

Research Paper

Targeting ER α degradation by L-Tetrahydropalmatine provides a novel strategy for breast cancer treatment

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Abstract

The incidence and mortality of breast cancer (BCa) are the highest among female cancers. There are approximate 70% BCa that are classified as estrogen receptor alpha (ER α) positive. Therefore, targeting ER α is the most significantly therapeutic schedule. However, patients with breast cancer develop resistance to ER α or estrogen (E2) antagonists such as fulvestrant and tamoxifen. In the present study, we found that L-Tetrahydropalmatine (L-THP) significantly suppressed cell proliferation in ER α ⁺ BCa cells via inducing cell cycle arrest rather than apoptosis. Additionally, L-THP enhanced the sensitivity of ER α ⁺ BCa cells to tamoxifen and fulvestrant. Mechanically, the application of L-THP promotes ER α degradation through accumulating ubiquitin chains on ER α . Overexpressing ER α abrogates L-THP induced-antiproliferation in ER α ⁺ BCa cells. Collectively, our work indicates that L-THP may represent a potentially novel therapeutic medicine for ER α ⁺ breast cancer patient.

Key words: breast cancer, L-THP, ER α , proliferation, cell cycle

Introduction

The incidence and mortality of breast cancer (BCa) have been studied to be the No.1 in female all over the world, resulting in seeking some more effective therapeutic schedules is an urgent matter [1]. BCa is divided into many subtypes basing on different criteria, including in tumor size, lymph nodes, molecular subtypes classification and so on [2]. Among these characteristic, molecular subtype is considered to be classical and significant advance. Based on hormone receptor status, breast cancer is divided into estrogen receptor, human growth factor, progesterone receptor and triple negative tumor [3]. It has been demonstrated nearly 80 years ago that there are a closed relationship between breast cancer and

estrogen receptor signaling [4].

Estrogen binds to receptor and activates the estrogen receptor (ER) signaling. The ER is considered as a transcription factor of nuclear receptor and owns two subtypes, including ER α and ER β [5]. ER α is expressed in approximately 70% breast cancer and has a draving and proliferative potential in breast cancer [6]. Overexpressing ER α promotes the G0/G1 to S phase transition by increasing protein expression of p21 and inhibiting Cyclin D1, which are essential factors for cell cycle progression [7, 8]. Clinically, ER α expression is correlated with the prognosis of breast cancer patients and thus is regarded as an important target to endocrine therapy [9]. Some inhibitors are

developed to target ER α . For example, tamoxifen and fulvestrant, inhibitors of antiestrogen, have been applied for ER α positive breast cancer patients and bring a significant prognostic [10, 11]. However, with the disease progression, acquired resistance to the endocrine therapy has been developed in many patients who received tamoxifen and fulvestrant treatment [12]. Thus, developing alternatively new agents is necessary to win the battle against BCa in future.

Rotundine (L-Tetrahydropalmatine, L-THP) is a natural tetrahydro protoberberine isoquinoline alkaloid isolated from *stephania and corydalis* [13]. Clinically, L-THP is traditional Chinese medicine for 40 years and used for the treatment of cardiovascular and a lot of pains [14, 15]. Studies have been demonstrated that L-THP shows a protective role in cardio, blood vessels and neural tissues via anti-apoptosis, anti-oxidant and anti-inflammation [16-18]. As previously reported, natural alkaloids have also been shown their brilliant anti-cancer activities in some human cancers, such as leukemia and prostate cancer [19, 20]. The alkaloids can induce cell apoptosis and cell cycle arrest by interacting with DNA in vitro or inhibiting G2/M transition [20, 21]. Considering that L-THP is an isoquinoline alkaloid, some researchers explore the potential effect against cancers. L-THP plays an antiproliferative role in breast cancer MCF-7 cell line through regulating the uptake of ⁹⁹Tc^m-MIBI [22]. Moreover, L-THP has been indicated to enhance the sensitivity of leukemia cell to doxorubicin and in combination of L-THP and berberine triggers anti-growth in MDA-MB231 breast cancer cell [23, 24].

In this study, L-Tetrahydropalmatine, as a natural product, inhibits the growth and progression of ER α positive breast cancer cells by cell cycle arrest. Mechanistically, we demonstrate that L-THP downregulated ER α protein via increasing the ubiquitination on ER α . Moreover, the transcriptional activity of ER α and its downstream signaling pathway are significantly suppressed by L-THP. Additionally, the application of L-THP enhances the anti-cancer effects of tamoxifen and fulvestrant on ER α positive breast cancer cells. Collectively, these results indicate that L-THP as a clinically painkiller could be applied to treat breast cancer via regulating ER α degradation and transcriptional activity.

Materials and methods

Materials

Rotundine (S2437), tamoxifen (S1238) and fulvestrant (S1191) were obtained from Sellckchem (Houston, TX, USA). Antibodies are as follows:

anti-ER α (#8644), anti-PARP (#9532), anti-p27 (#3686), anti-Rb (#9309), anti-p-Rb (#8516), anti-CDK4 (#12790), anti-p27 (#3686), anti-Cyclin D1 (#2922), anti-Bcl-2 (#15071), anti-GAPDH (#5174), anti-ubiquitin (#3936), anti-K48-Ub (#12805). All of these antibodies above were purchased from Cell Signaling Technology (MA, USA).

Cell culture

MCF-7, BT474, T47D, MDA-MB453 and MDA-MB468 cells were purchased from ATCC (American Type Culture Collection, Manassas, VA, USA), grown in DMEM contained with 10% FBS. MCF-7 needs add 10 ug/ml insulin. 5% CO₂ and 95% air were kept at 37°C for cell culture.

Cell viability

MTS (catalog no. G111) was applied for cell viability analysis. This reagent was from Promega Corporation (Madison, WI, USA). As previously reported in our studies [25, 26], cells were grown in 96-well plates for 16-24 h. Adherent cells were treated with L-THP at the indicated doses for different times, followed by 20 μ l MTS for 2 or 3 h.

Edu staining

The assay was worked as we reported previously [27, 28] using cell-light EdU Apollo 567 in vitro kit (Cat number: C10310-1, RiboBio, Guangzhou, China). MCF-7 and T47D cells were plated onto chamber slides. The indicate treatment was performed. Edu reagents were incubated for the additional 2 h. According to the manufacturer's instructions, cells were fixed, then incubated with reaction buffer. DAPI for cell nucleus staining.

Cell cycle and apoptosis

Cells were placed into 6-well plates and then performed treatment of L-THP. Cells were digested and washed with PBS. As previously describe in our studies [29, 30], for cell cycle assay, the mixture containing 500 μ l PBS and 2000 μ l 70% ethanol were used to fix cell for one night. The stained reagents were applied in dark for 30 min, including 50 μ g/ml of PI, 0.2% of Triton-X-100 and 100 μ g/ml of RNase. For cell apoptosis, 500 μ l binding buffer with 5 μ l Annexin V-FITC and PI was used to resuspended. The kit for apoptosis was from Keygen Company (Nanjing, China). Lastly, cells were subjected to flow cytometry.

Cell clone formation

BCa cells were plated and exposed to the indicated treatment. The treated cells were cultured into six-well plates until colonies formation. The cells were fixed and stained with crystal violet for 15 min.

The images were captured by camera.

