

Supplemental Information

Smoke-induced SAV1 gene promoter hypermethylation disrupts YAP negative feedback and promotes malignant progression of non-small cell lung cancer

Supplemental Experimental Procedures

RNA extraction and quantitative real-time PCR analyses (qRT-PCR)

The TRIzol reagent was used to extract total RNA. cDNA was generated with the PrimeScriptRT reagent kit. GAPDH was performed as an internal reference for cytoplasmic gene expression. The expression change of genes was calculated by the $2^{-\Delta\Delta C_t}$ method.

Western Blot

Total protein was extracted from cells using RIPA buffer (Thermo Scientific, Rockford, IL, USA) in the presence of protease inhibitors (Protease Inhibitor Cocktail, Thermo Scientific). The protein concentration of lysates was measured using a BCA Protein Assay Kit (Thermo Scientific). Equivalent amounts of protein were mixed with 5×Lane Marker Reducing Sample Buffer (Thermo Scientific), and resolved by electrophoresis in a 10% SDS–polyacrylamide gel and then transferred onto Immobilon-P Transfer Membrane (Merck Millipore). The membranes were blocked with 5% non-fat milk in Tris-buffered saline and then incubated with the primary antibodies followed by secondary antibody. The signal was detected using enhanced chemiluminescence western blot detection kit. The primary antibody for GAPDH (abs132004) was from absin. The primary antibody for SAV1 (#13301) was from Cell Signaling Technology. The primary antibody for β -Actin (A5316) was from sigma.

Flow Cytometry Analysis

Cell cycle experiment was carried out using the cell cycle staining kit (MultiSciences (Lianke) Biotechnology) following the manufacture's instruction. Briefly, cells were trypsinized and incubated with the primary antibody from the kit. The cell cycle was evaluated using flow cytometry.

Immunohistochemistry

The sections were dried at 60°C for 2 h and then deparaffinized in xylene and rehydrated using a series of graded alcohol washes. The tissue slides were then treated with 3% hydrogen peroxide in methanol for 15 min to quench endogenous peroxidase activity and antigen retrieval then performed by incubation in 0.01 M sodium citrate buffer (pH 6.0) and heating using a microwave oven. After a 1 h preincubation in 10% goat serum, the specimens were incubated with primary antibody overnight at 4°C. The tissue slides were treated with a non-biotin horseradish peroxidase detection system according to the manufacturer's instruction (DAKO, Glostrup, Denmark). Two different pathologists evaluated the immunohistological samples.

Primers

Primers for qRT-PCR

GAPDH

Forward: 5'-ATTCCATGGCACCGTCAAGGCTGA-3'

Reverse: 5'-TTCTCCATGGTGGTGAAGACGCCA-3'

YAP

Forward: 5'-TAGCCCTGCGTAGCCAGTTA-3'

Reverse: 5'-TCATGCTTAGTCCACTGTCTGT-3'

CTGF

Forward: 5'-CAGCATGGACGTTTCGTCTG-3'

Reverse: 5'-AACCACGGTTTGGTCCTTGG-3'

CYR61

Forward: 5'-CTCGCCTTAGTCGTCACCC-3'

Reverse: 5'-CGCCGAAGTTGCATTCCAG-3'

SAV1

Forward: 5'- AAGGCCCAATACAGGCATCC -3'

Reverse: 5'- TGGTCATATTTACAGGGGCT -3'

SURVIVIN

Forward: 5'- ATGACGACCCCATAGAGGAAC -3'

Reverse: 5'- CGCACTTTCTCCGCAGTTTC -3'

MST1

Forward: 5'- GGAGACGGTACAGCTGAGGA -3'

Reverse: 5'- GCTGCCATAGGACCCTTCTC-3'

