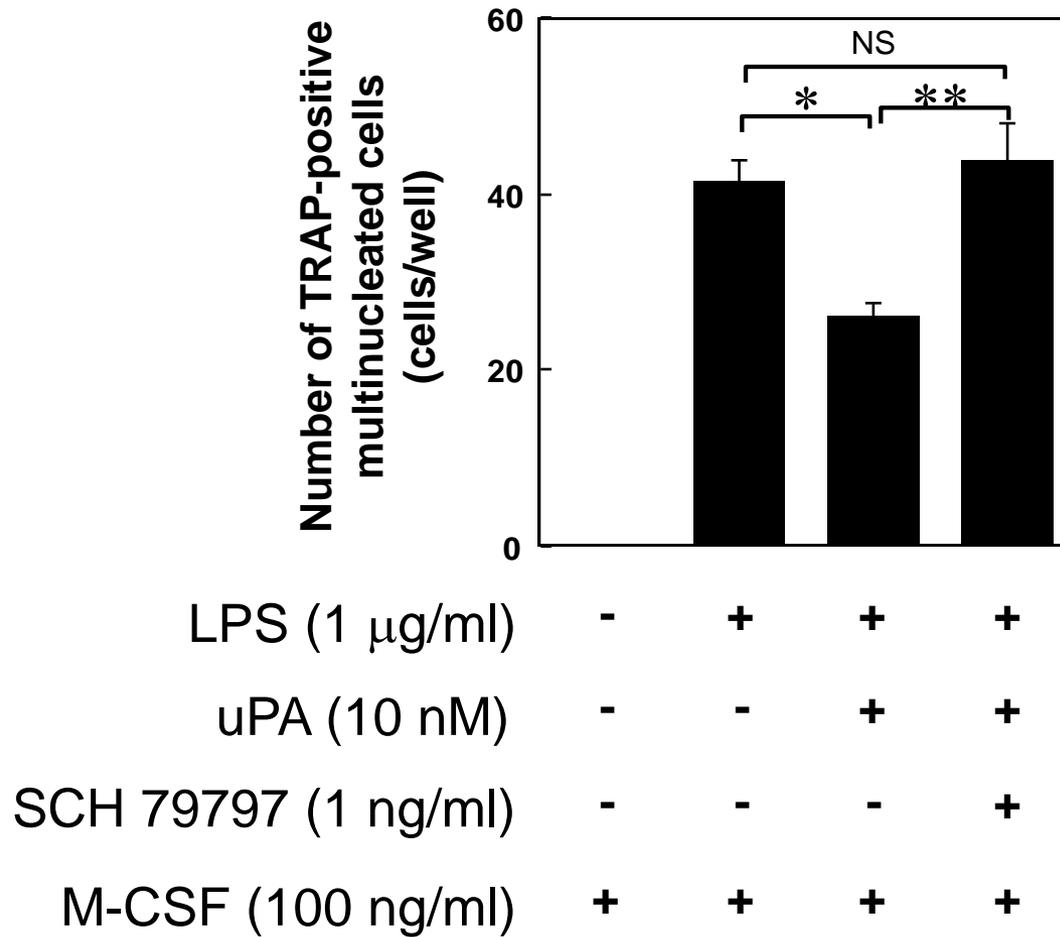
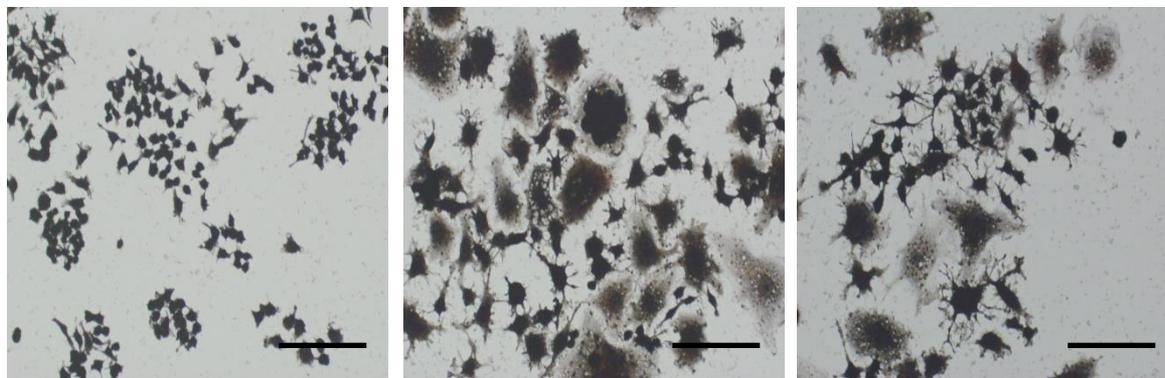


Fig. S1L



LPS (1 $\mu\text{g/ml}$ )	-	+	+
TFLLRN-NH <sub>2</sub> (1 $\mu\text{g/ml}$ )	-	-	+

