

Research Paper

Yorkie Facilitates Organ Growth and Metamorphosis in *Bombyx*

Shumin Liu¹, Panli Zhang^{1,2}, Hong-Sheng Song², Hai-Sheng Qi³, Zhao-Jun Wei³, Guozheng Zhang⁴, Shuai Zhan¹, Zhihong Liu⁵ ✉, Sheng Li¹ ✉

1. Key Laboratory of Insect Developmental and Evolutionary Biology, Institute of Plant Physiology and Ecology, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, Shanghai 200032, China
2. College of Life Sciences, Shanghai University, Shanghai 200444, China
3. School of Biotechnology and Food Engineering, Hefei University of Technology, Hefei 230009, China
4. College of Biotechnology, Jiangsu University of Science and Technology, Zhenjiang Jiangsu 212018, China
5. Department of Urology, Shanghai General Hospital, Medical School of Shanghai Jiao Tong University, Shanghai 200080, China

✉ Corresponding authors: lisheng01@sibs.ac.cn, drzhihongliu@sjtu.edu.cn

© Ivyspring International Publisher. Reproduction is permitted for personal, noncommercial use, provided that the article is in whole, unmodified, and properly cited. See <http://ivyspring.com/terms> for terms and conditions.

Received: 2016.01.03; Accepted: 2016.05.06; Published: 2016.06.07

Abstract

The Hippo pathway, which was identified from genetic screens in the fruit fly, *Drosophila melanogaster*, has a major size-control function in animals. All key components of the Hippo pathway, including the transcriptional coactivator Yorkie that is the most critical substrate and downstream effector of the Hippo kinase cassette, are found in the silkworm, *Bombyx mori*. As revealed by microarray and quantitative real-time PCR, expression of Hippo pathway genes is particularly enriched in several mitotic tissues, including the ovary, testis, and wing disc. Developmental profiles of Hippo pathway genes are generally similar (with the exception of Yorkie) within each organ, but vary greatly in different tissues showing nearly opposing expression patterns in the wing disc and the posterior silk gland (PSG) on day 2 of the prepupal stage. Importantly, the reduction of Yorkie expression by RNAi downregulated Yorkie target genes in the ovary, decreased egg number, and delayed larval-pupal-adult metamorphosis. In contrast, baculovirus-mediated Yorkie^{CA} overexpression upregulated Yorkie target genes in the PSG, increased PSG size, and accelerated larval-pupal metamorphosis. Together the results show that Yorkie potentially facilitates organ growth and metamorphosis, and suggest that the evolutionarily conserved Hippo pathway is critical for size control, particularly for PSG growth, in the silkworm.

Key words: Hippo pathway; Yorkie function; size-control; ovary; silk gland; wing disc; domestication; *Bombyx mori*

Introduction

One fundamental biology question is how animals grow to the right size. The mechanism by which animals orchestrate the growth of their individual cells, tissues and organs is a long-standing mystery. Hormones and nutrients play certain roles in the regulation of tissue growth, but organ size is modulated in intact animals by genetic signaling networks. During the last decade, the Hippo pathway, which was first shown to regulate cell proliferation and apoptosis during development and regeneration, has been recognized as one of the most important mechanisms of size control in animals. Many of the key components in the Hippo pathway and its basic signal transduction steps were originally discovered

from genetic screens in the fruit fly, *Drosophila melanogaster*, and this pathway was later found to be the major size control mechanism that is evolutionarily and functionally conserved in vertebrates [1-6].

In *Drosophila*, key components of the Hippo pathway fall into three main classes: the Hippo kinase cassette, downstream transcriptional regulators, and upstream inputs (Fig. S1).

(i) The Hippo kinase cassette. Hippo, Salvador (Sav), Warts, and Mob-as-tumor-suppressor (Mats), which form the core of the Hippo pathway, were discovered in tumor suppressor genetic screens in *Drosophila* [7-17]. Both Hippo and Warts are Ser/Thr

kinases, and Sav and Mats are the adaptor proteins of Hippo and Warts, respectively. Importantly, Hippo-Sav phosphorylates and activates Warts-Mats, and the four proteins form a complex kinase cascade.

(ii) The Hippo downstream transcriptional regulators. The transcriptional coactivator Yorkie is the most critical substrate and downstream effector of the Hippo pathway [2, 18-24]. Via direct protein-protein interaction, Yorkie is phosphorylated at three Ser residues (Ser168 is the most critical one in *Drosophila*) by Warts and is thus inactivated. Upon phosphorylation, Yorkie is relocated to the cytoplasm. When the Hippo pathway is inactivated, Yorkie functions as an oncogene in the nucleus by acting as a transcriptional coactivator. The Hippo pathway negatively regulates Yorkie activity by preventing its accumulation within the nucleus. Once in the nucleus, Yorkie binds and activates several transcription factors, including Scalloped (Sd), Homothorax (Hth), Teashirt, and Mothers against DPP, to induce gene expression. For example, Yorkie-Sd induces expression of *cyclin E (cycE)* and *inhibitor of apoptosis protein 1 (diap1)* to promote cell proliferation and to inhibit apoptosis, respectively [20, 21]. In addition, the microRNA *bantam*, the oncogene *Myc*, and several Hippo upstream regulators are potent target molecules of Yorkie.

(iii) The Hippo upstream inputs. Many of the Hippo upstream regulators control cellular processes such as apical-basal cell polarity, planar cell polarity, and cell-cell adhesion [1, 2, 5]. Fat is a large transmembrane protein that is critical for maintaining both planar cell polarity and the Hippo pathway. Dachshous (Ds) is a ligand for Fat [25-27]. Crumbs (Crb) is an apical transmembrane protein that organizes both apical-basal polarity and the Hippo signaling [28-30]. Expanded (Ex), Merlin (Mer), and Kibra colocalize at the apical domains of polarized epithelial cells, forming the Ex-Mer-Kibra complex. The three proteins physically interact with each other, and are partially redundant in activating the Hippo kinase cassette [31-34]. Crb binds to Ex, affecting Ex stability and localization and thus the Hippo pathway, but Ds-Fat signaling is genetically distinguishable from Hippo regulation by the Ex-Mer-Kibra complex [2]. Two other polarity complexes, the Scribble (Scrib) complex (Scrib; Discs large, Dlg; Lethal giant larva, Lgl) and the Par complex (Par3; Par6; atypical Protein kinase C, aPKC), are also involved in the Hippo pathway. The Lgl complex regulates apical-basal polarity by modulating the components of the Crb complex and the aPKC complex. In general, the Lgl complex activates the Hippo pathway, while the aPKC complex antagonizes the Lgl complex [35-37].

In mammals, a conserved Hippo kinase cascade and Yorkie function exist. However, the Hippo upstream inputs have additional complexity and, in some cases, are divergent from *Drosophila*. Mechanotransduction and G-protein-coupled receptor (GPCR) signaling were identified as additional Hippo upstream regulators. Importantly, actin cytoskeleton and cellular tension appear to be the master mediators that integrate and transmit upstream signals to the Hippo kinase cascade and Yorkie function [3-6]. The Hippo pathway is largely conserved in *Caenorhabditis elegans* [38]. Moreover, key components in the Hippo pathway are found in the cnidarians, a very basal group of metazoans, and even in the unicellular ancestors of animals, the amoeboid holozoans, showing that the hippo pathway evolved well before the origin of Metazoa [39, 40]. Moreover, a distinct interaction interface between Yorkie and Sd became structurally fixed in the eumetazoan common ancestor [41], showing that the ancient evolutionary history of the Hippo pathway as a key developmental mechanism in all animals.

Apart from *Drosophila*, the domesticated silkworm, *Bombyx mori*, is another model insect for general biology [42]. In this study, we found that the Hippo pathway is evolutionarily conserved in the silkworm. Temporal and spatial expression patterns suggest that all of the Hippo pathway genes coordinately regulate organ growth and growth of the whole body. Moreover, Yorkie facilitates organ growth and accelerates metamorphosis. This study provides potential for promoting growth of the silk gland and thus silk yield by genetic modification of Yorkie in the silk gland, and suggests that the evolutionarily conserved Hippo pathway might play a crucial role in size control in the silkworm.

Materials and Methods

Animals

Bombyx larvae (P50 strain) were raised and provided by the Sericultural Research Institute, Chinese Academy of Agricultural Sciences. The silkworms were reared with fresh mulberry leaves in the laboratory using previously described conditions [43].

Gene identification

Gene identification was performed according to a method we reported previously [43]. We mainly used SilkDB, <http://silkworm.genomics.org.cn/>, to search for potential Hippo pathway genes. First, we used the sequences of 18 key proteins in the *Drosophila* Hippo pathway to search against the GLEAN gene collection to identify Hippo pathway genes in the silkworm by local BLASTP, with an E-value threshold

of 10^{-6} . KAIKObase, <http://sgp.dna.affrc.go.jp/KAIKObase/>, was used for obtaining some full length cDNA sequences. The identified putative Hippo pathway genes were validated by searching the NCBI protein database with the putative Hippo pathway gene sequences as queries. Furthermore, each potential Hippo pathway gene was analyzed using Pfam to identify its domains. Finally, the identified silkworm Hippo pathway genes were used to search both SilkDB and KAIKObase to avoid missing any genes.