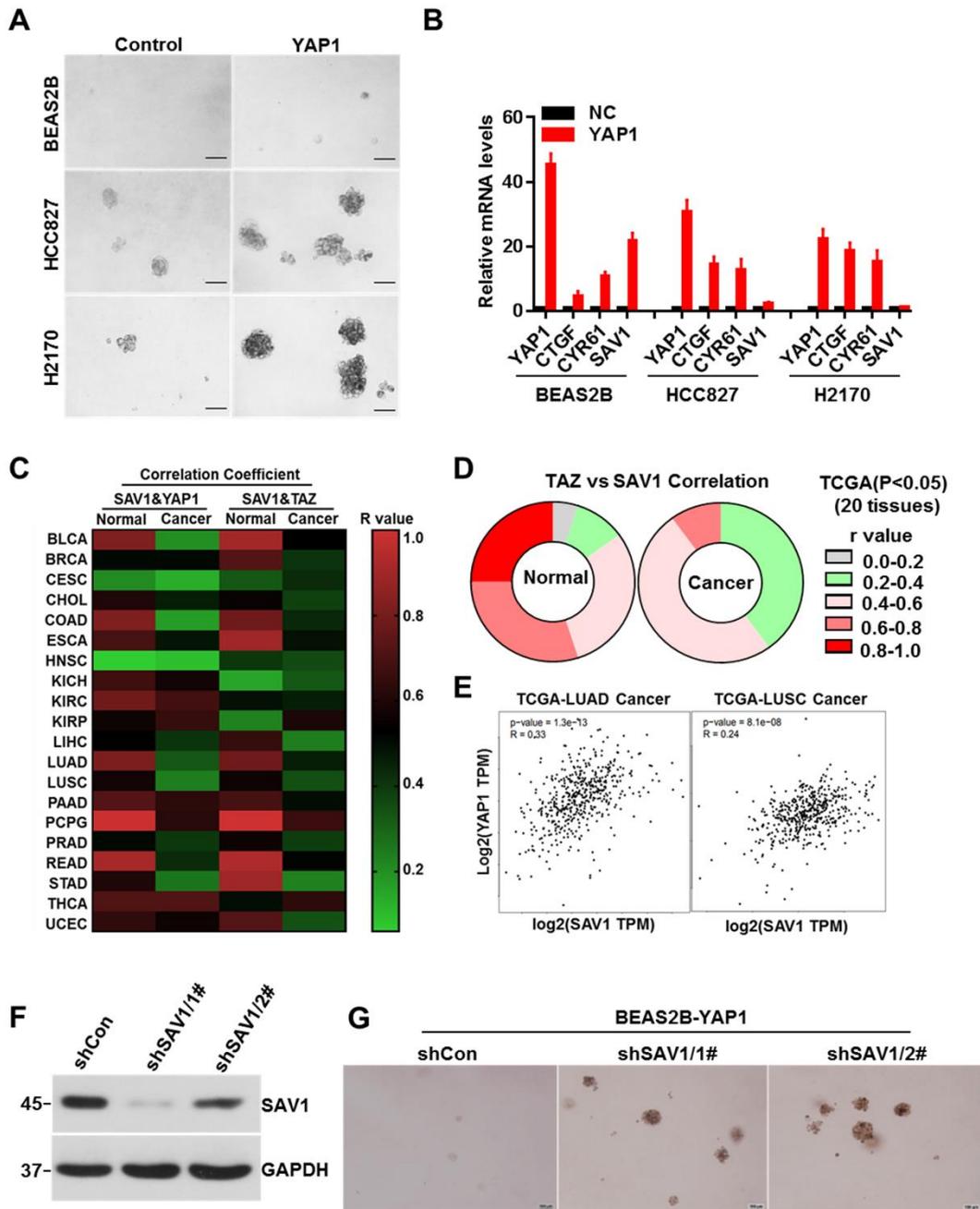
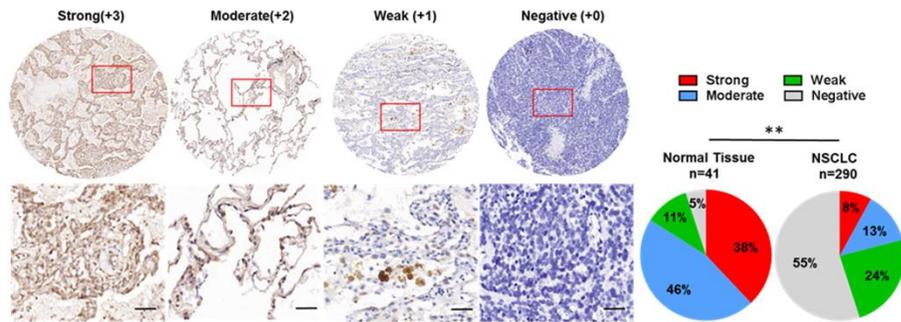