Fig. S1M

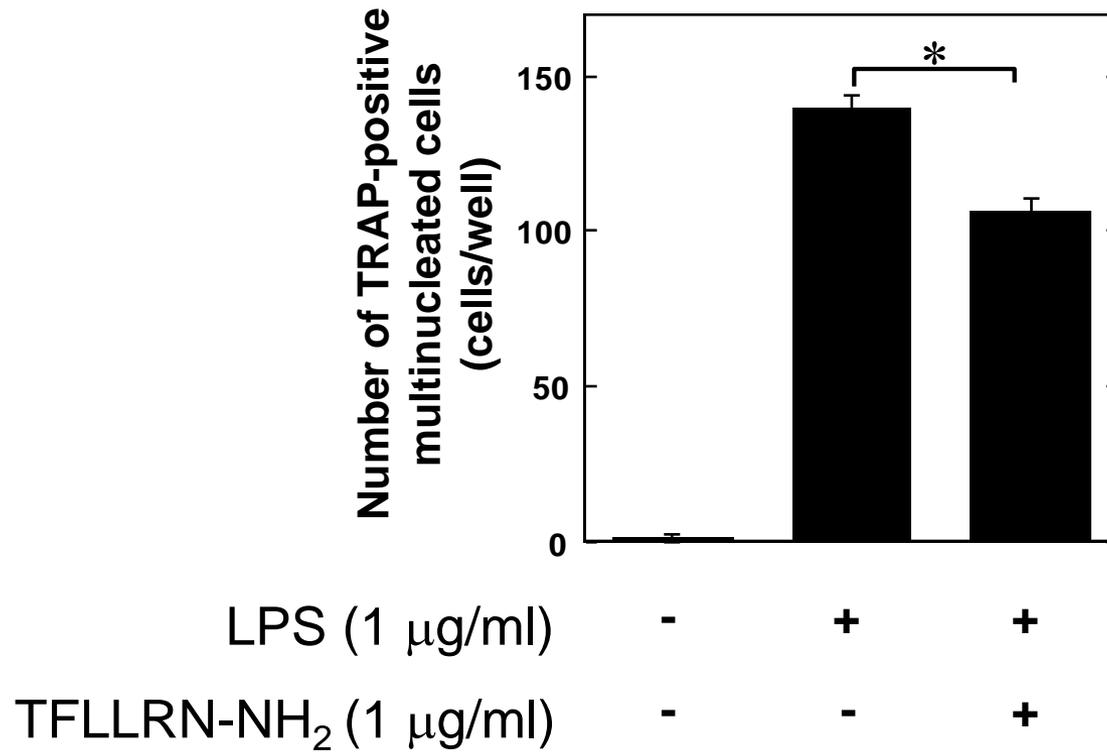


Fig. S1N

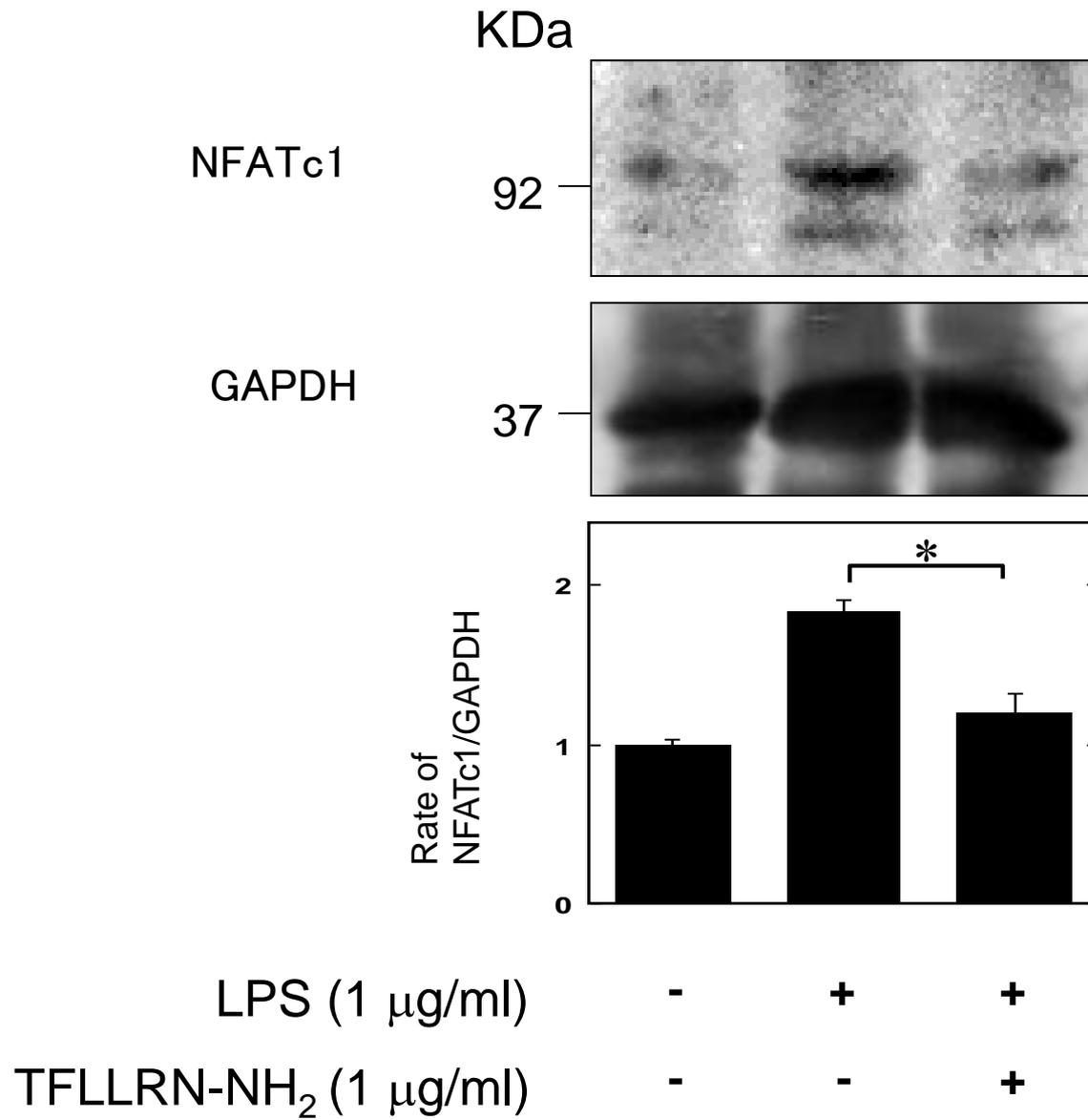