Phylogenetic analysis

At least one representative genome was selected for most orders of sequenced arthropod species, whose genomic information is available from NCBI, <http://www.ncbi.nlm.nih.gov> or predicted from the genome databases. The arthropod species (and Yorkie sequences) used are *Ixodes scapularis* (XP_002399565.1), *Stegodyphus mimosarum* (KFM74842.1), *Strigamia maritima* (SMAR013599-PA, predicted), *Daphnia pulex* (EFX70433.1), *Timema cristinae* (Tcri27797, predicted), *Zootermopsis nevadensis* (KDR09902.1), *Drosophila melanogaster* (XP_001976544.1), *Tribolium castaneum* (XP_970492.1), *Apis mellifera* (XP_391844.3), *Pediculus humanus humanus* (XP_002433100.1), *Acyrtosiphon pisum* (XP_001948042.1), *Plutella xylostella* (XP_011558709.1), *Amyeloides transitella* (XP_013194179.1), *Papilio polytes* (XP_013134895.1), *Helicoverpa armigera* (ALO18798.1), and *Bombyx mori* (NP_001116819.1). The protein sequence of *Drosophila* Yorkie was used as the seed to search for putative orthologs across the whole genome by reciprocal BLAST. Multiple alignments of Yorkie proteins were then performed using MUSCLE [44], and the two WW domains of these alignments were extracted using Gblocks [45]. Maximum-likelihood phylogenies were calculated using PhyML [46] with the JTT model for 100 replicates.

Microarray analysis

We initially used the genome-wide microarray data of the silkworm, which was previously performed in 2007 [47], to profile the expression patterns of Hippo pathway genes in multiple larval tissues and during metamorphosis. From the public SilkDB, we downloaded the normalized microarray data for genome-wide gene expression in a number of tissues on day 3 of the fifth larval instar (L5-3) and at 19 sequential time points during metamorphosis. The expression pattern of the Hippo pathway genes was estimated from intensity values. If the normalized intensity of a Hippo pathway gene value exceeded 0, the gene was considered to be expressed [47].

Quantitative real-time PCR

Total RNA was extracted for quantitative real-time PCR (qPCR) analysis as previously described in detail [43]. qPCR was carried out in a 20 μ l reaction solution, which contains 10 μ l of SYBR Green real-time PCR Master Mix (Bio-Rad, USA), 5 μ l of first strand cDNA template, and 0.5 mM of each primer. The iQ5 Real-Time PCR Detection System (Bio-Rad, USA) was employed. Rp49 was chosen as a reference gene for qPCR analysis. Table S1 shows the primers used in this paper.

RNA interference of Yorkie in *Bombyx larvæ*

EGFP (full length) and *Yorkie* (101-500 bp) dsRNA was generated using the T7 RiboMAXTM Express RNAi system (Promega, USA). At the initiation of the early wandering stage (IW), each larva was injected with either *EGFP* dsRNA (30 μ g) or *Yorkie* dsRNA (30 μ g). Twenty-four h after RNA interference (RNAi) treatment, the larvae were sacrificed [43]. Ovary/testis, posterior silk gland (PSG), wing disc, and fat body tissues were collected for further analysis. For the RNAi experiments, 30 animals were used for each group, and 3 biological replicates were conducted.

Baculovirus-mediated overexpression of Yorkie and Yorkie^{CA} in *Bombyx larvæ*

The *Bombyx* Yorkie Ser97 corresponds to the *Drosophila* Yorkie Ser168. According to the original discovery in *Drosophila* [19], Ser97 of the *Bombyx* Yorkie was mutated to Ala97 to generate the constitutive-active form of Yorkie (Yorkie^{CA}) by using PCR-mediated site-directed mutagenesis. The V5 tag (encoding the amino acid sequence of GKPIPPLLGLDST) sequence was fused in the 5' end of Yorkie or Yorkie^{CA} to generate V5-Yorkie or V5-Yorkie^{CA}. V5-Yorkie or V5-Yorkie^{CA} was cloned into the EcoRI-NotI sites of the pFastBac-HTa (Invitrogen) plasmid, and *DsRed2* (a type of red fluorescent protein, RFP) was used as a control.

We have previously employed the AcNPV ecdysteroid UDP-glucosyl transferase (*egt*) mutant for gene overexpression in *Bombyx* [48, 49], but this baculovirus does not produce sufficient Yorkie and Yorkie^{CA} proteins for functional studies. Here, using the homologous recombination technique, we generated an *egt* mutant of *Bombyx mori* nucleopolyhedrosis baculovirus (BmNPV). This virus allows silkworms to survive until pupation and to produce sufficient Yorkie and Yorkie^{CA} proteins. The pFastBac-HTa plasmids of V5-Yorkie and V5-Yorkie^{CA} were then transformed into DH10Bac Δ EGT bacteria 47 to prepare bacmid DNA according to the protocol of the Bac-to-Bac system. Bm-N cells were transfected

with bacmid DNA (1 µg/ml) using Cellfectin II transfection reagent (Invitrogen). After 7 days, P1 virus was collected. The P1 virus was then used to prepare P2 virus after another 3 days. Bm-N cells infected with the P2 virus were collected for silkworm injection experiments on L5-2. One hundred and twenty h after virus treatment, the larvae were sacrificed for analyses. For the overexpression experiments, 30 animals were used for each group, and 3 biological replicates were conducted.

Western blotting and immunohistochemistry

The V5 antibody (1:1000) (Invitrogen, USA) was used to detect the overexpressed *V5-Yorkie* or *V5-Yorkie^{CA}* in the PSG by Western blotting as previously described in detail [49].

For detecting the overexpressed *V5-Yorkie* or *V5-Yorkie^{CA}* by immunohistochemistry, the newly dissected fat body tissues were fixed in 4% paraformaldehyde for 45 min at room temperature. Fat body tissues were blocked in phosphate buffered saline (PBS) containing 5% BSA and 1% Triton-X (PBSBT) for 1 h, and then incubated with the V5 antibody (diluted 1:50) at 4°C overnight. Fat body tissues were washed multiple times for 1 h in PBSBT, and then incubated with a FITC-(green) conjugated secondary antibody (diluted 1:200) for 2 h. DAPI (Sigma) was added to label nuclei. Fat body tissues stained with V5 antibody and DAPI were imaged using an FV10-ASW confocal microscope (Olympus, Japan) at 20×magnification [49].

Statistics

Throughout the study, the experimental data were analyzed using Student's t-test: *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$. Meanwhile, the values are represented as the mean ± standard deviation.

Results

Identification of key genes in the Hippo pathway

Eighteen key proteins in the *Drosophila* Hippo pathway were used in BLASTP queries against the SilkDB and KAIKObase databases. Each potential protein in the *Bombyx* Hippo pathway was validated by searching its known orthologs from the NCBI protein database and further searching its conserved domain(s) with the Pfam program. As a result, a total of 18 putative genes in the Hippo pathway were identified in the silkworm genome. Their encoding proteins fall into three main classes: the Hippo kinase cassette (Hippo, Sav, Warts, and Mats), downstream transcriptional regulators (Yorkie and Sd), and upstream inputs (Ds and Fat; Crb; Ex, Mer, and Kibra; Scrib, Dlg, and Lgl; and Par3, Par6, and aPKC) (Fig.

S1). As revealed by domain analysis (Fig. 1), Hippo is a Ste20 family protein kinase, Sav contains one WW domain, Warts is an NDR family protein kinase, and Mats has a Mob1/phocein domain. Yorkie contains two WW domains, and Sd belongs to the TEA transcriptional enhancer factor superfamily. The transmembrane proteins Ds and Fat contain 22 and at least 10 (the Fat sequence is likely incomplete) cadherin repeats, respectively. The transmembrane protein Crb has multiple epidermal growth factor (EGF)-like domains and Laminin G-like domains. Ex and Mer are similar to each other, containing FERM (4.1 protein, ezrin, radixin, moesin) domains; Kibra has two WW domains, one C2 domain, and one component of oligomeric Golgi complex 7 (COG7) domain. Scrib has multiple leucine-rich repeats; Dlg contains one Guanylate kinase domain and three PDZ domains, and Lgl belongs to the WD40 superfamily. Par3 contains two PDZ domains and one DUF3534 domain; Par6 has one PDZ domain and one PB1 domain, while aPKC is an atypical protein kinase C.

In SilkDB, eight Hippo pathway genes have evidence for mRNA expression with expressed sequence tags (ESTs), which were collected from 36 different cDNA libraries. Based on the EST information, *Sd* shows the highest transcript level with 10 hits. The largest Hippo pathway protein (2764 amino acids) and the smallest one (217 amino acids) are encoded by *Ds* and *Mats*, respectively. Of the 18 Hippo pathway genes, 17 are dispersed on 11 chromosomes and 1 (*Mer*) on an unmapped scaffold. With the exception of *Fat*, the other 17 Hippo pathway genes have accession numbers in NCBI (Table 1).

The 18 Hippo pathway genes were also searched from 16 arthropod genomes. Each arthropod contains at least one copy (with occasionally gene duplication) of each of the 18 genes, indicating that the *Drosophila* Hippo pathway is evolutionarily conserved in Arthropoda. We selected Yorkie, which is the most critical substrate and downstream effector of the Hippo pathway, to perform a multiple sequence alignment (Table S2). The comparison analysis reveals that the Yorkie proteins among five lepidopteran species share the highest identity (73-89%) and that the identity of any two arthropod Yorkie proteins is greater than 34%. A previous phylogenetic analysis shows that arthropod Yorkie proteins form a separate clade among all the animal Yorkie orthologs [39]. We then constructed a phylogenetic tree on the basis of the two WW domains of Yorkie proteins from the 16 arthropods (Fig. 2), including the five lepidopteran insects for more informative comparison. Importantly, the nodes within the lepidopteran clade present a strong bootstrap support, and the

phylogenetic tree shows a clear separation between Insecta and non-Insecta[50].