Figure.S1

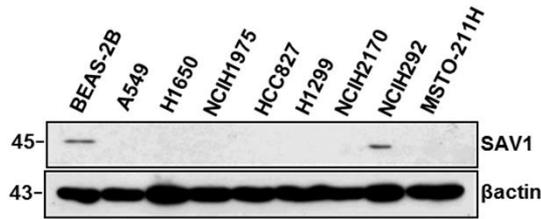
Figure S1, related to Figure 1. YAP-SAV1 negative feedback loop maintain lung epithelial cell

(A) Representative pictures of colony in soft agar. Scale bar: 100 μ m. (B) YAP, CTGF, CYR61 and SAV1 mRNA levels in different lung cell lines were detected by qRT-PCR. Student's t-test, Mean \pm s.d. (n=3), **** p < 0.0001. (C) The correlation between SAV1 and YAP1/TAZ in normal group or cancer group. (D) TCGA data shows the stronger correlation between SAV1 and TAZ in normal tissue than cancer group in 20 tissues. (E) Using TCGA transcriptome data, the transcripts per million (TPM) expression value shows the positive correlation between SAV1 and YAP in LUAD and LUSC. (F) The knockdown efficiency of shRNA was tested by WB assay. (G) Representative pictures of colony in soft agar. Scale bar: 100 μ m.

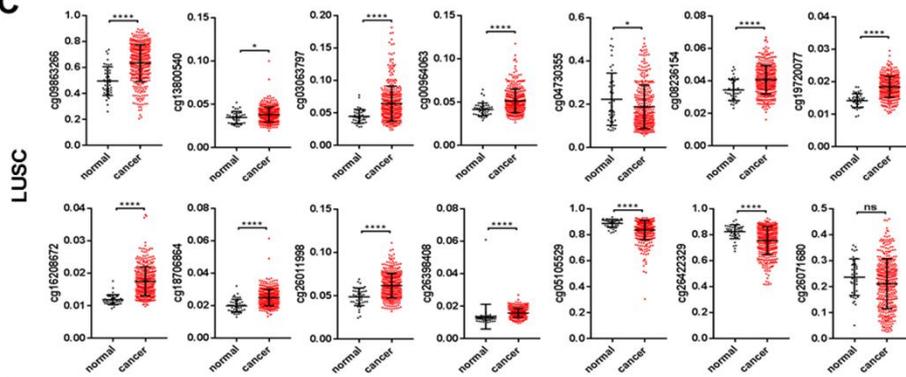
A



B



C



D

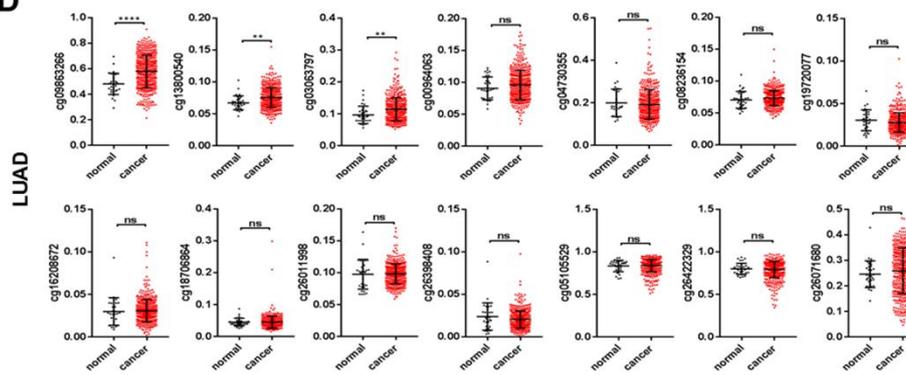


Figure.S2

Figure S2, related to Figure 2. SAV1 promoter methylation inhibits YAP-SAV1 negative feedback regulation in lung cancer cells.

(A) Immunohistochemistry (IHC) was used to determine the level of SAV1 expression in clinical lung tissues. Scale bar: 50 μm . (B) WB was used for detecting SAV1 protein expressions in different lung cell lines. (C, D) Analysis of the genome-wide DNA methylation data of LUAD and LUSC samples obtained from TCGA dataset (<http://cancergenome.nih.gov/>) found that a total of 14 cg sites are on the SAV1 gene. Student's t-test, Mean \pm s.d. **** $p < 0.0001$.