Fig. S10

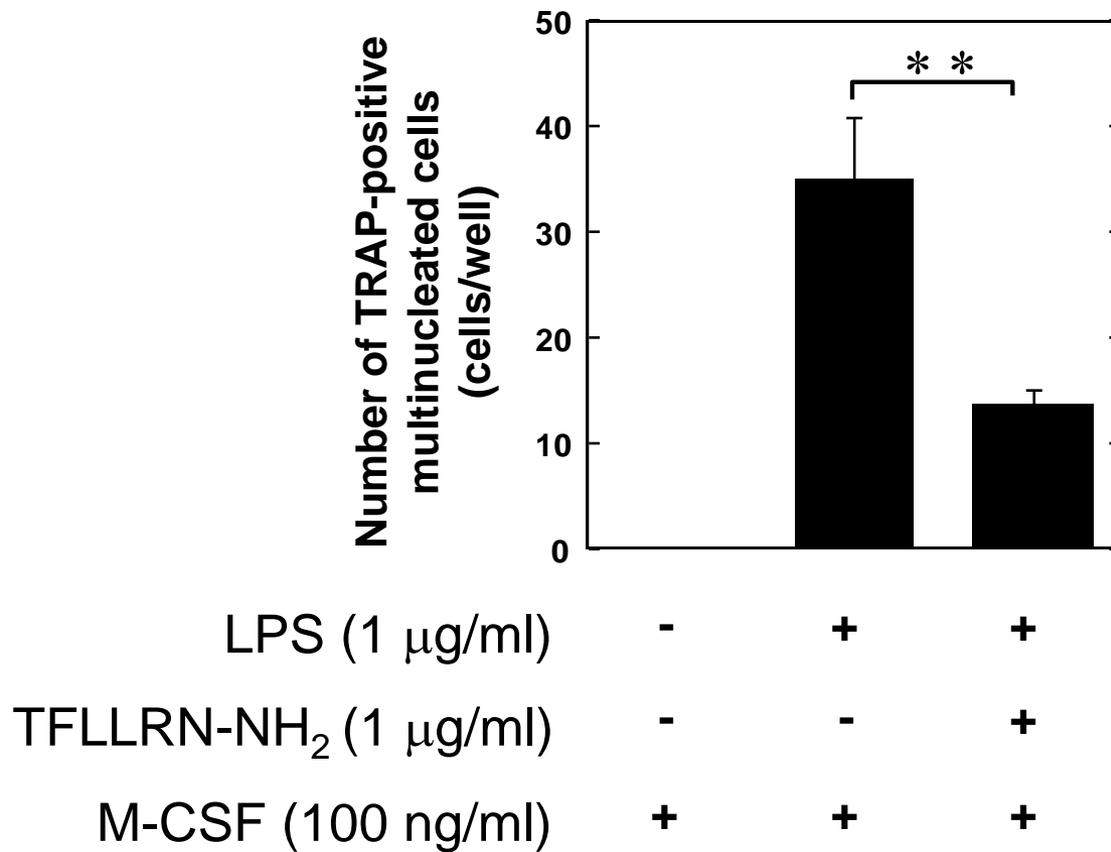
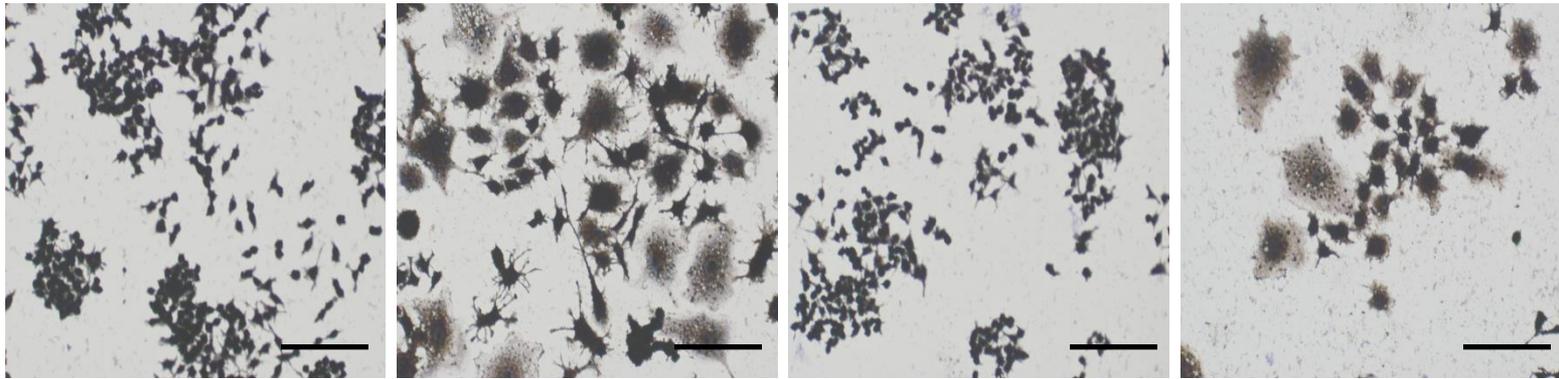