Expression of the Hippo pathway genes is enriched in mitotic tissues

To understand the physiological function of the Hippo pathway in *Bombyx*, we initially surveyed the temporal and spatial expression patterns of all 18 Hippo pathway genes by microarray analysis. All of the 18 genes have corresponding probes on the oligonucleotide chip [47]. We investigated the tissue

distribution of the 18 genes on L5-3 using the microarray data of tissue-specific expression [47]. The expression of 7 genes (including *Yorkie*) can be detected in at least one of the selected tissues, including testis, ovary, head, integument, Malpighian tubules, fat body, hemocytes, anterior and middle silk glands, and PSG (Fig. S2). Significantly, most genes were highly expressed in the gonad (ovary and testis) and the head compared to other tissues.

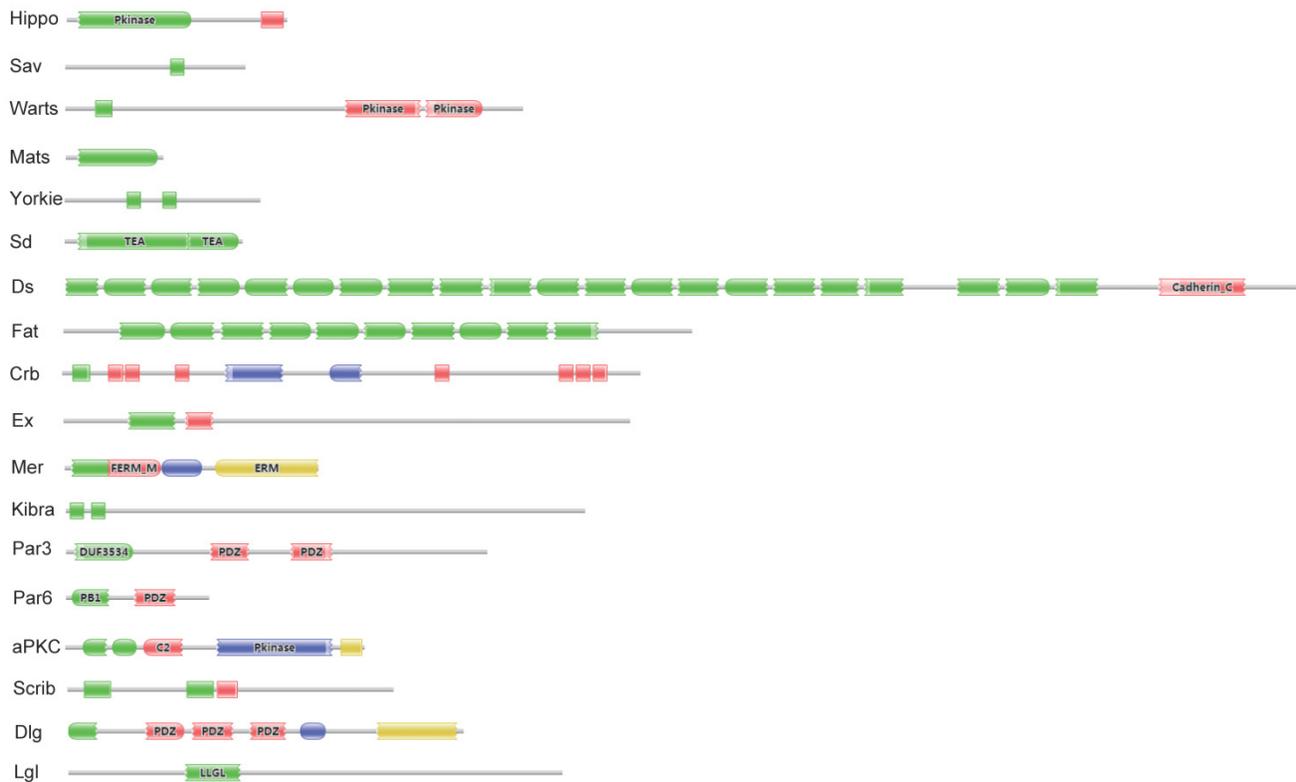


Fig. 1. Significant domains of 18 Hippo pathway genes in *Bombyx*. The program Pfam was used for domain identification. Please see the details of domain analysis in the text. * The *Fat* sequence is likely incomplete in the *Bombyx* genome.

Table I: Summary of the 18 Hippo pathway genes in the *Bombyx* genome.

NO.	gene	SilkDB ID	Chr.	Scaffold	Position	EST	Probe	Size (AA)	NCBI ID
1	<i>Hippo</i>	BGIBMGA013478	5	nscaf3075	(+)1302878..1290675	0	sw08605	492	NP_001116818.1
2	<i>Sav</i>	BGIBMGA004937	25	nscaf2822	(-)912627..918022	0	sw11181	402	XP_004924516.1
3	<i>Warts</i>	BGIBMGA006330	6	nscaf2852	(+)1986876..1992247	1	sw05368	1025	XP_004923559.1
4	<i>Mats</i>	BGIBMGA013463	6	nscaf3074	(+)146991..151388	0	sw05526	217	XP_004926597.1
5	<i>Yorkie</i>	BGIBMGA003682	5	nscaf2674	(+)4049793..4052521	2	sw03123	437	NP_001116819.1
6	<i>Sd</i>	BGIBMGA001129	13	nscaf1898	(+)4585777..4618439	10	sw14392	397	XP_004928021.1
7	<i>Ds</i>	BGIBMGA010453	12	nscaf2993	(-)1494834..1480580	0	sw11749	2764	XP_004929431.1
8	<i>Fat</i>	BGIBMGA010597	12	nscaf2995	(+)253973..259252	0	sw21999	1407	NO
9	<i>Crb</i>	BGIBMGA007609	15	nscaf2888	(-)9505337..9522337	0	sw11252	1294	XP_004928825.1
10	<i>Ex</i>	BGIBMGA010558	12	nscaf2993	(+)6011036..6015638	5	sw03959	1268	XP_004929583.1
11	<i>Mer</i>	BGIBMGA014502	UN	nscaf749	(-)6487..18440	0	sw11071	567	XP_004933720.1
12	<i>Kibra</i>	BGIBMGA009269	14	nscaf2943	(+)1377742..1396051	2	sw05915	1162	XP_004922701.1
13	<i>Scrib</i>	BGIBMGA005373	8	nscaf2828	(-)1127705..1141772	1	sw18238	729	XP_004932102.1
14	<i>Dlg</i>	BGIBMGA010382	12	nscaf2993	(-)6758200..6776168	0	sw19956	884	XP_004929498.1
15	<i>Lgl</i>	BGIBMGA005570	17	nscaf2829	(-)1712186..1720814	3	sw12635	1105	XP_004921966.1
16	<i>Par3</i>	BGIBMGA006557	10	nscaf2855	(+)6535850..6549347	0	sw11025	942	XP_004930528.1
17	<i>Par6</i>	BGIBMGA004440	20	nscaf2795	(+)718639..722191	1	sw09724	319	XP_004922525.1
18	<i>aPKC</i>	BGIBMGA014132	8	nscaf463	(+)1220257..1230024	0	sw05322	669	NP_001036978.1

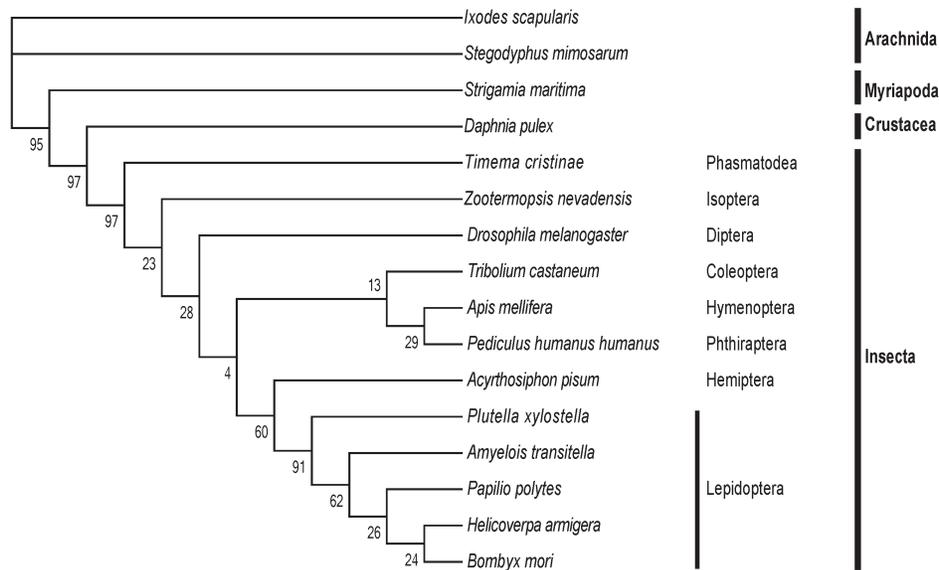


Fig. 2. The phylogenetic tree of Yorkie in Arthropoda. At least one representative genome was selected for most orders of sequenced arthropod species. In total, 16 arthropods were used, including Arachnida (2 species), Myriapoda, Crustacea, and Insecta (12 species, including five lepidopteran species). The protein sequence of *Drosophila* Yorkie was used to search putative orthologs across the whole genomes of the 16 arthropods by reciprocal BLASTP. Multiple alignments of the two WWV domains of Yorkie proteins were then performed using MUSCLE, and the conserved blocks of these alignments were extracted using Gblocks. The phylogeny was calculated using PhyML with the JTT model for 100 replicates.

All 18 genes were also analyzed by qPCR using total RNA isolated from 9 tissues, including fat body, midgut, ovary, wing disc, trachea, Malpighian tubules, hemocyte, prothoracic gland, and silk gland on L5-2. Except *Sd* and *Fat*, the other 16 genes were abundantly expressed in the gonad. *Fat* and *Ex* were highly expressed in the wing disc. *Sd* expression was high in the prothoracic glands (Fig. 3). The tissue distribution patterns of *Scrib*, *Dlg*, and *Lgl* were previously reported by Qi et al [51]. In consistent with the microarray data, qPCR analysis revealed that gene expression of the Hippo pathway components is particularly enriched in several mitotic tissues, including the gonad and wing disc on L5-2.