Western blotting and Co-IP

To prepare protein for the two assays, cells were harvested after treatment of L-THP using lysis buffer (PMSF, protein inhibitor). In our previously describe [31], The concentration of protein was measured to the same. For Co-IP, the assay was performed to explore the interaction between the target proteins. Dynabeads were incubated with indicated antibody for 16-24h. Cell lysates were then added into antibody-conjugated dynabeads, shaking incubated for 1-2h. The kit was used to wash the immunocomplexes following the manufacturer's instructions, then PBST washes for thrice. Immunoprecipitants and protein lysates were subjected to western blot. Proteins were fractionated by SDS-PAGE and then transferred to a membrane. The membranes were incubated with the indicated antibodies including primary and secondary antibodies. The protein expression was reflected using X-ray films (Kodak, Japan).

RT-qPCR

RNA was extracted using RNAiso plus (TakaRa Biotechnology, Dalian, China).

Dual-luciferase reporter promoter. The same concentration cDNAs were measure in each sample and then quantitative PCR in triplicate was performed using SYBR Green Dye. The amplification was obtained to analysis the expression of targeted genes. GAPDH was as a normal level. The primers of genes were as follows: human ER α -F: 5' TCTTGGACAGGAACCAGGGA 3', human ER α -R: 5' CAGAGACTTCAGGGTGCTGG 3'; human PS2-F: 5'-TTGTGGTTTTCTGGIGTCA-3', human PS2-R: 5'-GCAGATCCCTGCAGAAAGTGT-3; human Cyclin D1-F:5'-GCTGCGAAGTGGAAACCATC-3', human Cyclin D1-R:5'-CCTCCTTCTGCACACATTGAA-3'; human GAPDH-F 5'TCCCATCACCATCTTCCA3', human GAPDH-R 5' CATCACGCCACAGTTTCC3'.

Confocal

As previously described immunofluorescence staining were done [32, 33]. In brief, the treated cells were fixed and washed with PBS twice time followed by permeabilization with 0.1% Triton X-100 for 15 min. Then primary antibody was used to incubated with cells overnight at 4 °C, followed by second antibody for 1 h. DAPI was applied for nuclei.

Statistical analysis

The showed data are as mean SD from three independent experiments. Unpaired Students's t test is applied where appropriate to analysis the statistical probabilities. GraphPad Prism8.0 software and SPSS

16.0 are used to perform statistical test. P value of < 0.05 is judged statistically significant.

Results

L-THP inhibits the growth of ER α ⁺ breast cancer cell lines

L-THP is traditional Chinese medicine and mainly used for the treatment of cardiovascular and a lot of pains. Recently, increasing studies have shown that L-THP displays anti-proliferative effects on several cancer models, such as leukemia and ovarian cancer. But the anti-cancer activity of L-THP on ER α ⁺ breast cancer cell lines has not been reported. To detect the anti-tumor effect on BCa, we choosed the three ER α ⁺ breast cancer cell lines, including MCF-7, T47D and BT474 cells. Firstly, we used cell viability assay to evaluate the proliferation of BCa cells in the treatment of escalating doses of L-THP. The value of OD indicated that L-THP significantly suppressed the growth on ER α ⁺ breast cancer cell lines (Figure 1A). Secondly, we also test effect of L-THP on ER α negative breast cancer cell lines (MDA-MB453 and MDA-MB468). MTS results showed that cell growth was suppressed by L-THP on ER α negative BC cells and it has been shown in Figure S1A. This clarify that L-THP may target other molecular, in addition to ER α . To further evaluate the function of ER α on inhibitory effect of L-THP on ER α ⁺ breast cancer cells. We calculate the IC₅₀ at 48 h on each cell lines. The IC₅₀ is lower in ER α ⁺ breast cancer cells than in ER α ⁻ breast cancer cells (Figure S1B), suggesting ER α ⁺ breast cancer cells is more sensitivity to L-THP treatment. ER α plays a significant role on L-THP's inhibitory effect. L-THP suppresses BCa cell growth depending on ER α , but just not only ER α . To determine the long effect of L-THP on BCa cell lines, colony formation assay was performed. The results showed that L-THP decreased the cloning ability of BCa (Figure 1B, C). Moreover, we further explored the cell growth using Edu staining which is a thymidine analog binding to replicated chromosomal DNA. The stained cells were decreased in the treated group of L-THP (Figure 1D, E). These findings support an inhibitory role of L-THP on ER α ⁺ BCa cells.

Anti-proliferative ability of L-THP depends on cell cycle arrest on ER α ⁺ BCa cells

We have demonstrated that L-THP inhibited the growth on ER α ⁺ BCa cells. To study the mechanism of anti-cancer of L-THP, we considered two main aspects to explore, including cell cycle arrest and cell apoptosis. Firstly, we did the cell cycle analysis to evaluate the cell cycle distributions in present of L-THP in MCF-7 and T47D cells. The cell number was

upregulated in G0/G1 phases after the treatment of L-THP, clarifying L-THP inhibited G1 to S transition in ERα⁺ BCa cells (Figure 2A, B). Furthermore, related molecular mechanisms were explored. We detected the protein expressions of p27, CDK4, Cyclin D1, Rb and p-Rb using western blotting. The expression of p27 was increased. CDK4, Cyclin D1, Rb and p-Rb were downregulated (Figure 2C). The results indicated L-THP inhibited protein expression promoting G1-S transition and upregulated protein expression suppressing transition. On the other aspect, we speculated that cell apoptosis could be induced by L-THP. Flow cytometry assay was applied and showed no apoptotic cells in the treatment of L-THP in MCF-7 cells (Figure 2D, E). The expressions of pro-apoptotic and anti-apoptotic

proteins have no change by L-THP treatment (Figure 2F). The results were consistent with the flow cytometry. Therefore, we suggest that L-THP induced inhibition of cell growth depends on cell cycle arrest rather than cell apoptosis.

L-THP induces the downregulation of ERα protein

Given that inhibitory effect of L-THP on ERα positive breast cancer cells, we assessed the role of L-THP on the ERα expression. Western blotting assay showed that the expression of ERα protein was decreased in MCF-7 and T47D cells (Figure 3A, B). ERα is a transcriptional factor. Estrogen (E2) can binds to ERα and then activates the transcriptional activity. E2 can promote the degradation of ERα. We sought to