Fig. S1P



LPS (1 μg/ml)	-	+	-	+
AICAR (100 μM)	-	-	+	+

Fig. S2A

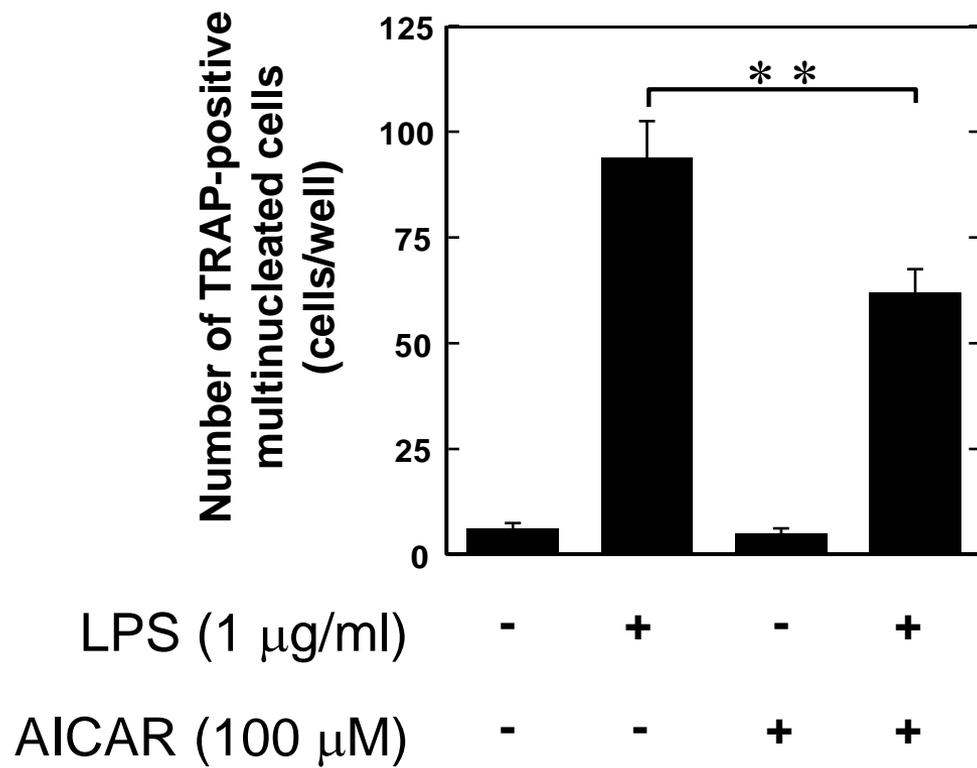


Fig. S2B

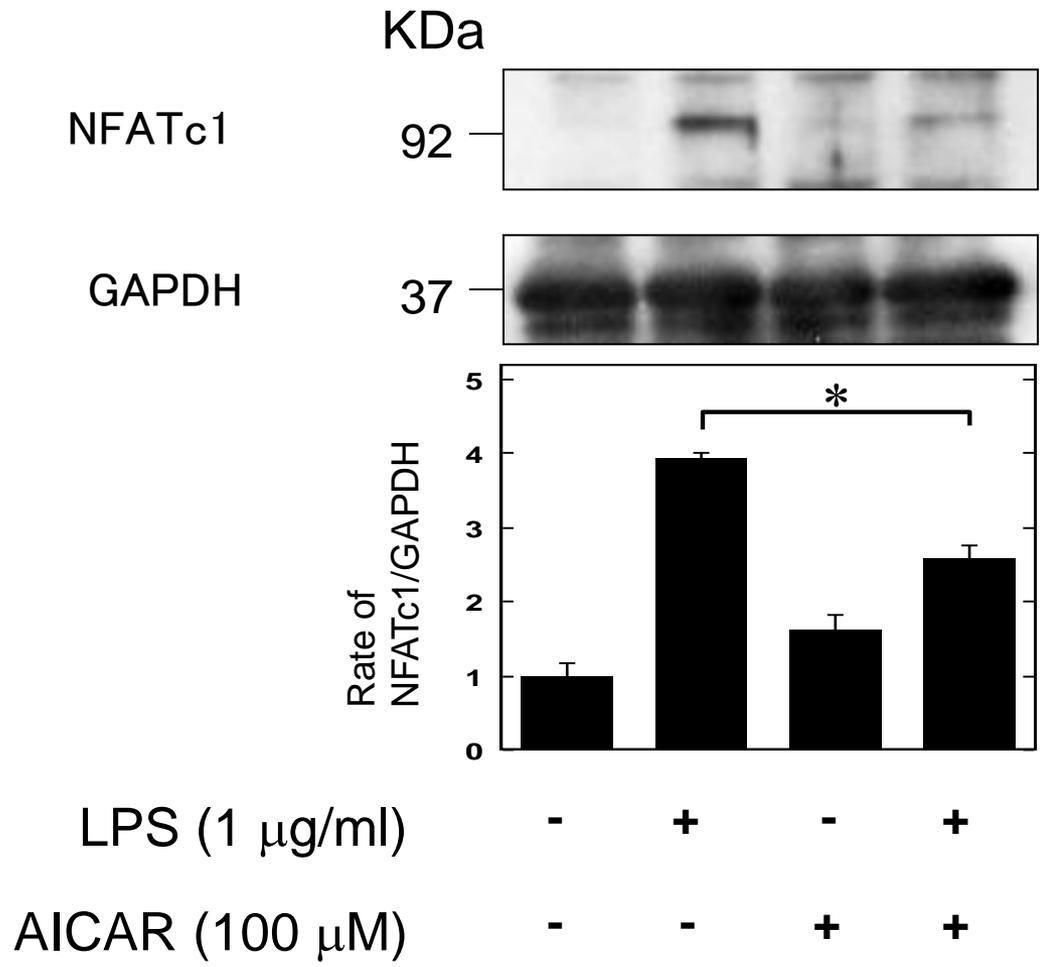
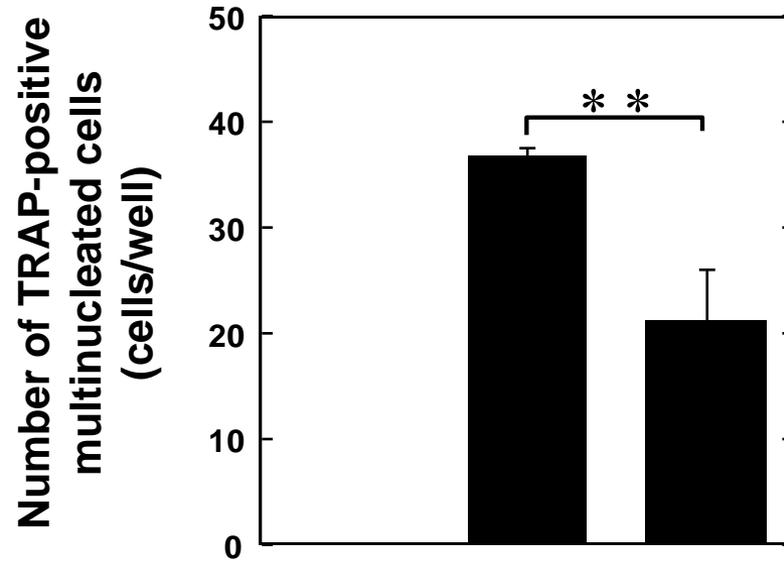