Developmental profiles of the Hippo pathway genes are generally similar within each organ but vary greatly in different tissues

We initially investigated the temporal expression patterns of 18 Hippo pathway genes from L5-4 to the adult stage by using the microarray data of developmental changes of mRNA [47]. The expression of 11 of the 18 genes (including *Yorkie*) was detected (Fig. S3). With the exception of *Par3* and *Fat*, which were only detected in male pupae, the expression of the other 9 genes gradually rises from the larval stage to the pupal stage to the adult stage.

To understand the detailed developmental profiles of the 18 genes during *Bombyx* larval-pupal metamorphosis, we performed qPCR analysis using total RNA isolated from the ovary, wing disc, and

PSG dissected from 4L-3 to day 2 of the prepupal stage (PP2). In the ovary, with the exception of *Sd*, *Fat*, *Par3*, and *Scrib*, the expression levels of the other 14 genes were higher in the fourth instar than in the fifth instar, and the expression levels of 11 of the 18 genes (*Hippo*, *Warts*, *Mats*, *Yorkie*, *Crb*, *Mer*, *Kibra*, *Dlg*, *Lgl*, *Par3*, and *Par6*) increased on PP2 (Fig. 4). In the wing disc, the expression levels of all 18 genes were low on 4L-3 and relatively high from 4L-4 to PP1; with the exception of *Yorkie*, the expression levels of the other 17 genes decreased on PP2 (Fig. S4). In the PSG, the expression levels of 9 of the 18 genes (*Hippo*, *Mats*, *Sd*, *Crb*, *Ex*, *Mer*, *Scrib*, *Par6*, and *aPKC*) were low from 4L-3 to PP1; the expression levels of 6 of the 18 genes (*Sav*, *Ds*, *Fat*, *Kibra*, *Dlg*, and *Lgl*) gradually decreased from 4L-3 to PP1, while the expression levels of *Warts*, *Yorkie*, and *Par3* were quite stable during this period. Apart from *Yorkie*, the expression levels of the other 17 genes significantly increased on PP2 (Fig. 5). Collectively, the developmental profiles of the 18 Hippo pathway genes were quite similar (with the exception of *Yorkie*) within each organ but varied greatly in different tissues, and all 18 Hippo pathway genes show nearly opposing expression patterns in the wing disc and the PSG on PP2.

Yorkie RNAi delays metamorphosis and ovary maturation

In *Drosophila*, *Yorkie* mutants display reduced organ growth [18]. Thus, we investigated whether *Yorkie* is important for *Bombyx* development using

RNAi to suppress *Yorkie* expression (*Yorkie* RNAi) at IW. Interestingly, the wandering behavior was apparently delayed 12 h after *Yorkie* RNAi, compared with the *EGFP* RNAi control animals (Figs. 6A and 6A'). Moreover, all of the *Yorkie* RNAi animals remained at the pupal stages 240 h after RNAi treatment, while all of the control animals had already emerged as adults. The delay of adult emergence in the *Yorkie* RNAi animals was about 12 h (Fig. 6B). Notably, 24 h after *Yorkie* RNAi, *Yorkie* expression in the ovary decreased by approximately 90% compared

with the control levels; moreover, six *Yorkie* target genes, including *cycE*, *IAP*, *Ex*, *Mer*, *Hth*, and *Sd*, were significantly downregulated by *Yorkie* RNAi (Fig. 6C). Importantly, the *Yorkie* RNAi females laid fewer eggs than the control females (Fig. 6D and D'). In addition, the efficiency of *Yorkie* RNAi in the wing disc was moderate, and *Yorkie* RNAi only slightly decreased its organ size (Figs. S5A and S5A'). The RNAi experiment indicates that *Yorkie* is required for timely metamorphosis and ovary maturation in *Bombyx*.

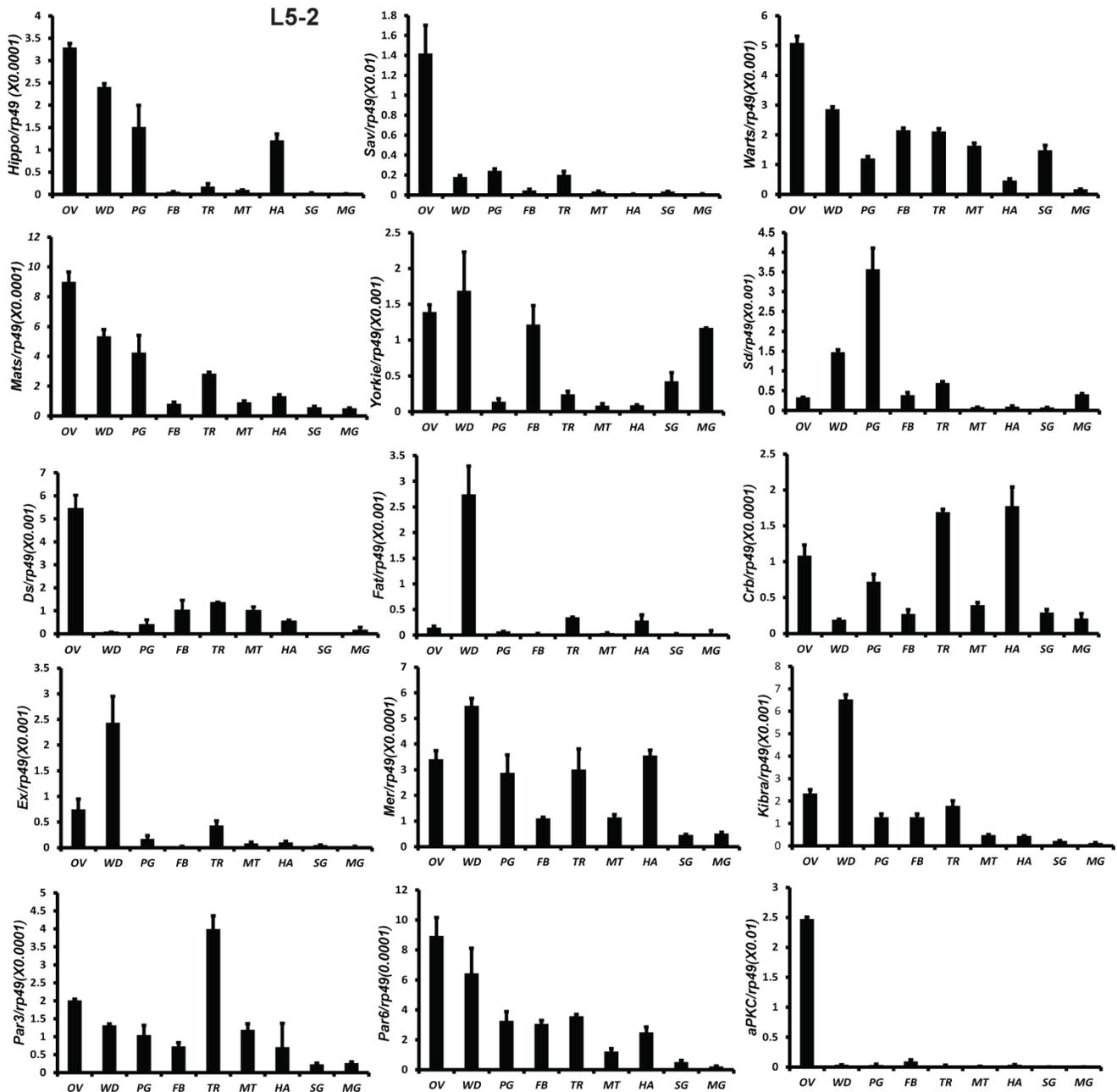


Fig. 3. Tissue distribution patterns of Hippo pathway genes on day 2 of the fifth larval instar. OV (ovaries); WD (Wing disc); PG (Prothoracic gland); FB (Fat body); TR (Trachea); MT (Malpighian tubule); HA (Hemocytes); SG (Silk gland); MG (Midgut). *Rp49* is used as an internal control. Relative mRNA levels are indicated as the ratios of mRNA levels between the target gene and *Rp49*. Error bars represent the SDs of three independent replicates in this and all subsequent figures. In each replicate, ten animals were used, and triplicate qPCR analyses were conducted.