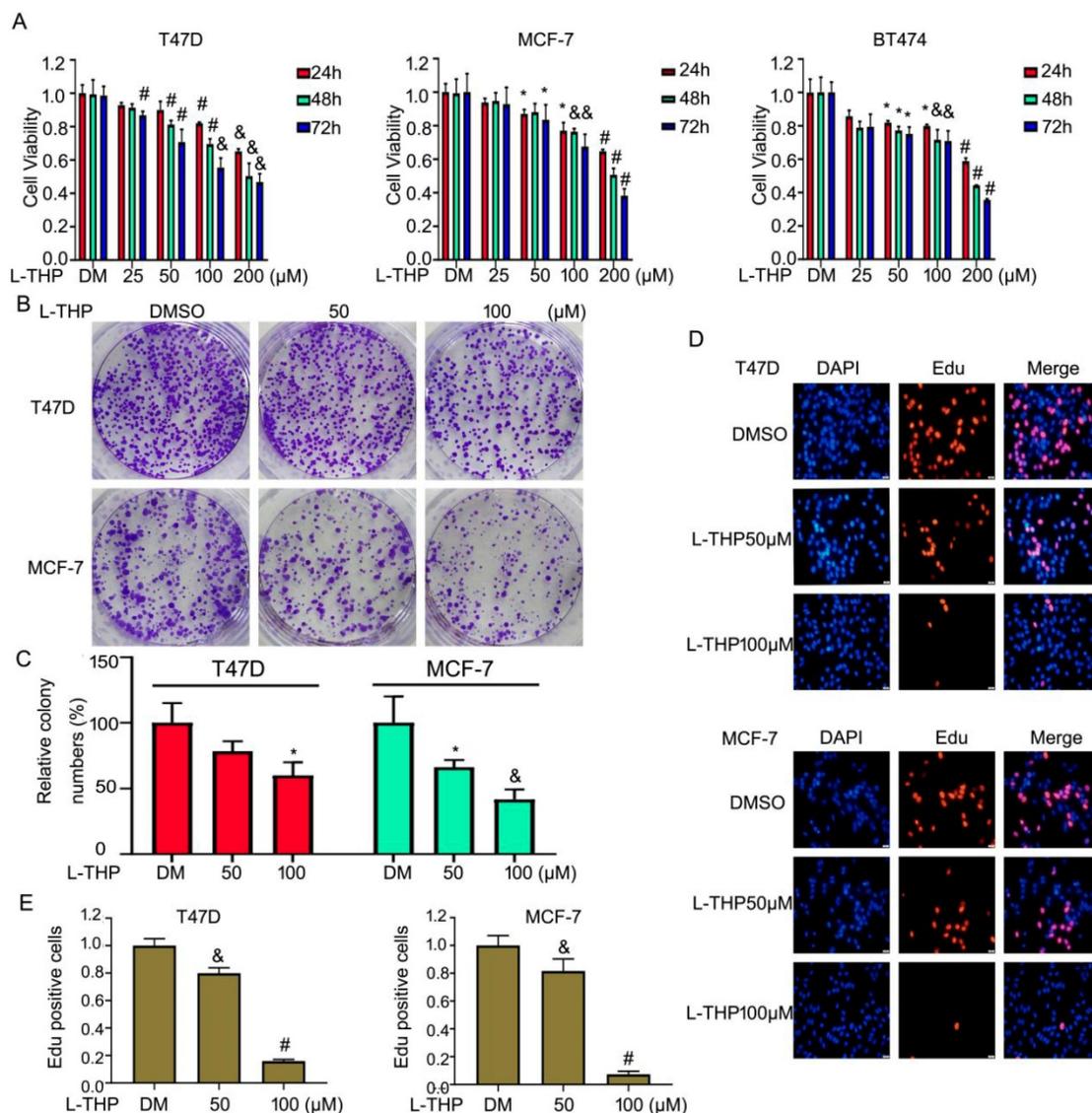


Figure 1. L-THP inhibits the growth of ERα⁺ breast cancer cell lines. (A) Cell viability was performed in MCF-7, T47D and BT474 cells post different concentrations of L-THP treatment for 24, 48 and 72h. (B) Colony formation assay was performed in BCa cells post L-THP for various concentrations for 10-14 days. (C) The colony numbers have been calculated. (D) Edu staining assay was performed in MCF-7 and T47D cells treated with L-THP for 48 h. (E) Counting stained cells which is in proliferation in each group. *p<0.05, *p<0.01, #p<0.001 versus each vehicle control. DM: DMSO.

explore the effect of L-THP in the present of E2 on the expression of ER α protein. The decreased expression of ER α protein was more obvious induced by L-THP after adding the treatment of E2 (Figure 3C, D). Furthermore, we speculated whether L-THP can regulate co-translated in breast cancer cells. We

applied the confocal microscope to observe the expression in the cytoplasm and nuclear. The images showed that L-THP significantly reduced the abundance of ER α . However, the translocate of ER α did not happen in the treatment of L-THP in MCF-7 and T47D cells (Figure 3E, F).

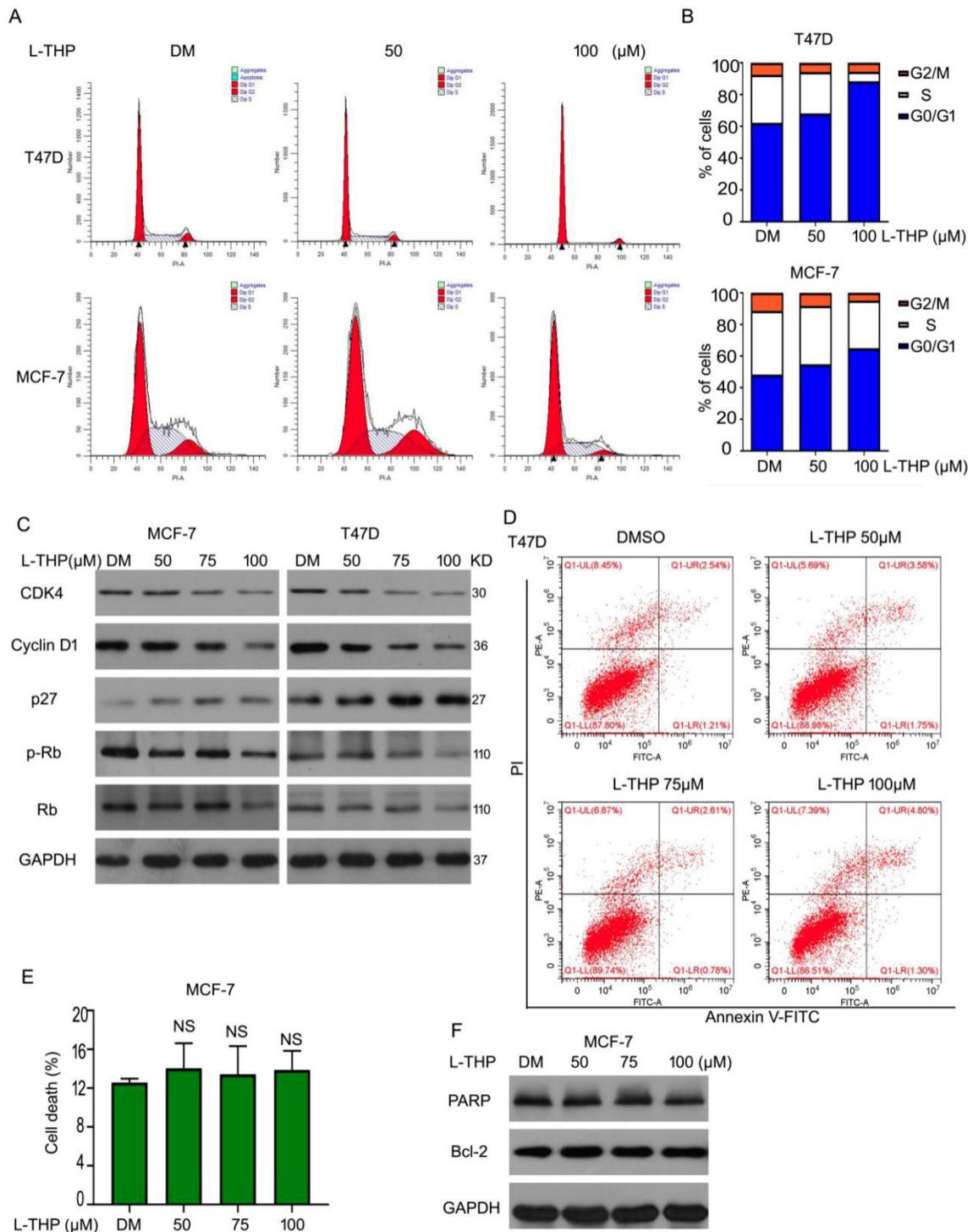


Figure 2. Anti-proliferative ability of L-THP depends on cell cycle arrest on ER α + BCa cells. (A, B) The cells with L-THP treatment for 48 h were subjected to fluorescence-activated cell sorting analysis (FACS) for cell cycle distributions. (C) Cells were treated with L-THP (50, 75, 100 μ M) for 48 h. And then proteins were collected for western blot assay to test the expression of CDK4, Cyclin D1, p27, Rb, p-Rb. (D) Apoptosis assay was performed on MCF-7 cells posted with L-THP treatment and (E) showed are pooled data. NS is no significant. (F) Western blot assay was performed for expression of PARP and Bcl-2.