Fig. S2C



LPS (1 $\mu$ g/ml)	-	+	+
AICAR (100 $\mu$ M)	-	-	+
M-CSF (100 ng/ml)	+	+	+

Fig. S2D

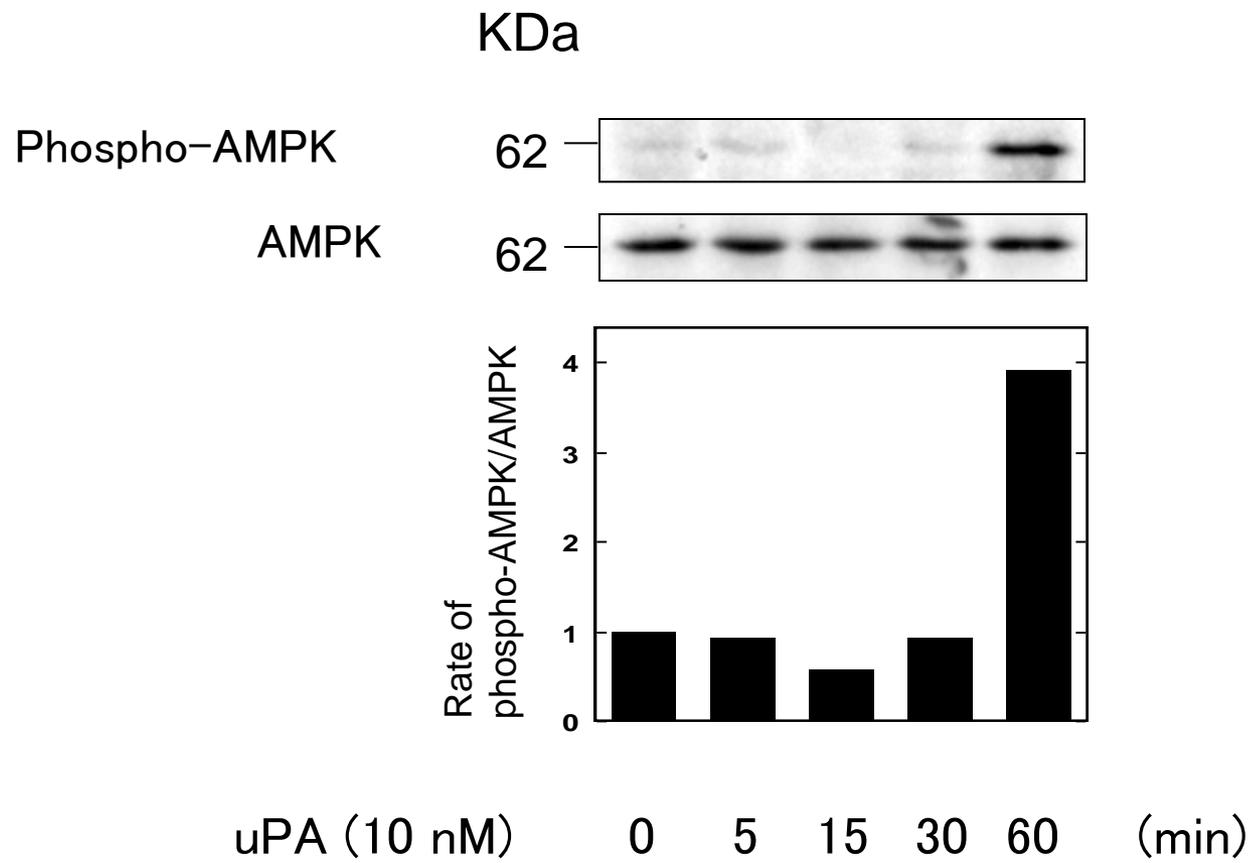
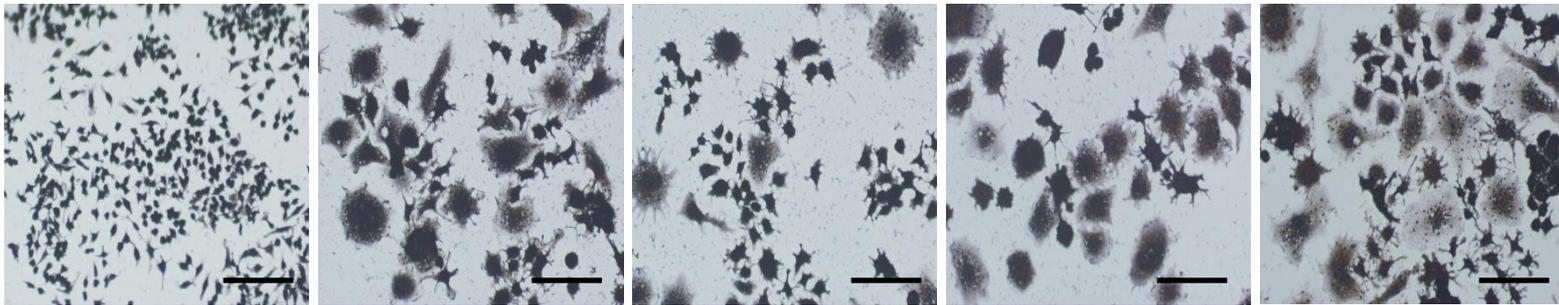


Fig. S2E



LPS (1 $\mu\text{g/ml}$ )	-	+	+	+	+
uPA (10 nM)	-	-	+	+	+
Compound C ( $\mu\text{M}$ )	-	-	-	<b>0.1</b>	<b>0.5</b>