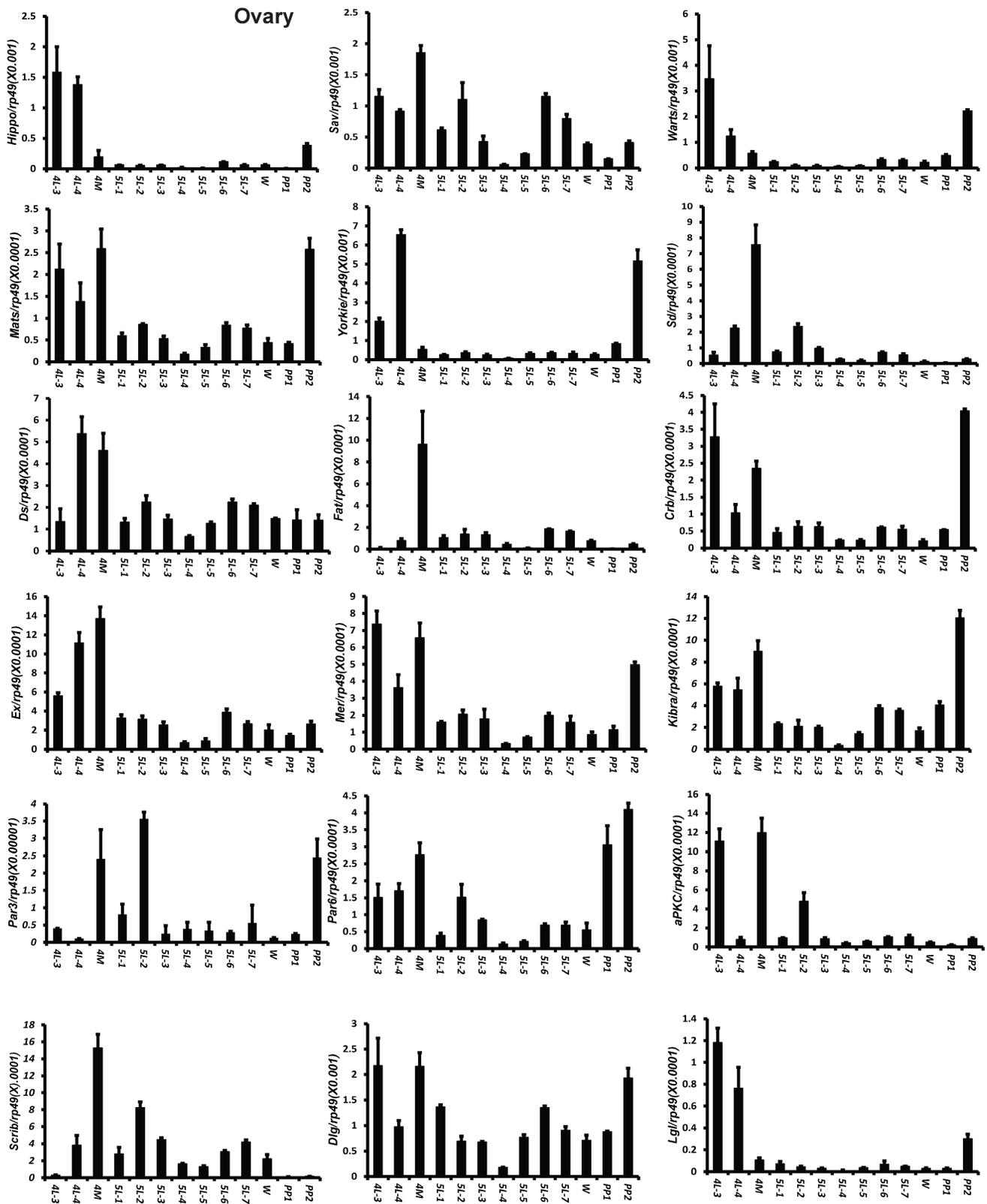


Fig. 4. Developmental expression profiles of 18 Hippo pathway genes in the ovary from day 3 of the fourth larval instar to day 2 of the prepupal stage. 4L-3 and 4L-4, day 3 and 4 of the fourth larval instar; 4M, the 4th molting; 5L-1 to 5L-7, day 1 to 7 of the fifth larval instar; W, the wandering stage; PP1 and PP2, day 1 and 2 of the prepupal stage.

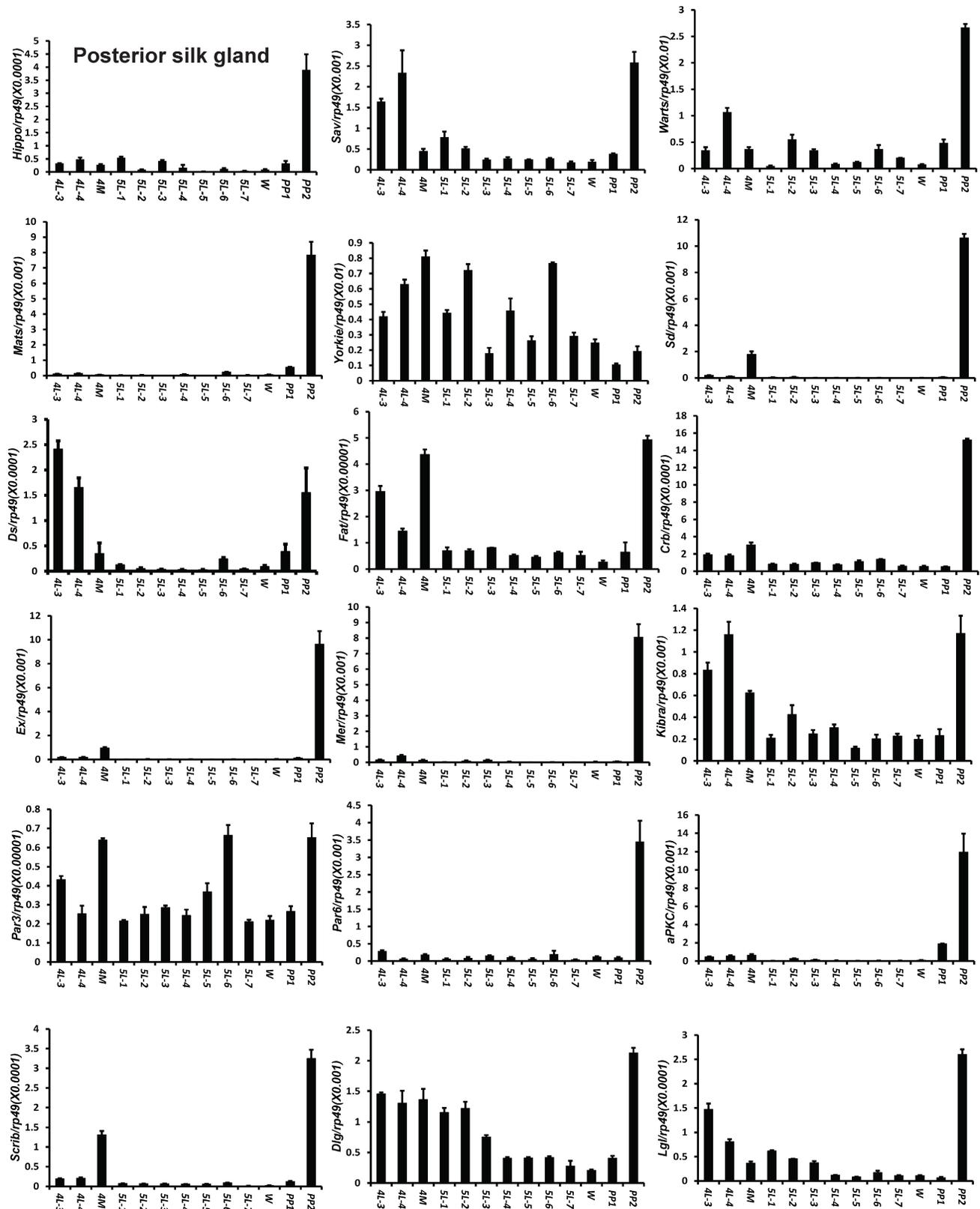


Fig. 5. Developmental expression profiles of 18 Hippo pathway genes in the posterior silk gland from day 3 of the fourth larval instar to day 2 of the prepupal stage. 4L-3 and 4L-4, day 3 and 4 of the fourth larval instar; 4M, the 4th molting; 5L-1 to 5L-7, day 1 to 7 of the fifth larval instar; W, the wandering stage; PP1 and PP2, day 1 and 2 of the prepupal stage.