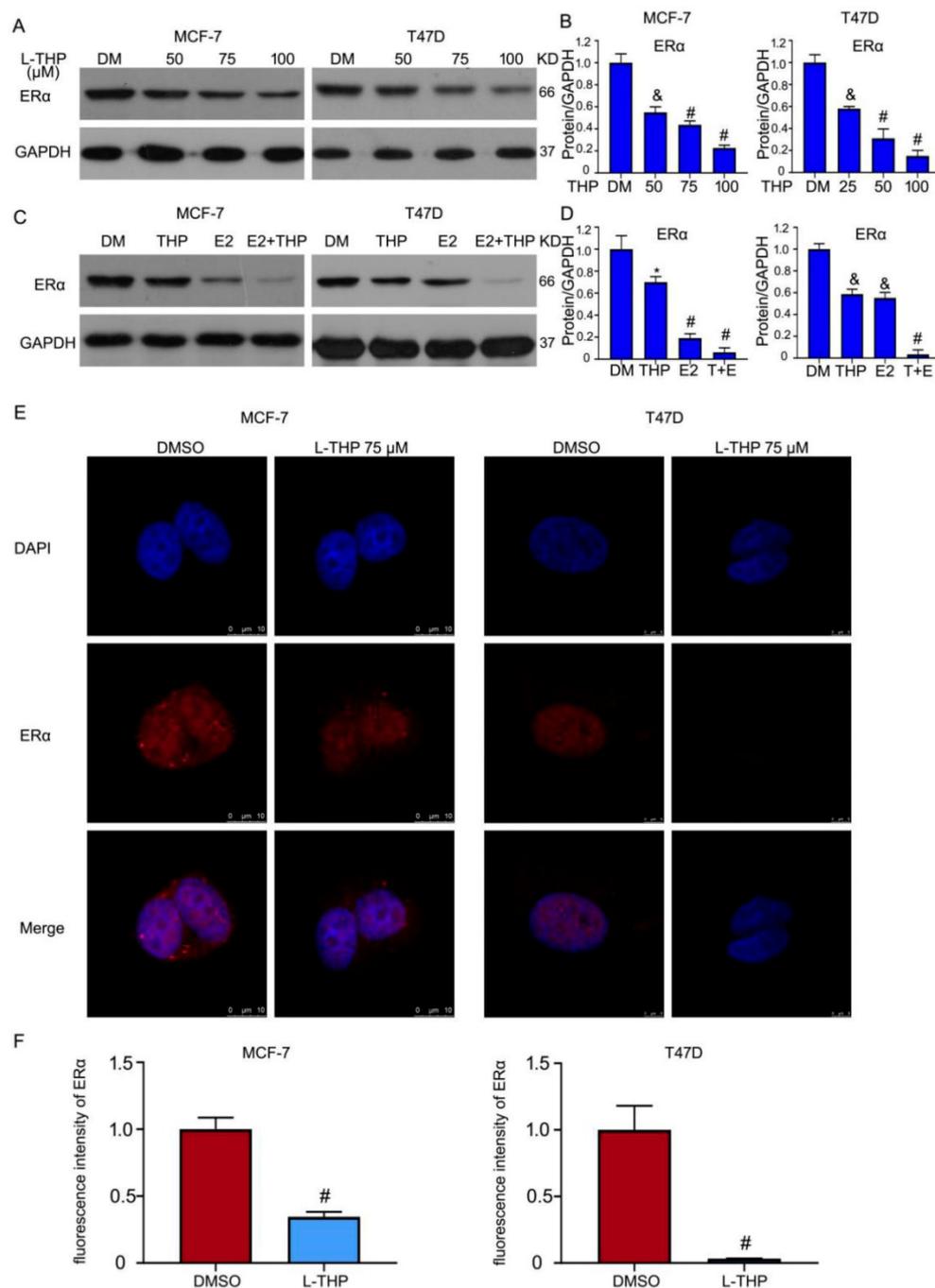


Figure 3. L-THP induces the downregulation of ER α protein. (A, C) Western blot assay was done on BCa cells posted with L-THP or estrogen (E2) treatment for 48 h. (B, D) The expressions of ER α were quantified. (E) MCF-7 and T47D cells exposed to L-THP (THP) (75 μ M) for 48 h were subjected to immunofluorescence assay. Images were captured by confocal microscopy. (F) The quantification of fluorescence intensity of ER α from three images were performed. * p <0.05, * p <0.01, # p <0.001 versus each vehicle control.

L-THP decreases the expression of ER α protein resulting from promoting its degradation

We have demonstrated that the protein expression of ER α can be influenced by L-THP. As we all known, the protein expression is from translation and transcription methods. Next, we further explored which results in L-THP induced-decreased ER α expression between translation and transcription

levels. RT-qPCR was applied to test the mRNA level of ER α . L-THP did not induce a significant change on the mRNA level of ER α , but the downstream genes of ER α were regulated by L-THP (Figure 4A). Considering the stability mRNA level of ER α , we speculated that L-THP modulated the transcription level. Cycloheximide (CHX) was used to inhibit protein synthesis. Under the treatment of CHX, L-THP accelerated the decrease of ER α protein,

indicating L-THP inhibits the transcription of ERα (Figure 4B, C). In addition, in the E2 present, the downregulated speed of ERα protein was more rapid in the L-THP group (Figure 4D, E). Obtaining that L-THP inhibited the protein expression of ERα via suppressing protein synthesis, we further explored

how L-THP promotes the degradation of ERα protein. Co-IP results showed that L-THP increased the poly- and K48-ubiquitination levels on ERα (Figure 4F). Moreover, ERα is a transcriptional factor. The transcriptional activity of ERα was reduced by L-THP (Figure 4G).