Fig. S2F

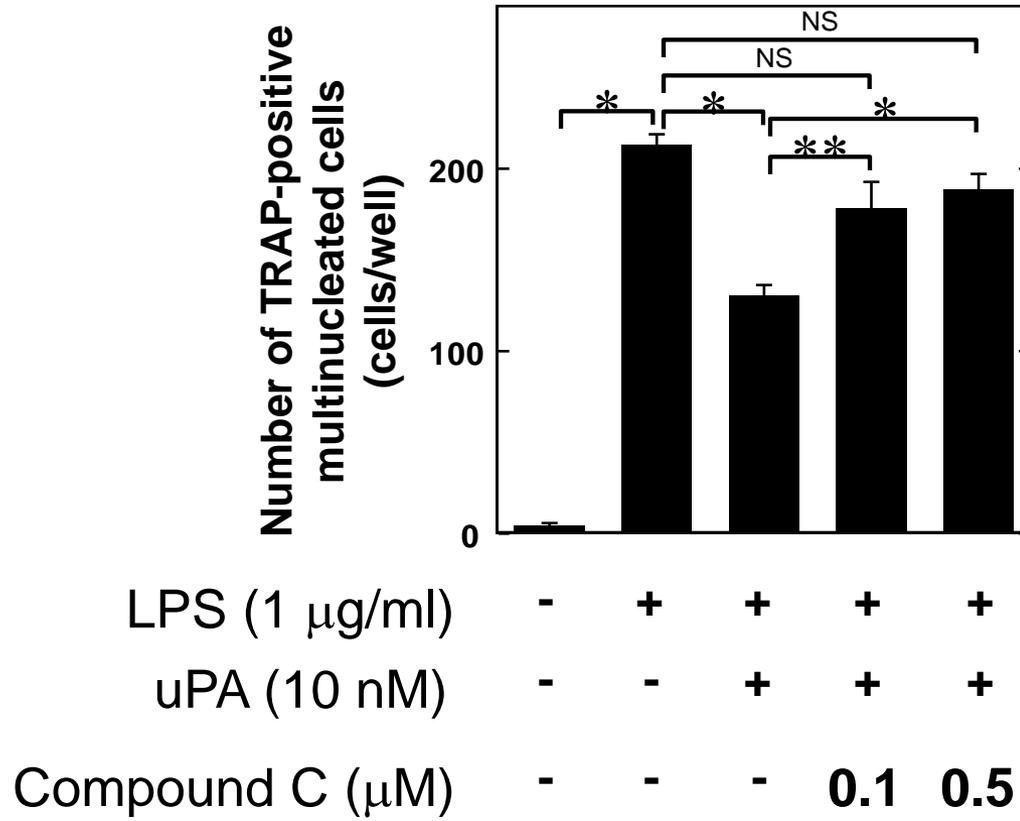


Fig. S2G

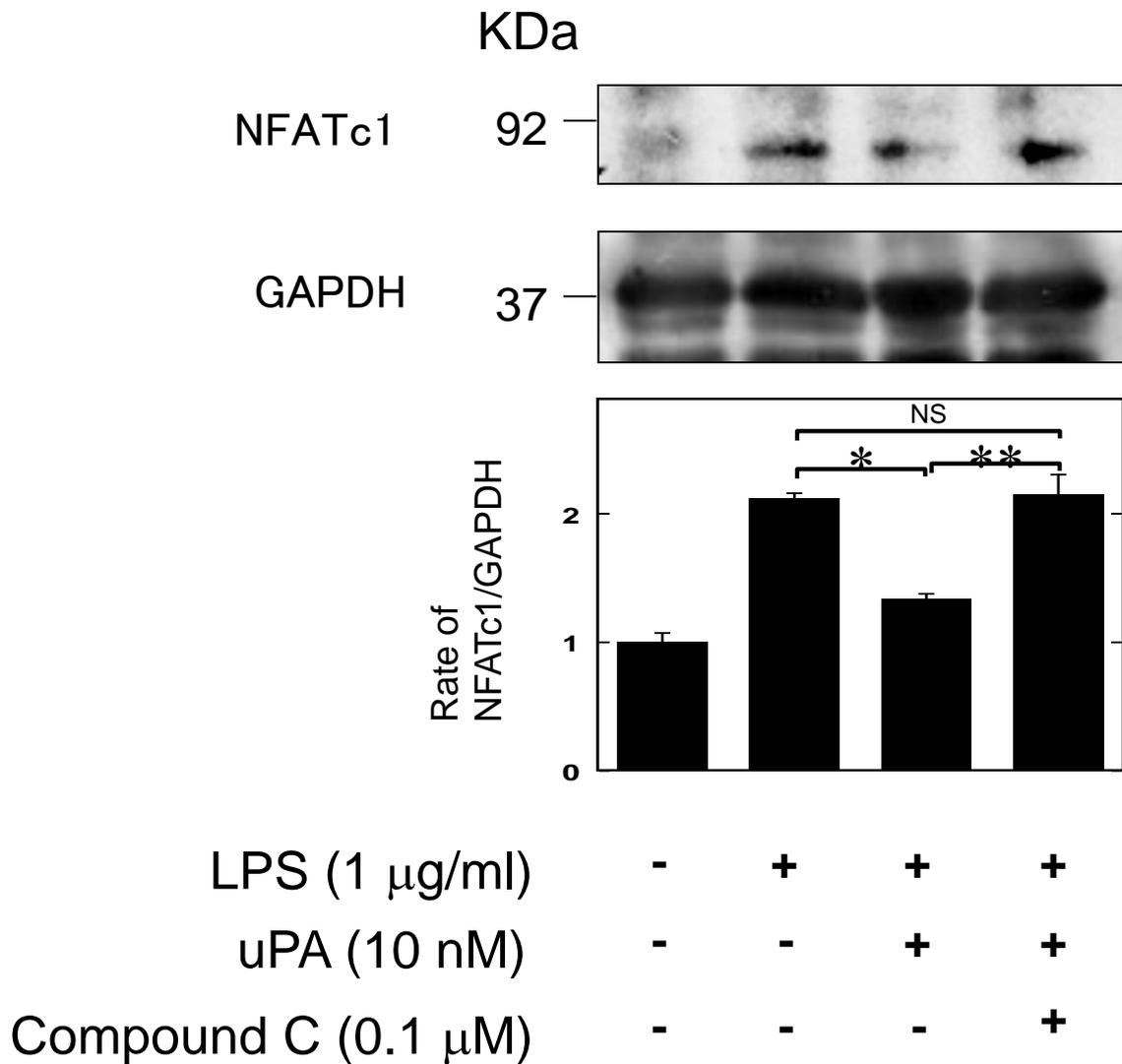
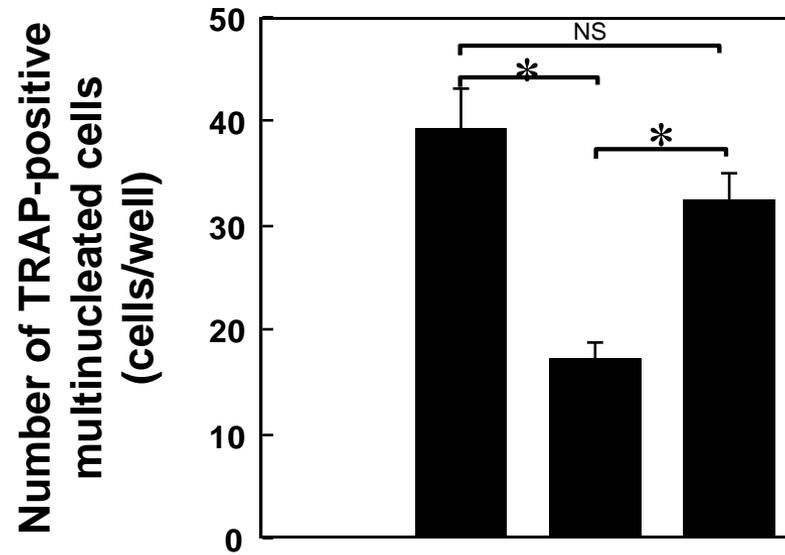


Fig. S2H



LPS (1 $\mu$ g/ml)	-	+	+	+
uPA (10 nM)	-	-	+	+
Compound C (0.1 $\mu$ M)	-	-	-	+
M-CSF (100 ng/ml)	+	+	+	+