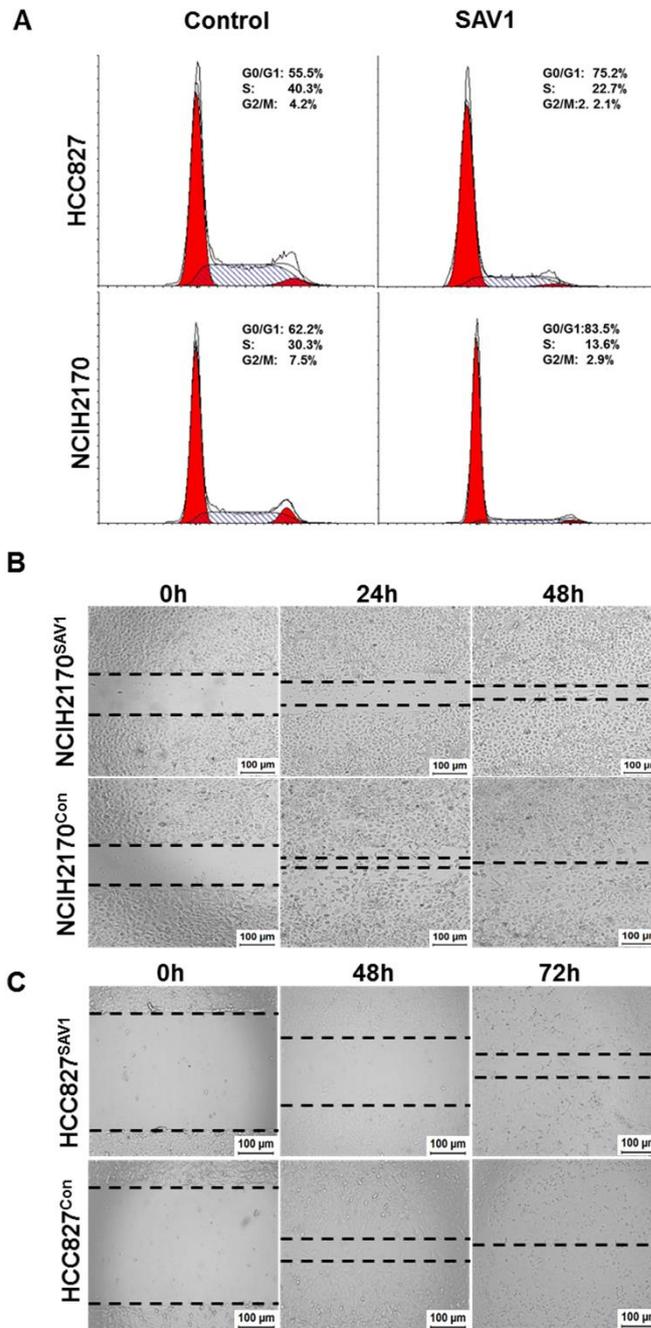


Figure.S3

Figure S3, related to Figure 4. Exogenous expression of SAV1 inhibits the malignancy of Lung Cancer Cell.

(A) Cell-cycle distribution analysis was performed by flow cytometry. (B, C) To measure cell migration, in vitro scratch wound assay was performed. Scale bar: 100 μ m.

A

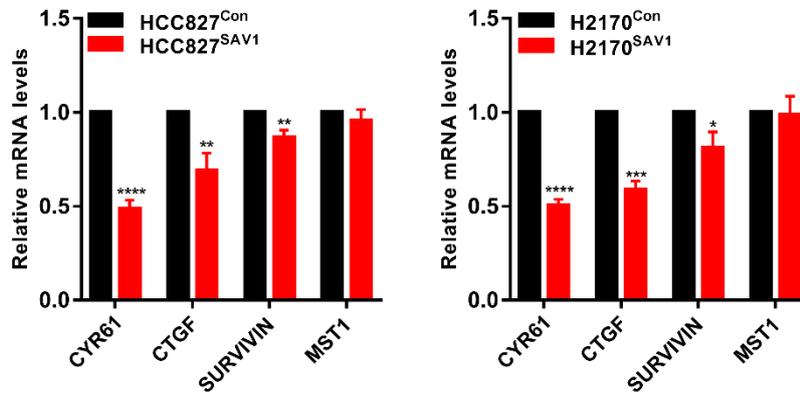


Figure.S4

Figure S4, related to Figure 5. Exogenous expression of SAV1 inhibits YAP transcriptional activity by stabilizing MST1 protein. (A) qPCR was performed to measure the expression of YAP target genes in HCC827 and H2170 cell lines with or without SAV1 overexpression.