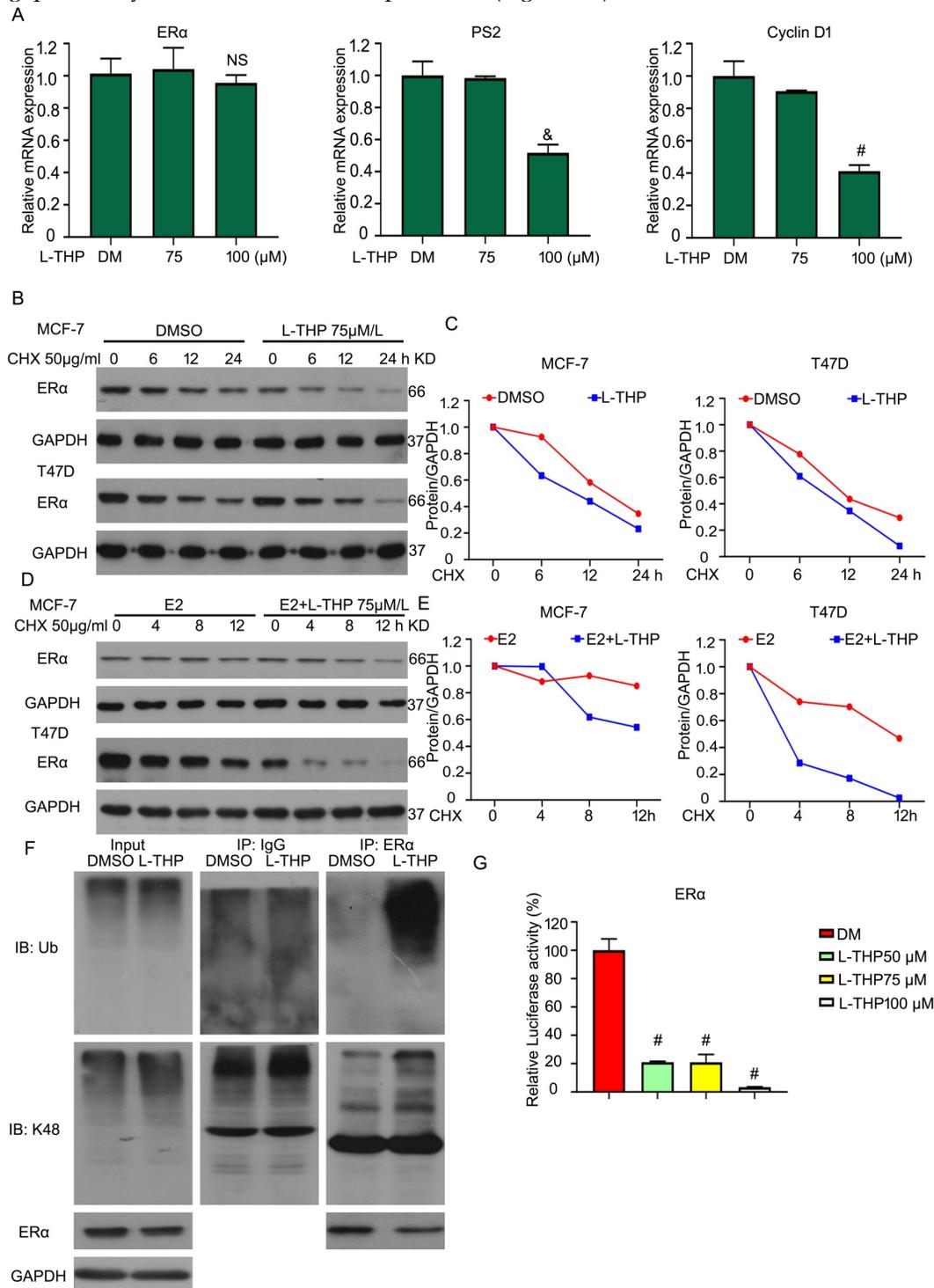


Figure 4. L-THP decreases the expression of ERα protein resulting from promoting its degradation. (A) The total RNAs were objected from T47D cells. RT-qPCR assay was performed for the expressions of ERα, Cyclin D1 and PS2 genes. (B) Proteins obtained from BCa cells exposed to CHX or CHX+L-THP treatment were subjected to western blot assay for ERα expression. (D) Western blot assay was performed on BCa cells treated with CHX+E2, CHX+E2+L-THP treatments for ERα. (C, E) The expressions of ERα were quantified. (F) BCa cells were treated with L-THP treatment for 72 h. Immunoprecipitated with ERα beads was subjected to immunoblotted for Ub and K48 expression. (G) Dual-luciferase assay was performed to detect transcriptional activity of ERα. *p<0.01, #p<0.001 versus each vehicle control.

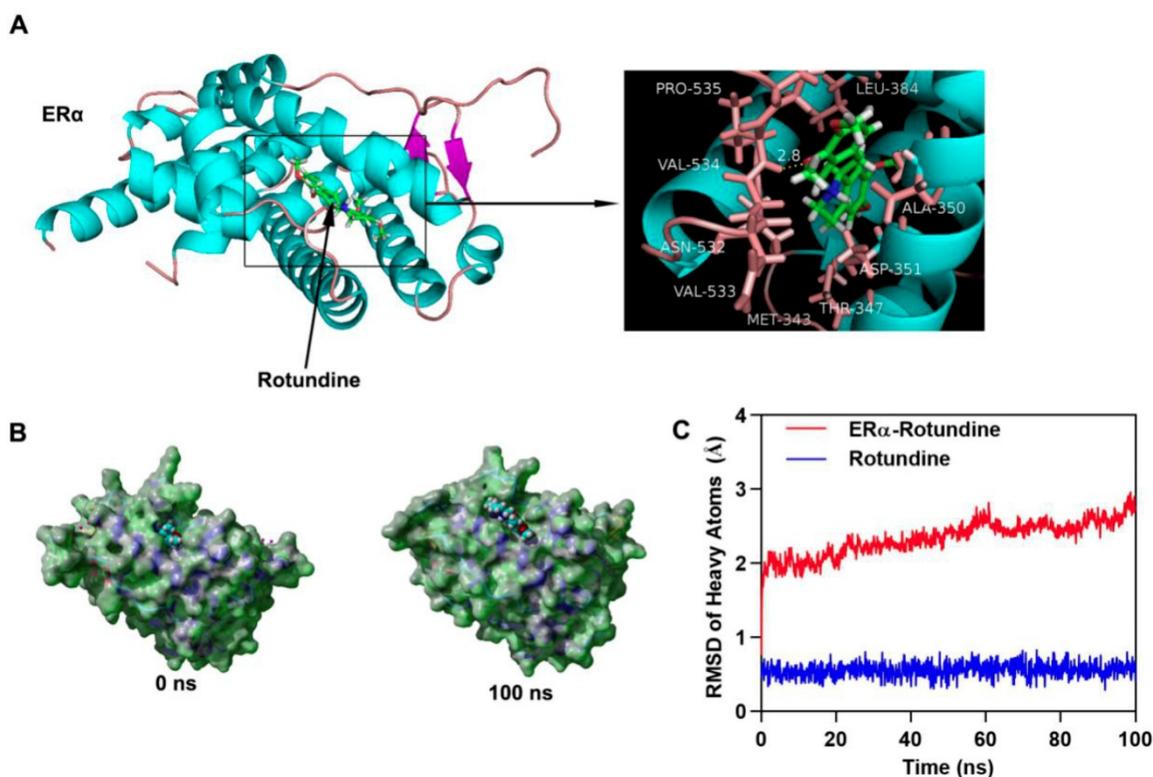


Figure 5. L-THP interacts with ER α . (A) Three dimensional crystal structure of L-THP (Rotundine) in complex with ER α (PDB ID: 5FQT). L-THP is shown in green, and the hydrogen bonds are indicated by the yellow line. (B) Surface presentation of the ER α -L-THP complex crystal structure at 0 ns and 100 ns. (C) Plots of root mean square deviation (RMSD) of heavy atoms of ER α (red) and L-THP (Rotundine) (blue) in ER α -L-THP (Rotundine) complex.