Fig. S2I

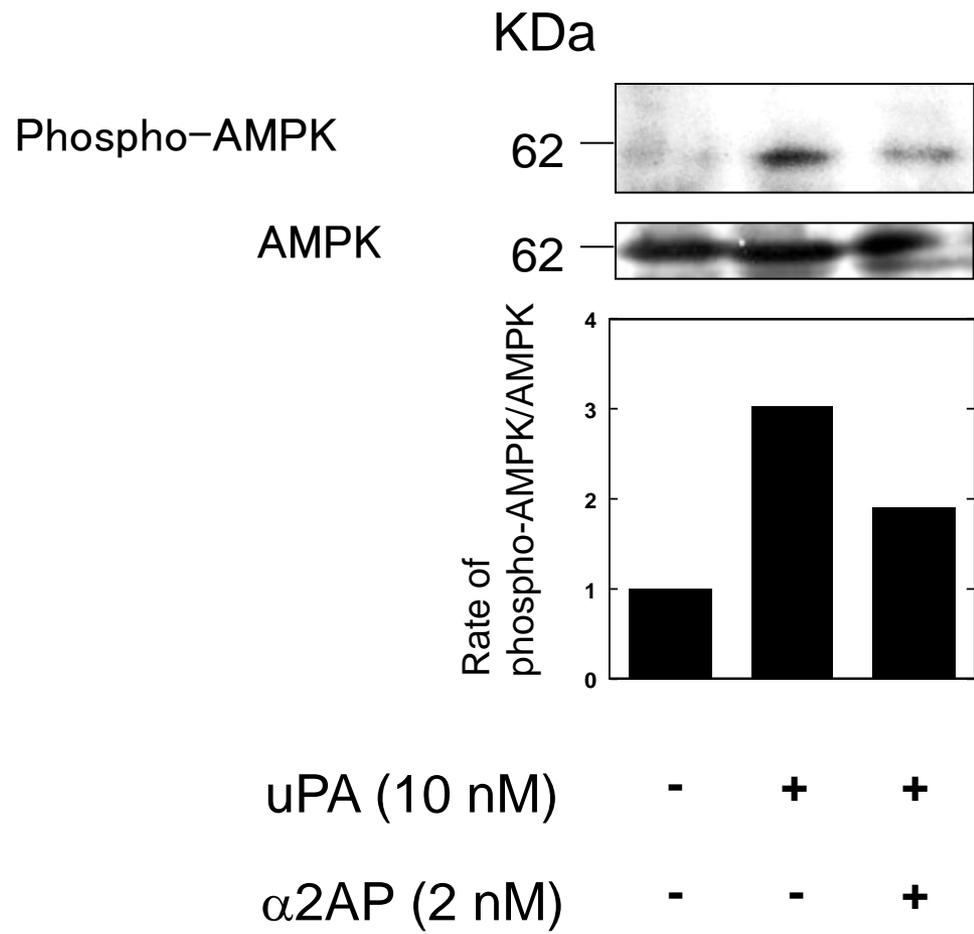


Fig. S2J

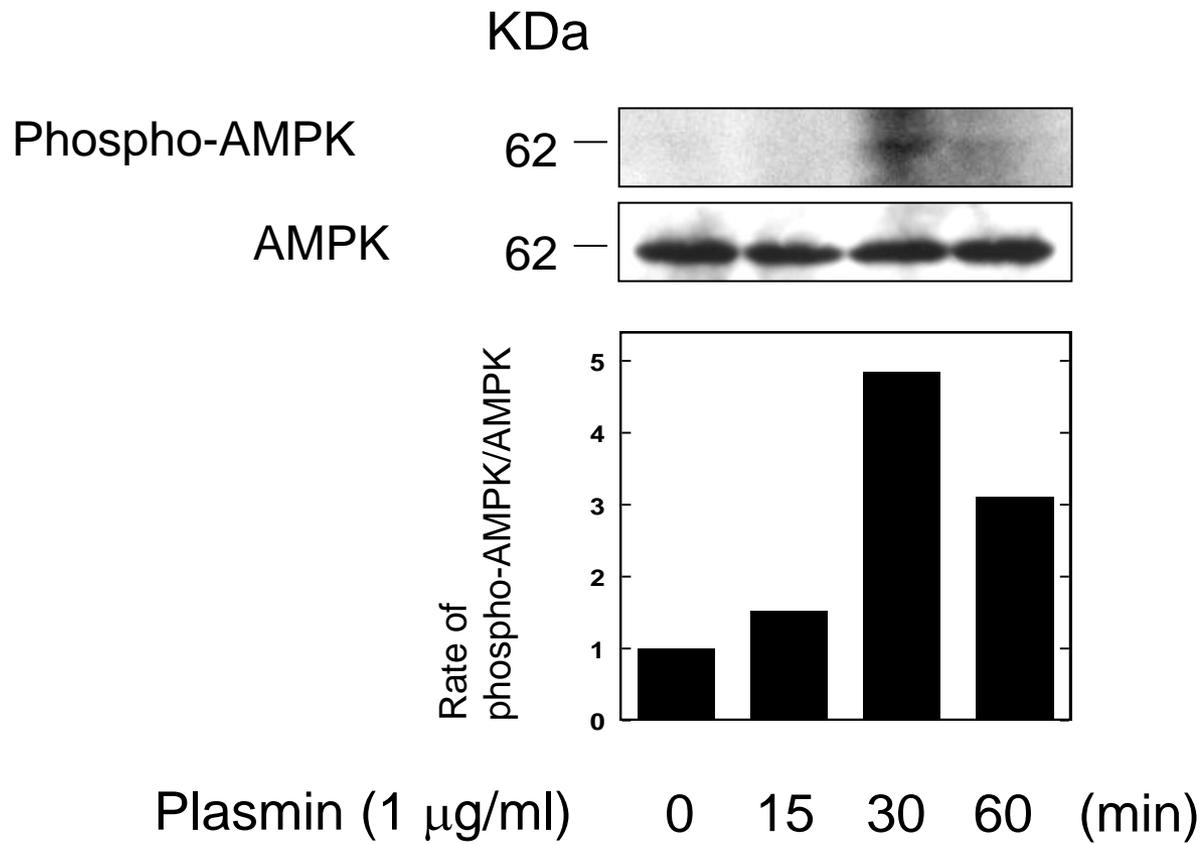