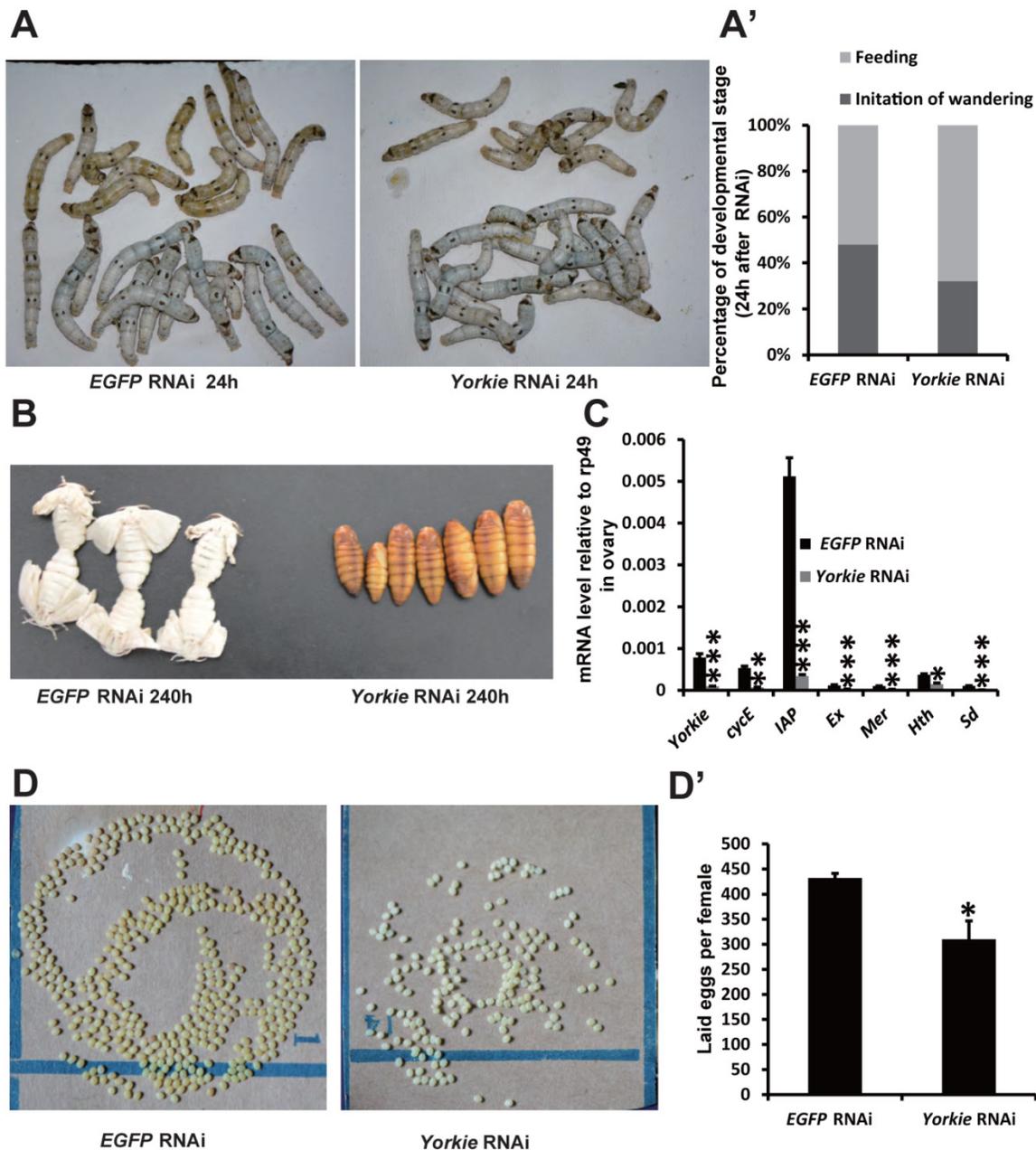


Fig.6. Yorkie RNAi delays metamorphosis and ovary maturation. (A and A') Compared to the EGFP RNAi control silkworms, the Yorkie RNAi silkworms showed delayed larval-pupal metamorphosis at 24 h after dsRNA treatment. The chart (A') shows the quantification of silkworms in (A): at the feeding stage or at the initiation of the wandering stage. (B) Compared to the EGFP RNAi control silkworms, the Yorkie RNAi silkworms showed delayed emergence at 240 h after dsRNA treatment. (C) A comparison of the mRNA levels of the Yorkie target genes in the ovary 24 h after dsRNA treatment. (D and D') Compared to the EGFP RNAi control silkworms, the Yorkie RNAi silkworm moths showed reduced egg numbers. The chart (D') shows the quantification of silkworm egg numbers in (D).

Yorkie^{CA} overexpression accelerates pupation and PSG organ size

In *Drosophila*, Yorkie overexpression induces organ overgrowth [18]. To complement the Yorkie RNAi experiments, we conducted V5-Yorkie^{CA} overexpression on L5-2 and observed the effects on *Bombyx* metamorphic development. The wandering behavior was apparently accelerated 120 h after V5-Yorkie^{CA} overexpression, compared with the RFP overexpression control animals (Figs. 7A and 7A').

Interestingly, PSG size was increased approximately 20% by V5-Yorkie^{CA} overexpression (Fig. 7B). V5-Yorkie^{CA} expression in the ovary increased by 6-fold; meanwhile, six Yorkie target genes were upregulated 3-10-fold by Yorkie overexpression (Fig. 7C). In addition, the efficiency of V5-Yorkie^{CA} overexpression in the wing disc was moderate, and Yorkie overexpression only slightly increased its organ size (Figs. S5B and S5B'). The overexpression experiment shows that Yorkie facilitates metamorphosis and promotes PSG growth in *Bombyx*.

In preliminary experiments, we found that compared to *RFP* overexpression, *V5-Yorkie* overexpression had very weak stimulatory effects on metamorphosis and PSG growth. As detected by Western blotting in the PSG using the V5 antibody, *V5-Yorkie^{CA}* overexpression yielded approximately 5-fold higher protein level than *V5-Yorkie* overexpression (Fig. 7D). As monitored by immunohistochemistry using the V5 antibody, the

overexpressed *V5-Yorkie^{CA}* mainly localized in the nuclei of the fat body cells, while the overexpressed *V5-Yorkie* localized in both nuclei and cytoplasm of the fat body cells (Fig. 7E). The more abundant protein level and the nuclear localization of the overexpressed *V5-Yorkie^{CA}* are likely responsible for its better stimulatory effects on metamorphosis and PSG growth compared to the overexpressed *V5-Yorkie*.

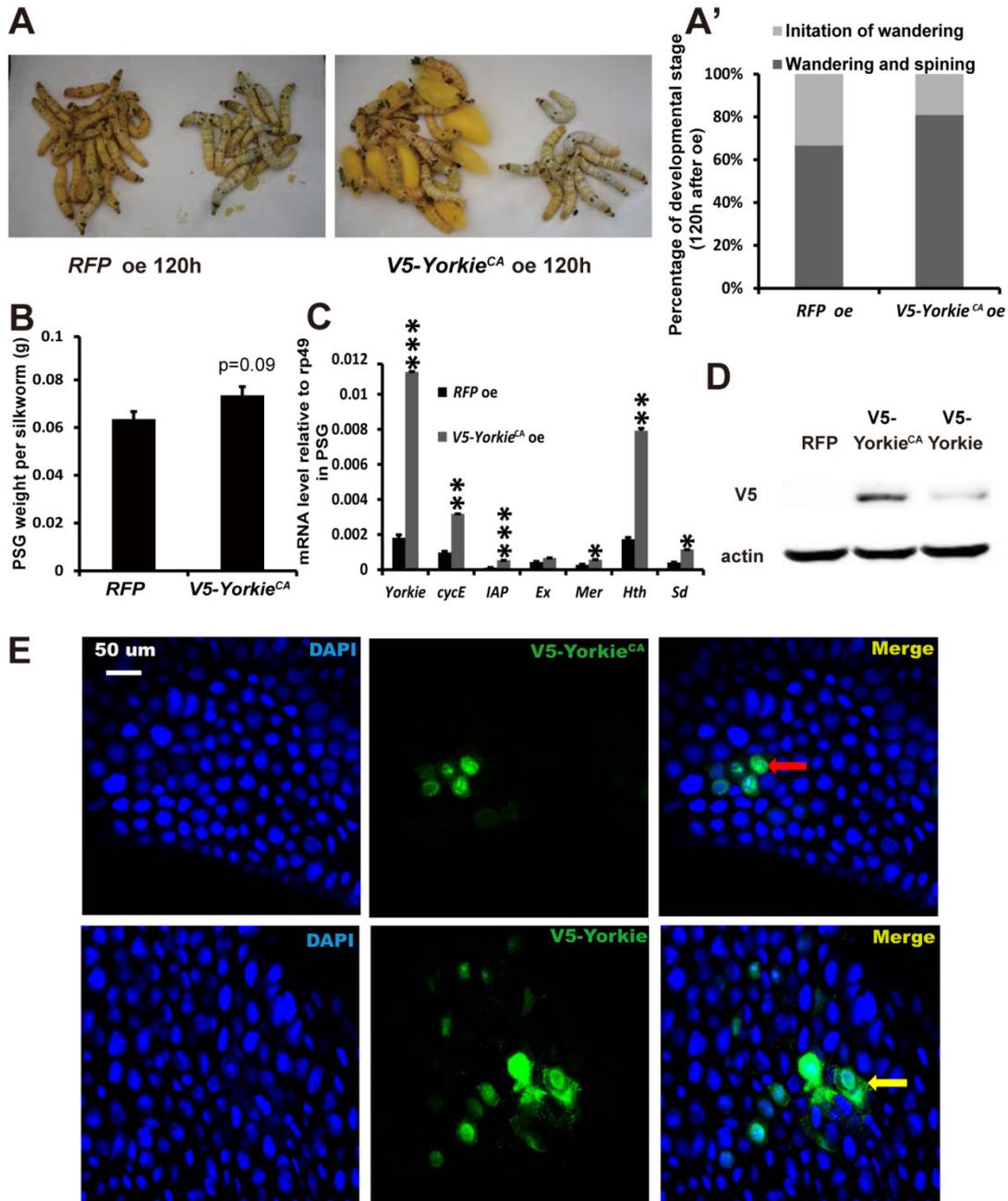


Fig. 7. *Yorkie^{CA}* overexpression accelerates pupation and PSG size. (A and A') Compared to the *RFP* overexpression (*RFP* oe) control silkworms, the *Yorkie^{CA}* overexpression (*Yorkie^{CA}* oe) silkworms showed accelerated larval-pupal metamorphosis at 120 h after baculovirus infection. The chart (A') shows the quantification of silkworms in (A): at the initiation of the wandering stage or at the wandering and spinning stages. (B) Compared to the *RFP*-overexpressed control silkworms, the *Yorkie^{CA}*-overexpressed silkworms showed size increases of the posterior silk gland (PSG) at 120 h after baculovirus infection. (C) A comparison of mRNA levels of the *Yorkie* target genes in the posterior silk gland at 120 h after baculovirus infection. (D) Western blotting analysis of the protein levels of overexpressed *V5-Yorkie^{CA}* and *V5-Yorkie* in the posterior silk gland at 120 h after baculovirus infection. The V5 antibody detects the overexpressed *V5-Yorkie^{CA}* and *V5-Yorkie* at MW ~75 kDa. (E) Immunohistochemistry analysis of the protein levels of overexpressed *V5-Yorkie^{CA}* and *V5-Yorkie* in the fat body at 120 h after baculovirus infection. As denoted by the yellow and red arrows, respectively, the overexpressed *V5-Yorkie^{CA}* always localized in the nuclei of the fat body cells, while the overexpressed *V5-Yorkie* localized in both nuclei and cytoplasm of the fat body cells.