L-THP interacts with ER α

We have demonstrated the participation of L-THP on ER α protein expression. To further study how L-THP increases the ubiquitination level on ER α , we observed whether exists interaction between ER α and L-THP. We performed molecular docking by using Autodock vina. The binding energy of ER α -Rotundine complex was -7.509 kcal/mol. In Figure 5A has showed that the three dimensional and two-dimensional binding conformation of ER α -Rotundine complex is observed. We found a hydrogen bond was formed between VAL-534 of ER α and L-THP. Between ER α and L-THP, the distance of hydrogen bond was detected at 2.8 Å. It was also observed that L-THP interacted with PRO-535, VAL533, ASN-532, LEU-384, ALA-350, ASP-351, THR-347, and MET-343 *via* van der Waals force. The surface models of ER α -Rotundine complex are visualized and presented (Figure 5B). L-THP steadily showed at the center of ER α binding site until the end of MD simulation. The evolution of heave atoms root-mean-square deviation (RMSD) of the complex concerning the minimized structure. The heave atoms RMSD track of ER α in ER α -L-THP complex rose from 0.6 Å to 2 Å during the first 5 ns, fluctuated around 2 Å during 5 to 20 ns, and then the RMSD rose from 2 Å to 2.5 Å during 20 to 60 ns, then fluctuated around 2.5 Å during last 40 ns

(Figure 5C, red line). The heavy atoms RMSD track of Rotundine in ER α -Rotundine fluctuated around 0.6 Å during entire MD simulations (Figure 5C, blue line). These results suggest a strong binding between the kinase domain of ER α and Rotundine, indicating that Rotundine could directly target ER α .

ER α overexpression abrogates partly growth inhibition induced by L-THP

Given that L-THP inhibits cell growth and ER α expression in protein level in breast cancer cells, we lastly asked whether L-THP induced cell inhibition resulted from decreased ER α protein. To detect this idea, we purchased the plasmid of human ER α and transduced into T47D and MCF-7 cells. We found that overexpressing ER α protein promoted cell cycle progression. Importantly, under full-length plasmid of ER α treatment, cell cycle arrest induced by L-THP weaken in ER α positive breast cancer cells (Figure 6A, B). What is more, we used western blot to test the ER α and FLAG expression to ensure the plasmid into cells. The expression of two proteins were remarkably overexpressed. Rb and p-Rb expression were evaluated using western blot. The FLAG-ER α rescued the downregulation induced by L-THP. These results suggested that L-THP-inhibited breast cancer cell growth depends on ER α status (Figure 6C).

L-THP increases the sensitivity of BCa cells to tamoxifen

Considering the role of L-THP on ER α expression and the growth in breast cancer cells, we hypothesized whether L-THP increases the effect of inhibitor targeting ER α in ER α positive BCa cells. Tamoxifen, as an antagonist of estrogen, binds to estrogen receptor. Cell viability assay were used to measure proliferation under the treatment of L-THP. We found that L-THP enhanced the inhibitory ability of tamoxifen in MCF-7 and T47D cells (Figure 7A). The long-proliferation also was suppressed by L-THP or tamoxifen. The more anti-proliferation was happened in the combination of the two drugs (Figure 7B, C). Additionally, we explore the molecular regulation, such as ER α and its targeted protein using western blotting assay. The result showed that the decreased expression of ER α and Cyclin D1 proteins were more in the group of L-THP + tamoxifen than the signal treatment (Figure 7D). In previous finding, we have demonstrated that L-THP did not induce cell apoptosis. Hence, we seek to explore whether L-THP enhanced apoptosis induced by tamoxifen in ER α BCa cells. Flow cytometry and western blotting assay

clarified that cell apoptosis was increased by adding L-THP treatment (Figure 7E-G).

L-THP and fulvestrant synergistically suppress the growth of BCa cells

Unlike tamoxifen, Fulvestrant is an antagonist of estrogen receptor that blocking the binding to estrogen to inhibit its receptor and changing receptor morphological to decrease its concentration. As the same, to better investigate the effect of L-THP on ER α protein, we test the joint effect between L-THP and fulvestrant in ER α positive breast cancer cells. We used MTS assay to evaluate cell growth. We found that L-THP enhanced the anti-proliferative ability of fulvestrant in breast cancer cells (Figure 8A). The expression of ER α and Cyclin D1 proteins were reduced when L-THP in combination with fulvestrant than the signal treatment (Figure 8B). In addition, the colony formation results also showed that effect of the combination of L-THP and fulvestrant was more significant (Figure 8C, D). We have performed the Edu staining assay and found that L-THP enhanced the sensitivity of ER α positive BCa cells to fulvestrant (Figure 8E).

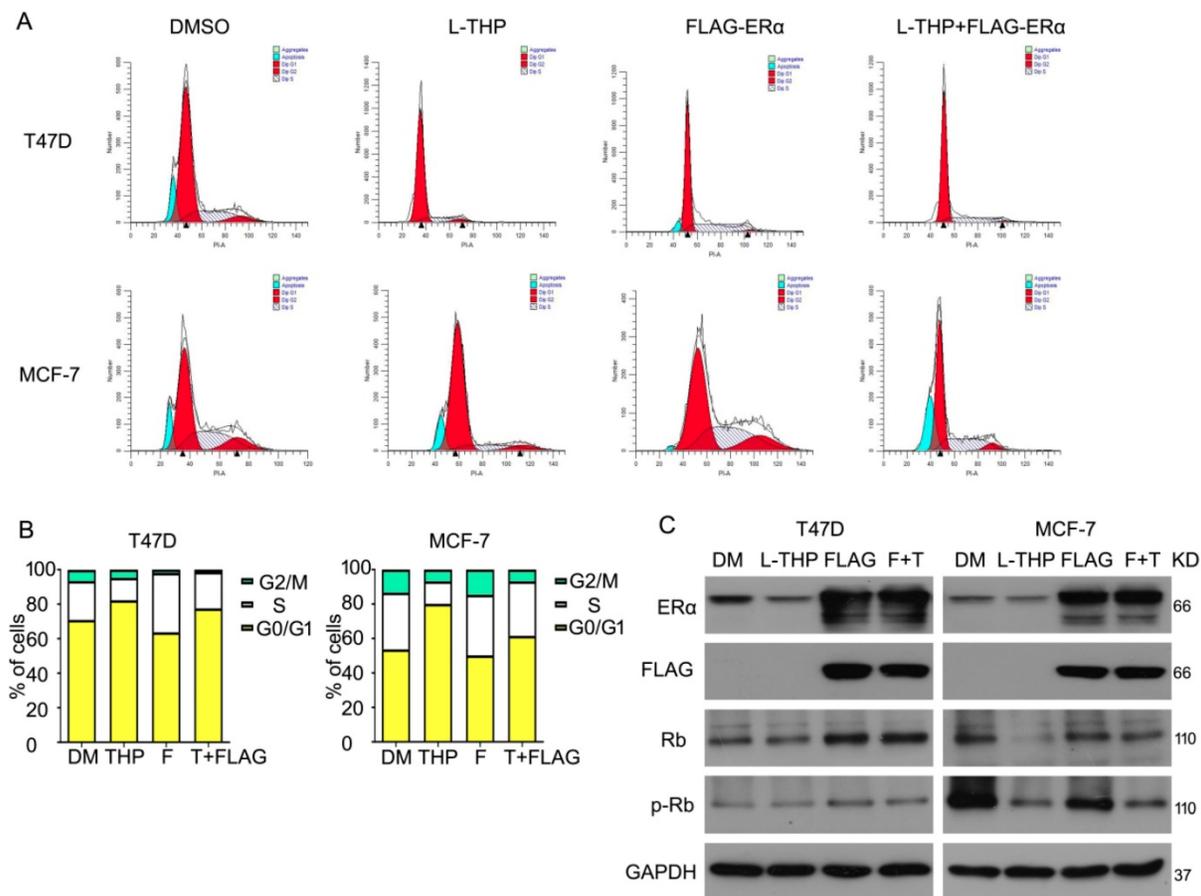


Figure 6. ER α overexpression abrogates partly growth inhibition induced by L-THP. (A) Cell cycle assay was performed on MCF-7 and T47D cells exposed to L-THP, FLAG-ER α or FLAG-ER α +L-THP. (B) Cell number was counted at different distribution. (C) Western blot assay was performed to test expression of ER α , FLAG, p-Rb and Rb proteins.