Fig. S2K

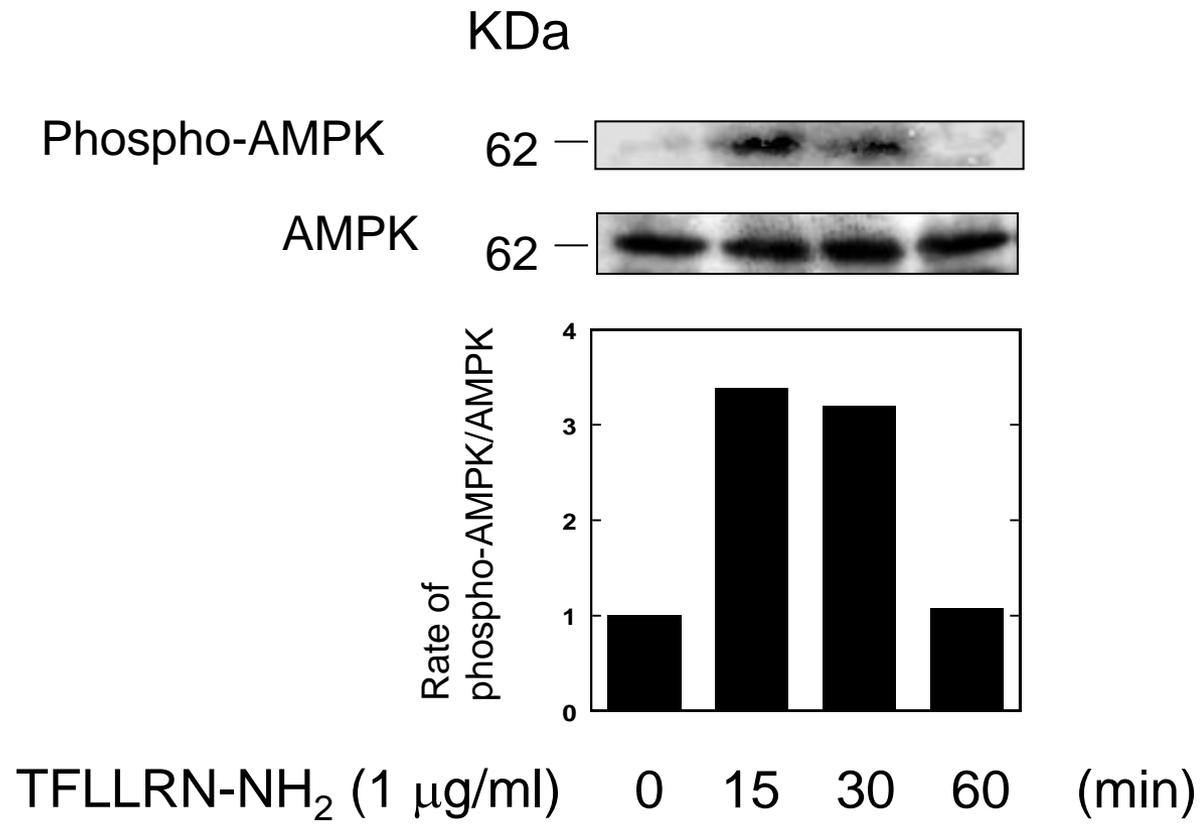


Fig. S2L

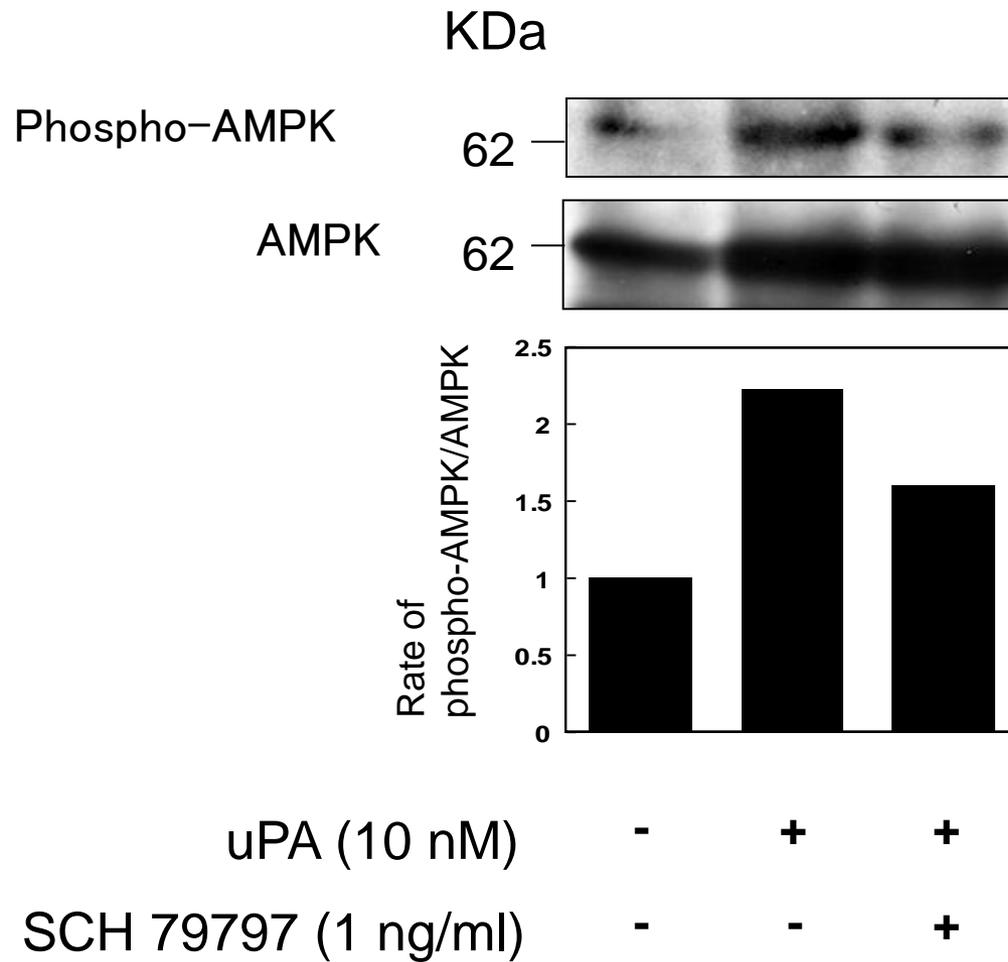


Fig. S2M