Discussion

As revealed by microarrays and qPCR, the tissue distribution and developmental profiles of the Hippo pathway genes suggest that the Hippo pathway regulates *Bombyx* organ growth in a context-specific manner. Expression of the Hippo pathway genes on L5-3 or L5-2 is particularly enriched in several mitotic tissues, including the gonad and wing disc (Figs. S2 and 3). It is likely that the growth of mitotic tissues is restricted by the Hippo pathway at this period. In consistent with this hypothesis, during the larval stages, the mitotic tissues do not grow as dramatically as other larval tissues, such as the silk gland and the fat body that undergo rapid endocycles. During the larval stages, expression levels of the Hippo pathway genes are generally higher at the fourth instar than those at the fifth instar (Figs. 4, 5, and Fig. S4), in agreement with a dramatic body growth at the fifth instar. Moreover, as revealed by microarray analysis, expression levels of the Hippo pathway genes gradually increase from the larval stage through the pupal stage to the adult stage (Fig. S3), agreeing that the obsolete larval tissues undergo massive programmed cell death and histolysis during metamorphosis [52]. It is possible that the growth of larval tissues is tightly limited and their death is promoted by the Hippo pathway during the non-feeding pupal and adult stages. Interestingly, some Hippo pathway genes are expressed in males but not in females during the pupal stages, implying that pupal development is more restricted by the Hippo pathway in males than in females.

Pronouncedly, the developmental profiles of Hippo pathway genes are generally similar to each other (with the exception of *Yorkie*) within each tissue, but vary greatly in different tissues, including the ovary, wing disc, and PSG. It is likely that the Hippo pathway genes coordinate and control the growth of each organ, but how the pathway regulates this growth might be different in distinct organs. For example, the expression patterns of 18 Hippo pathway genes on PP2 are opposite between the wing disc and PSG (Fig. 5 and Fig. S4). On PP2, the wing disc undergoes dramatic growth [53], while the PSG stops cell proliferation and begins programmed cell death with decreased expression of *cycE* and *IAP* [54-57]. Surprisingly, expression of all genes in the Hippo kinase cassette and upstream inputs is low in the wing disc on PP2, but expression of *Yorkie* remains high (Fig. S4), suggesting that *Yorkie* might promote its dramatic growth at this developmental stage. In contrast, expression of all genes in the Hippo kinase cassette and upstream inputs sharply increases in the PSG on PP2, but expression of *Yorkie* decreases (Fig.

5), implying that a low *Yorkie* activity might allow programmed cell death and histolysis to occur in the PSG at this developmental stage. Together, the nearly opposing expression patterns of the Hippo pathway genes between the wing disc and the PSG on PP2 raise the possibility that the Hippo pathway might differently regulate growth of distinct tissues at particular developmental stages.

To understand if physiological function of the Hippo pathway is conserved in *Bombyx*, we performed both loss- and gain-of-function studies of *Yorkie*. It is necessary to note that *Yorkie* RNAi at IW did not cause lethal phenotypes. One reason could be the high expression levels of genes in the Hippo kinase cassette and upstream inputs during the pupal stages (Fig. S3) and these genes might have been lowered *Yorkie* activity during metamorphosis. Meanwhile, although the efficiency of *Yorkie* RNAi was high in the ovary, it was moderate in the wing disc and fat body but extremely low in the PSG (Fig. 6, Figs. S5, S6). The strongest phenotypes caused by *Yorkie* RNAi or *Yorkie* overexpression are changes in metamorphosis. To our understanding, this is the first time to report that *Yorkie* facilitates metamorphosis in insects. In accordance with the 20E signaling that determines *Bombyx* metamorphosis [53, 58-60], *Yorkie* promotes timely metamorphosis (Figs. 6A-6B, 7A, and 7A'). Compared with the 20E signaling that causes programmed cell death [53, 60-62], *Yorkie* promotes growth of organs, including the ovary, wing disc, and PSG (Figs. 6D, 6D', 7B, and Fig. S5). Using the newly developed CRISPR/Cas9-mediated gene knockout method [63] and the GAL4/UAS system [56], one might understand how *Yorkie* regulates organ growth and metamorphosis at the embryonic and early larval stages in *Bombyx*.

During thousands of years of artificial selection, the body size of the domesticated silkworm has been improved; moreover, the organ size of the silk gland has been even more significantly improved. During the fifth larval instar, the silk gland grows dramatically and produces a great amount of fibroin. In terms of protein synthesis, the silk gland is one of the most efficient organs in animals [42]. Using the GAL4/UAS system, we have previously demonstrated that Ras signaling, together with insulin-like peptides and nutrients, plays a critical role in the regulation of silk gland growth and silk yield [54, 56, 57]. Considering that baculovirus-mediated *Yorkie*^{CA} overexpression accelerates PSG growth (Fig. 7), we assume that *Yorkie*^{CA} overexpression in the PSG should have similar stimulatory effects on PSG growth to *Ras1*^{CA} overexpression. In order to improve silk yield more efficiently, we are considering simultaneous

overexpression of *Yorkie^{CA}* and *Ras1^{CA}* in the PSG. Combining the powerful tools of omics, genetics, and bioinformatics, we have sought to understand how the genetic signaling networks, including 20E signaling, Ras signaling and the Hippo pathway, coordinately regulate the size of the silk gland and the whole body during silkworm domestication.

Supplementary Material

Supplementary tables and figures.

<http://www.ijbs.com/v12p0917s1.pdf>

Acknowledgments

This study was supported by the National Science Foundation of China (31302034 and 31330072), the National Program for the Development of New Transgenic Species of China (2014ZX08010-016B), and Shanghai Municipal Nature Science Fund (13ZR1433100). English was polished by the Nature Publishing Group.

Authors' contributions

S Li conceived and designed the experiments. S Liu, PZ, LZ, and HQ performed the research and analyzed the data. SZ constructed the phylogenetic tree and analyzed the arthropod genomes. S Li, S Liu, and LZ wrote the paper. HS, GZ, and ZW provided important reagents. All authors read and approved the final manuscript.

Competing Interests

The authors have declared that no competing interest exists.

References

- Pan D. The Hippo signaling pathway in development and cancer. *Dev Cell*. 2010; 19:491-505.
- Staley BK, Irvine KD. Hippo signaling in *Drosophila*: recent advances and insights. *Dev Dyn*. 2012; 241:3-15.
- Tremblay AM, Camargo FD. Hippo signaling in mammalian stem cells. *Semin Cell Dev Biol*. 2012; 23:818-26.
- Enderle L, McNeill H. Hippo gains weight: added insights and complexity to pathway control. *Sci Signal*. 2013; 6:re7.
- Yu FX, Guan KL. The Hippo pathway: Regulators and regulations. *Genes Dev*. 2013; 27:355-71.
- Zhao B, Guan KL. Hippo pathway key to ploidy checkpoint. *Cell*. 2014; 158:695-6.
- Justice RW, Zilian O, Woods DF, Noll M, Bryant PJ. The *Drosophila* tumor suppressor gene warts encodes a homolog of human myotonic dystrophy kinase and is required for the control of cell shape and proliferation. *Genes Dev*. 1995; 9:534-46.
- Xu T, Wang W, Zhang S, Stewart RA, Yu W. Identifying tumor suppressors in genetic mosaics: the *Drosophila* *lats* gene encodes a putative protein kinase. *Development*. 1995; 121:1053-63.
- Kango-Singh M, Nolo R, Tao C, Verstreken P, Hiesinger PR, Bellen HJ, et al. Shar-pei mediates cell proliferation arrest during imaginal disc growth in *Drosophila*. *Development*. 2002; 129:5719-30.
- Tapon N, Harvey KF, Bell DW, Wahrer DC, Schiripo TA, Haber D, et al. Salvador Promotes both cell cycle exit and apoptosis in *Drosophila* and is mutated in human cancer cell lines. *Cell*. 2002; 110:467-78.
- Harvey KF, Pflieger CM, Hariharan IK. The *Drosophila* Mst ortholog, hippo, restricts growth and cell proliferation and promotes apoptosis. *Cell*. 2003; 114:457-67.
- Jia J, Zhang W, Wang B, Trinko R, Jiang J. The *Drosophila* Ste20 family kinase dMST functions as a tumor suppressor by restricting cell proliferation and promoting apoptosis. *Genes Dev*. 2003; 17:2514-9.