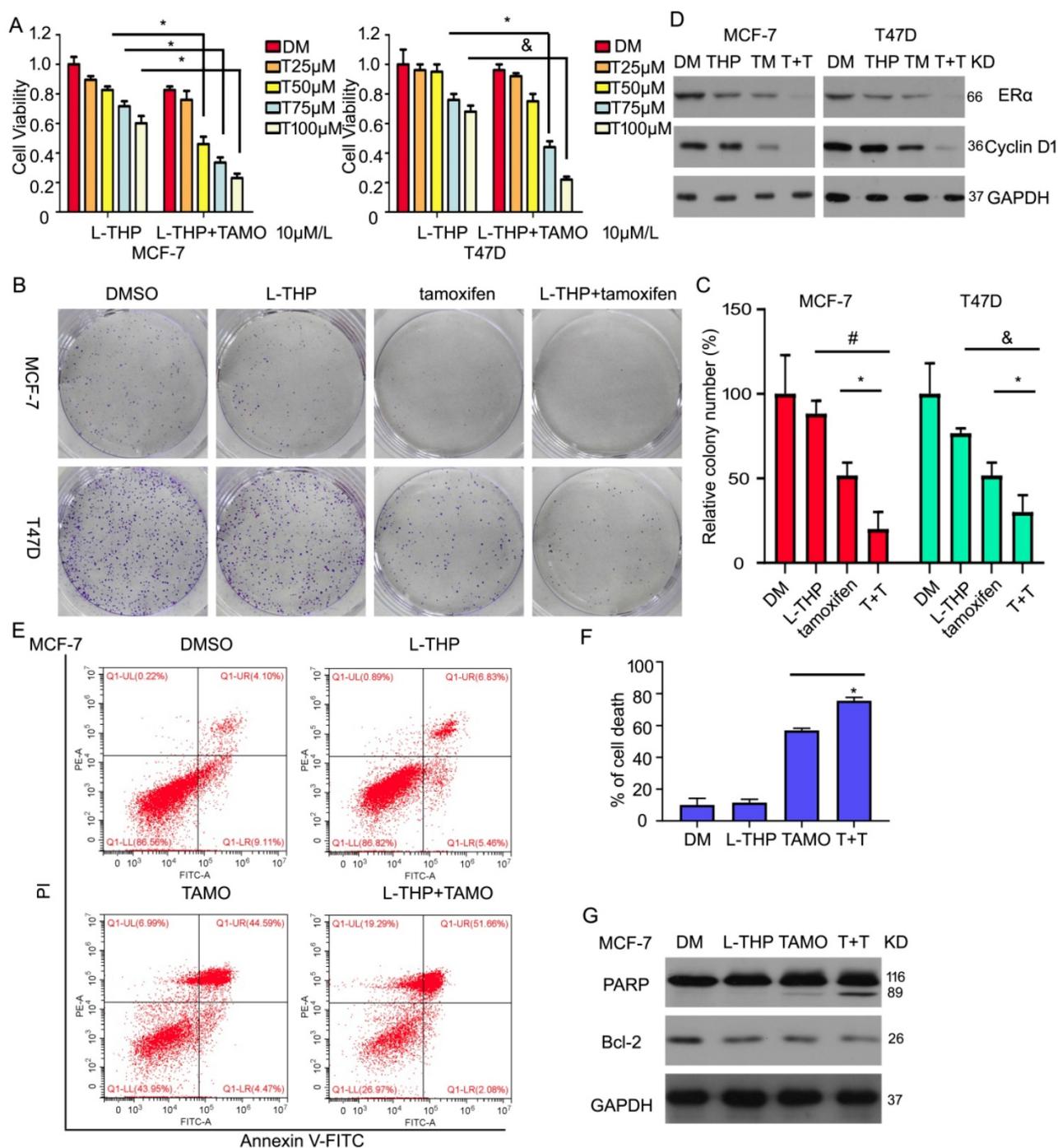


Figure 7. L-THP increases the sensitivity of BCa cells to tamoxifen. (A) Cell viability was performed on BCa cells post with L-THP at various doses or L-THP+tamoxifen (TAMO). (B) Colony formation assay was performed on MCF-7 and T47D exposed to L-THP or L-THP+tamoxifen treatment. (C) The colony numbers have been calculated. (D) Protein obtained from treated cells with L-THP or L-THP+tamoxifen was subjected to western blot assay to detect expression of ERα and Cyclin D1. (E) Cell apoptosis was performed with Annexin V-FITC staining to evaluate cell death. (F) Dead cells were counted. (G) The treated cells with L-THP or L-THP+tamoxifen was subjected to western blot for PARP and Bcl-2 expressions. *p<0.05, *p<0.01, #p<0.001.

Discussion

L-THP has been used for analgesic and sedative effect in clinical practice in China for more than 40 years. L-THP belongs to tetrahydro protoberberine isoquinoline alkaloid and is naturally extracted from *Corydalis* and genera *Stephania* [34]. To overcome the

dose-induced toxicity of chemotherapeutic reagents in the recent year, growing studies explored the potential function of a number of natural products with less toxicity and higher potency as chemotherapeutic drugs in cancer [35]. Considering that L-THP is a natural product and applied clinical therapy for pain, we ensure its safety. Some published

reports claimed that L-THP owns anticancer ability. L-THP enhances the sensitivity of leukemia to doxorubicin. Moreover, L-THP in combination with berberine induces cell growth-inhibition in MDA-MB231 cell [23, 24]. In this study, we discovered that L-THP suppressed cell proliferation in ERα positive breast cancer (ERα⁺ BCa).

Estrogen receptor alpha (ERα) plays a main driver in progression of breast cancer, resulting in breast cancer is classified as ERα positive breast cancer and others. ERα⁺ BCa is sensitivity to ERα inhibitor or estrogen (E2) antagonist which blocks

estrogen-ERα binding. The transcriptional activity of ERα triggers multiple estrogen-responsive genes which is essential in development and survival of BCa cells. Tamoxifen and fulvestrant are regarded as important drugs for ERα positive breast cancer patients. Unfortunately, resistance to chemotherapy drugs happens in the patients with breast cancer develop. Hence, we seek to find a safe and effective drug to replace or enhance tamoxifen/fulvestrant sensitivity to breast cancer patients to reduce resistance.

