

## **Supplemental Material**

### **Regulating Neutrophil PAD4/NOX-Dependent Cerebrovascular Thromboinflammation**

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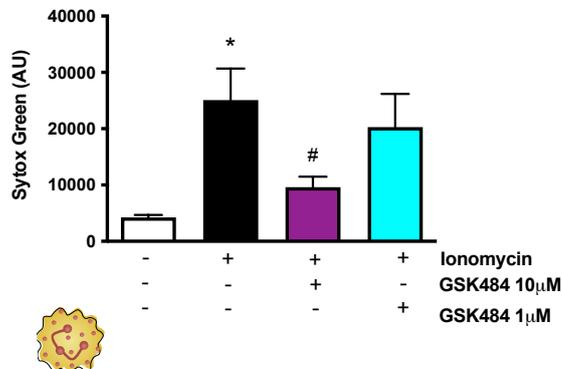
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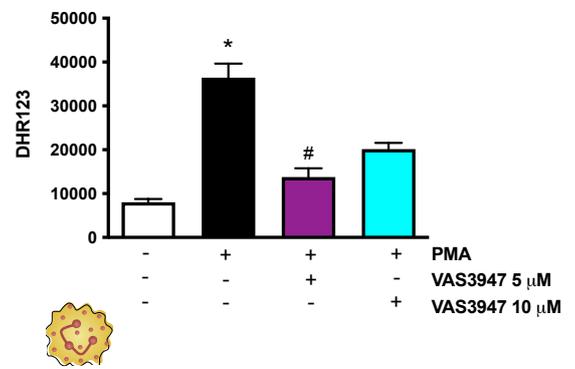
Email: [felicity.gavins@brunel.ac.uk](mailto:felicity.gavins@brunel.ac.uk)

## Supplemental Figure 1.

**A**



**B**

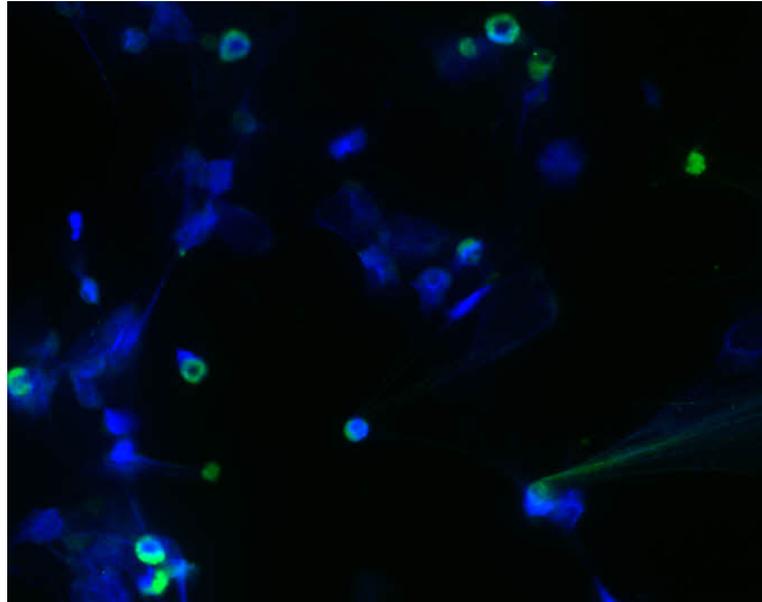


### Supplemental Figure 1: GSK484 and VAS3947 dose response

**(A)** NETs were quantified by Sytox green intensity using a plate reader (BioTek; excitation = 485 nm, emission=525 nm) from unstimulated and ionomycin (4 μM)-stimulated neutrophils isolated from human volunteers; unstimulated (n=4), ionomycin stimulated (n = 4), GSK484 (10 μM) pre-treated ionomycin-stimulated neutrophils (n=4), and GSK484 (1 μM) pre-treated ionomycin-stimulated neutrophils (n=4)

**(B)** DHR123 production was measured using a plate reader (BioTek; excitation = 485 nm, emission = 525 nm) from unstimulated neutrophils, PMA-stimulated neutrophils, VAS3947 (5 μM) treated PMA-stimulated neutrophils and VAS3947 (5 μM) treated PMA-stimulated neutrophils (n=3 in all groups). Graphs are expressed as mean±SEM from independent experiments. All imaging analysis was done in a double-blinded fashion.

**Supplemental Figure 2.**



**Supplemental Figure 2:** For immunocytochemistry, cells were incubated with NET specific antibodies histone H3 mouse anti-H3Cit (1:200) followed by species-specific secondary antibodies coupled with Alexa Fluor Dyes (1:1000, Alexa Fluor 488 goat anti-mouse, green) and co-stained with DAPI (nuclear stain, blue) After mounting (Fluoromont-G, Southern Biotech, Birmingham, Alabama, USA) the images were visualized by Nikon Eclipse Ti inverted epifluorescence microscope (Minato-ku, Tokyo, Japan). We quantified *in-vitro* NETs by measuring the percentage of CitH3 stained DNA over total number of cells (DAPI stained). Figure shows a protrusion or a thread like structure coming from the main cell body, which was positive for nuclear stain DAPI, and neutrophil elastase (NET bound protein), as well as citrullinated histone.

**Supplemental Table 1. Demographic and clinical characteristics of controls and Sickle Cell Disease (SCD) patients**

<b>Variable</b>	<b>Control</b>	<b>SCD patients</b>
Number	15	14
Gender	Eight males, seven females	Five males, nine females
Age range	25-54 years old	20-36 years old
History of cerebrovascular accidents	0	5 (38.46%)
Leukocytes (Ref: 3.6-11.2 K/ $\mu$ l)	Not recorded	10.73 $\pm$ 1.09
Red blood cells (Ref: 4.06-5.63 M/ $\mu$ l)	Not recorded	2.95 $\pm$ 0.19
Hemoglobin (Ref: 12.5-16.3 gm/dl)	Not recorded	9.12 $\pm$ 0.37
Hematocrit (Ref: 36.7-47.1 %)	Not recorded	26.84 $\pm$ 1.23
Platelet count (Ref: 159-386 K/ $\mu$ l)	Not recorded	310.6 $\pm$ 30.13
Neutrophils (Ref: 1.8-7.8 K/ $\mu$ l)	Not recorded	6.8 $\pm$ 0.90

'Ref:' indicates reference range. Data represented as mean  $\pm$  SEM in SCD patients.

### **Video Legends**

Video 1. Time-lapse video microscopy shows progression of thrombus formation in a cerebral arteriole of a control mouse demonstrating onset and complete cessation of blood flow.

Video 2. Time-lapse video microscopy shows progression of thrombus formation in a cerebral venule of a control mouse demonstrating onset and complete cessation of blood flow.

Video 3. Time-lapse video microscopy shows progression of thrombus formation in a cerebral arteriole of an STM mouse demonstrating onset and complete cessation of blood flow.

Video 4. Time-lapse video microscopy shows progression of thrombus formation in a cerebral venule of an STM mouse demonstrating onset and complete cessation of blood flow.