- Pantalacci S, Tapon N, Leopold P. The Salvador partner Hippo promotes apoptosis and cell-cycle exit in *Drosophila*. *Nat Cell Biol*. 2003; 5:921-7.
- Udan RS, Kango-Singh M, Nolo R, Tao C, Halder G. Hippo promotes proliferation arrest and apoptosis in the Salvador/Warts pathway. *Nat Cell Biol*. 2003; 5:914-20.
- Wu S, Huang J, Dong J, Pan D. Hippo encodes a Ste-20 family protein kinase that restricts cell proliferation and promotes apoptosis in conjunction with salvador and warts. *Cell*. 2003; 114:445-56.
- Lai ZC, Wei X, Shimizu T, Ramos E, Rohrbaugh M, Nikolaidis N, et al. Control of cell proliferation and apoptosis by mob as tumor suppressor, mats. *Cell*. 2005; 120:675-85.
- Wei X, Shimizu T, Lai ZC. Mob as tumor suppressor is activated by Hippo kinase for growth inhibition in *Drosophila*. *EMBO J*. 2007; 26:1772-81.
- Huang J, Wu S, Barrera J, Matthews K, Pan D. The Hippo signaling pathway coordinately regulates cell proliferation and apoptosis by inactivating Yorkie, the *Drosophila* Homolog of YAP. *Cell*. 2005; 122: 421-34.
- Dong J, Feldmann G, Huang J, Wu S, Zhang N, Comerford SA, et al. Elucidation of a universal size-control mechanism in *Drosophila* and mammals. *Cell*. 2007; 130:1120-33.
- Wu S, Liu Y, Zheng Y, Dong J, Pan D. The TEAD/TEF family protein Scalloped mediates transcriptional output of the Hippo growth-regulatory pathway. *Dev Cell*. 2008; 14:388-98.
- Zhang L, Ren F, Zhang Q, Chen Y, Wang B, Jiang J. The TEAD/TEF family of transcription factor Scalloped mediates Hippo signaling in organ size control. *Dev Cell*. 2008; 14:377-87.
- Goulev Y, Fauny JD, Gonzalez-Marti B, Flagiello D, Silber J, Zider A. SCALLOPED interacts with YORKIE, the nuclear effector of the hippo tumor suppressor pathway in *Drosophila*. *Curr Biol*. 2008; 18:435-41.
- Oh H, Irvine KD. In vivo regulation of Yorkie phosphorylation and localization. *Development*. 2008; 135:1081-8.
- Oh H, Irvine KD. In vivo analysis of Yorkie phosphorylation sites. *Oncogene*. 2009; 28:1916-27.
- Bennett FC, Harvey KF. Fat cadherin modulates organ size in *Drosophila* via the Salvador/Warts/Hippo signaling pathway. *Curr Biol*. 2006; 16:2101-10.
- Silva E, Tsatskis Y, Gardano L, Tapon N, McNeill H. The tumor-suppressor gene fat controls tissue growth upstream of expanded in the hippo signaling pathway. *Curr Biol*. 2006; 16:2081-9.
- Cho E, Feng Y, Rauskolb C, Maitra S, Fehon R, Irvine KD. Delineation of a Fat tumor suppressor pathway. *Nat Genet*. 2006; 38:1142-50.
- Chen CL, Gajewski KM, Hamaratoglu F, Bossuyt W, Sansores-Garcia L, Tao C, et al. The apical-basal cell polarity determinant Crumbs regulates Hippo signaling in *Drosophila*. *Proc Natl Acad Sci USA*. 2010; 107:15810-5.
- Ling C, Zheng Y, Yin F, Yu J, Huang J, Hong Y, et al. The apical transmembrane protein Crumbs functions as a tumor suppressor that regulates Hippo signaling by binding to Expanded. *Proc Natl Acad Sci USA*. 2010; 107:10532-7.
- Robinson BS, Huang J, Hong Y, Moberg KH. Crumbs regulates Salvador/Warts/Hippo signaling in *Drosophila* via the FERM-domain protein Expanded. *Curr Biol*. 2010; 20:582-90.
- Hamaratoglu F, Willecke M, Kango-Singh M, Nolo R, Hyun E, Tao C, et al. The tumour-suppressor genes NF2/Merlin and Expanded act through Hippo signalling to regulate cell proliferation and apoptosis. *Nat Cell Biol*. 2006; 8:27-36.
- Baumgartner R, Poernbacher I, Buser N, Hafen E, Stocker H. The WW domain protein Kibra acts upstream of Hippo in *Drosophila*. *Dev Cell*. 2010;18:309-16.
- Genevet A, Wehr MC, Brain R, Thompson BJ, Tapon N. Kibra is a regulator of the Salvador/Warts/Hippo signaling network. *Dev Cell*. 2010; 18:300-8.
- Yu J, Zheng Y, Dong J, Klusza S, Deng WM, Pan D. Kibra functions as a tumor suppressor protein that regulates Hippo signaling in conjunction with Merlin and Expanded. *Dev Cell*. 2010; 18:288-99.
- Grzeschik NA, Parsons LM, Allott ML, Harvey KF, Richardson HE. Lgl, aPKC, and Crumbs regulate the Salvador/Warts/Hippo pathway through two distinct mechanisms. *Curr Biol*. 2010; 20:573-81.
- Menendez J, Perez-Garijo A, Calleja M, Morata G. A tumor-suppressing mechanism in *Drosophila* involving cell competition and the Hippo pathway. *Proc Natl Acad Sci USA*. 2010; 107:14651-6.
- Sun G, Irvine KD. Regulation of Hippo signaling by Jun kinase signaling during compensatory cell proliferation and regeneration, and in neoplastic tumors. *Dev Biol*. 2011; 350:139-51.
- Yang Z, Hata Y. What is the Hippo pathway? Is the Hippo pathway conserved in *Caenorhabditis elegans*? *J Biochem*. 2013; 154:207-9.
- Hilman D, Gat U. The evolutionary history of YAP and the hippo/YAP pathway. *Mol Biol Evol*. 2011; 28:2403-17.
- Sebé-Pedrós A, Zheng Y, Ruiz-Trillo I, Pan D. Premetazoan origin of the hippo signaling pathway. *Cell Rep*. 2012; 1:13-20.
- Ikmi A, Gaertner B, Seidel C, Srivastava M, Zeitlinger J, Gibson MC. Molecular evolution of the Yap/Yorkie proto-oncogene and elucidation of its core transcriptional program. *Mol Biol Evol*. 2014; 31:1375-90.
- Xia QY, Li S, Feng QL. Advances in silkworm studies accelerated by genome sequencing of *Bombyx mori*. *Annu Rev Entomol*. 2014; 59:513-36.
- Liu S, Zhou S, Tian L, Guo E, Luan Y, Zhang J, et al. Genome-wide identification and characterization of ATP-binding cassette transporters in the silkworm, *Bombyx mori*. *BMC Genomics*. 2011; 12:491.
- Edgar RC. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucl Acids Res*. 2014; 32: 1792-97.

45. Talavera G, Castresana J. Improvement of phylogenies after removing divergent and ambiguously aligned blocks from protein sequence alignments. *Syst Biol.* 2007; 56: 564-77.
46. Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O. New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. *Syst Biol.* 2010; 59: 307-21.
47. Xia Q, Cheng D, Duan J, Wang G, Cheng T, Zha X, et al. Microarray-based gene expression profiles in multiple tissues of the domesticated silkworm, *Bombyx mori*. *Genome Biol.* 2007; 8:R162.
48. Hossain MS, Liu Y, Li K, Tian L, Li S. 20-Hydroxyecdysone-induced transcriptional activity of FoxO upregulates brummer and acid lipase-1 and promotes lipolysis in *Bombyx* fat body. *Insect Biochem Mol Biol.* 2013; 43:829-38.
49. Li K, Guo E, Hossain M, Li Q, Cao Y, Tian L, et al. *Bombyx* E75 isoforms display stage- and tissue-specific response to 20-hydroxyecdysone. *Sci Rep.* 2015; 5:12114.
50. Misof B, Liu S, Meusemann K, Peters RS, Donath A, Mayer C, et al. Phylogenomics resolves the timing and pattern of insect evolution. *Science.* 2014; 346:763-7.
51. Qi HS, Liu SM, Li S, Wei ZJ. Molecular expression of the Scribble complex genes, *Dlg*, *Scrib* and *Lgl*, in silkworm, *Bombyx mori*. *Genes (Basel).* 2013; 4:264-74.
52. Yin VP, Thummel CS. Mechanisms of steroid-triggered programmed cell death in *Drosophila*. *Semin Cell Dev Biol.* 2005; 16:237-43.
53. Liu X, Dai F, Guo E, Li K, Ma L, Tian L, et al. E93 transcriptionally modulates 20-hydroxyecdysone signaling during *Bombyx* larval-pupal metamorphosis. *J Biol Chem.* 2015; 290:27370-83.
54. Ma L, Liu S, Shi M, Chen XX, Li S. Ras1^{CA}-upregulated *bcp1* inhibits cathepsin activity to prevent tissue destruction of the *Bombyx* posterior silk gland. *J Proteome Res.* 2013; 12:1924-34.
55. Jia SH, Li MW, Zhou B, Liu WB, Zhang Y, Miao XX, et al. Proteomic analysis of silk gland programmed cell death during metamorphosis of the silkworm, *Bombyx mori*. *J Proteome Res.* 2007; 6:3003-10.
56. Ma L, Xu H, Zhu J, Ma S, Liu Y, Jiang RJ, et al. Ras1^{CA} overexpression in the posterior silk gland improves silk yield. *Cell Res.* 2011; 21:934-43.
57. Ma L, Ma Q, Li X, Cheng L, Li K, Li S. Transcriptomic analysis of differentially expressed genes in the Ras1^{CA}-overexpressed and wildtype posterior silk glands. *BMC Genomics.* 2014; 15:182.
58. Cheng D, Xia Q, Duan J, Wei L, Huang C, Li Z, et al. Nuclear receptors in *Bombyx mori*: insights into genomic structure and developmental expression. *Insect Biochem Mol Biol.* 2008; 38:1130-7.
59. Tian L, Guo E, Diao Y, Wang S, Liu S, Jiang RJ, et al. Developmental regulation of glycolysis by 20-hydroxyecdysone and juvenile hormone in fat body tissues of the silkworm, *Bombyx mori*. *J Mol Cell Biol.* 2010; 2:255-63.
60. Guo E, He Q, Liu S, Tian L, Sheng Z, Peng Q, et al. MET is required for the maximal action of 20-hydroxyecdysone during *Bombyx* metamorphosis. *PLoS ONE.* 2012; 7:e53256.
61. Tian L, Liu SM, Liu H, Li S. 20-hydroxyecdysone upregulates apoptotic genes and induces apoptosis in the *Bombyx* fat body. *Arch Insect Biochem Physiol.* 2012; 79:207-19.
62. Tian L, Ma L, Guo E, Deng X, Ma S, Xia Q, et al. 20-hydroxyecdysone upregulates Atg genes to induce autophagy in the *Bombyx* fat body. *Autophagy.* 2013; 9:1172-87.
63. Wang Y, Li Z, Xu J, Zeng B, Ling L, You L, et al. The CRISPR/Cas system mediates efficient genome engineering in *Bombyx mori*. *Cell Res.* 2013; 23:1414-6.