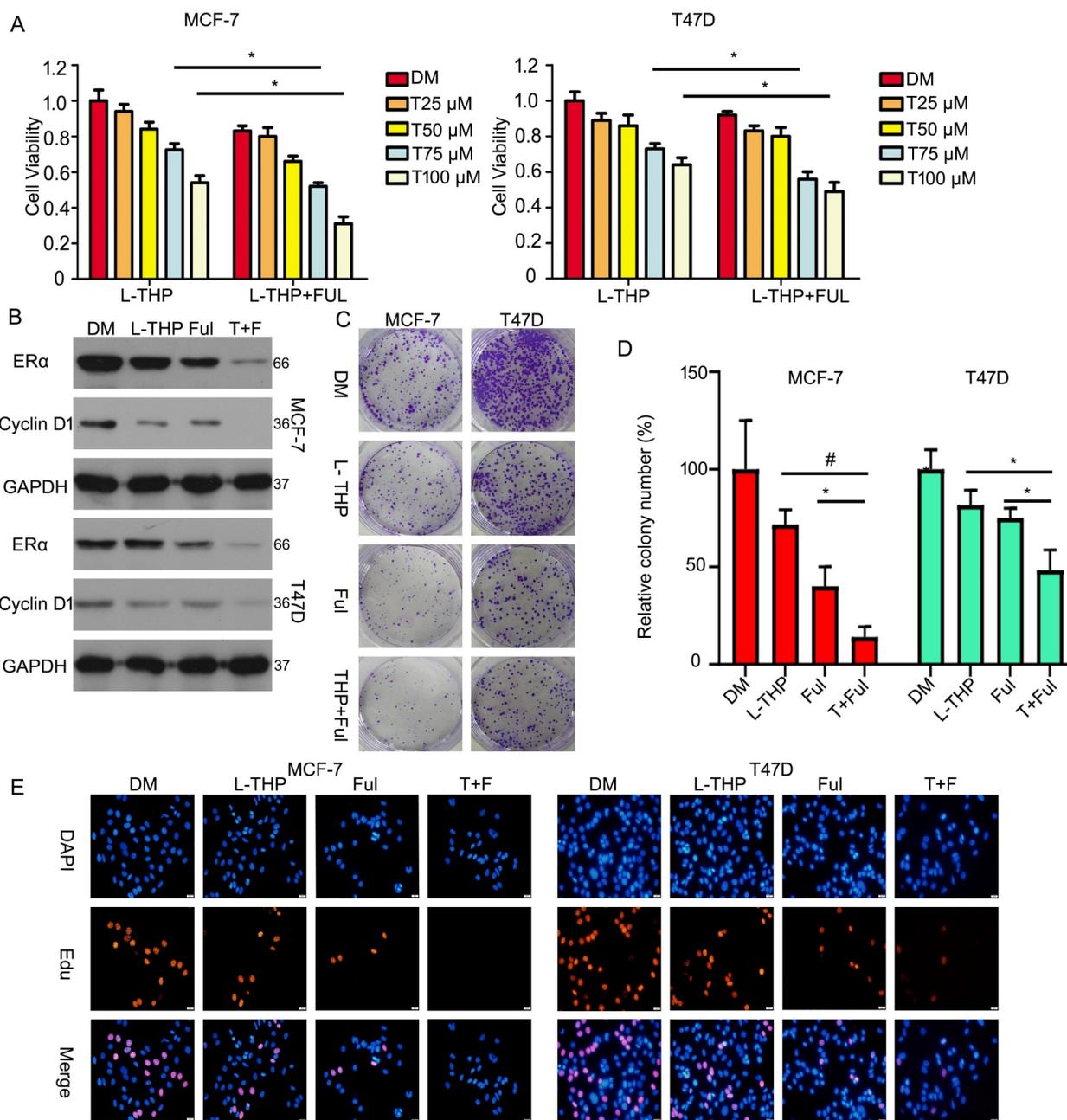


Figure 8. L-THP and fulvestrant synergistically suppress the growth of BCa cells. (A) Cell viability was performed on BCa cells post with L-THP at various doses or L-THP+fulvestrant (FUL). (B) Protein obtained from treated cells with L-THP or L-THP+fulvestrant (FUL) was subjected to western blot assay to detect expression of ERα and Cyclin D1. (C) Colony formation assay was performed on MCF-7 and T47D exposed to L-THP or L-THP+fulvestrant (FUL) treatment. (D) The colony numbers have been calculated. (E) Edu staining assay was performed on BCa cells treated with L-THP, fulvestrant or L-THP+fulvestrant. *p<0.05, #p<0.001.

Interestingly, we clarified that L-THP suppressed the growth of ER α ⁺ BCa cells depends on ER α status. Cell proliferation was inhibited by L-THP in BT474, MCF-7 and T47D ER α ⁺ BCa cells. The phenomenon mainly caused by cell apoptosis and cycle arrest. We found that L-THP increased cell rate at G0/G1 phase via blocking G0/G1 to S transition. But cell apoptosis be not observed. This findings suggested that anti-proliferative effect induced by L-THP results from cell cycle arrest rather than cell apoptosis. We speculated whether L-THP enhances the sensitivity of ER α BCa cells to estrogen or estrogen receptor antagonist. Tamoxifen, as estrogen antagonist, interdicts binding between estrogen and ER. Fulvestrant is an estrogen receptor antagonist. Both in combination of L-THP and tamoxifen or fulvestrant has a significant synergistic effect.

On the other hand, we further explored the most likely molecular mechanism. We found that L-THP decreased protein expression of ER α but not changed the translocation using western blotting and confocal assay. The decreased ER α may be happened in transcription and translation levels. RT-qPCR confirmed that L-THP did not induce the decreased mRNA level of ER α but downregulated its downstream genes, including PS2 and Cyclin D1. In addition to transcription, we used CHX to inhibit protein synthesis to detect the role of L-THP on ER α expression. We found that L-THP promoted the degradation of ER α protein in the presence and absence of estrogen. Based on many reported and our previously studies [30, 36, 37], we know that ubiquitin proteasome system (UPS) involves in the degradation of ER α . The level of ubiquitin on ER α was remarkably increased by L-THP treatment, suggesting L-THP promotes the degradation of ER α via accumulating the ubiquitin on ER α . Moreover, dual-luciferase reporter assay indicated L-THP inhibitory effect on the transcriptional activity of ER α . Additionally, the conformations of ER α and L-THP can be bind in three dimensional and two-dimensional, indicating that ER α interacts with L-THP. Moreover, we assessed whether the conjugate site between L-THP and ER α locates at the hinge region K302, K303 which participates degradation by UPS. However, the conjugate site is not at K302 or K303. We think that L-THP induced-ER α degradation may be via affecting some deubiquitinase or E3 ligase and so on or via changing ER α molecular structure. Delightedly, L-THP-induced cell cycle arrest can be partly abrogated by ER α overexpression.

The current study, our data confirmed that L-THP exists an inhibitory role in the growth of ER α ⁺ BCa cells via promoting the degradation of ER α and suppressing its transcriptional activity. Additionally,

L-THP enhances the sensitivity of ER α ⁺ BCa cells to tamoxifen and fulvestrant. Therefore, this study initiates a significant function of L-THP for cancer therapy and L-THP in combination with estrogen or estrogen receptor antagonist may provide a strategy to reduce resistance in ER α ⁺ breast cancer patients.

Supplementary Material

Supplementary figures and tables.

<http://www.ijbs.com/v16p2192s1.pdf>

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Competing Interests

The authors have declared that no competing interest exists.

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