

SGK1 aggravates idiopathic pulmonary fibrosis by triggering H3k27ac-mediated macrophage reprogramming and disturbing immune homeostasis

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Supplemental figure legends

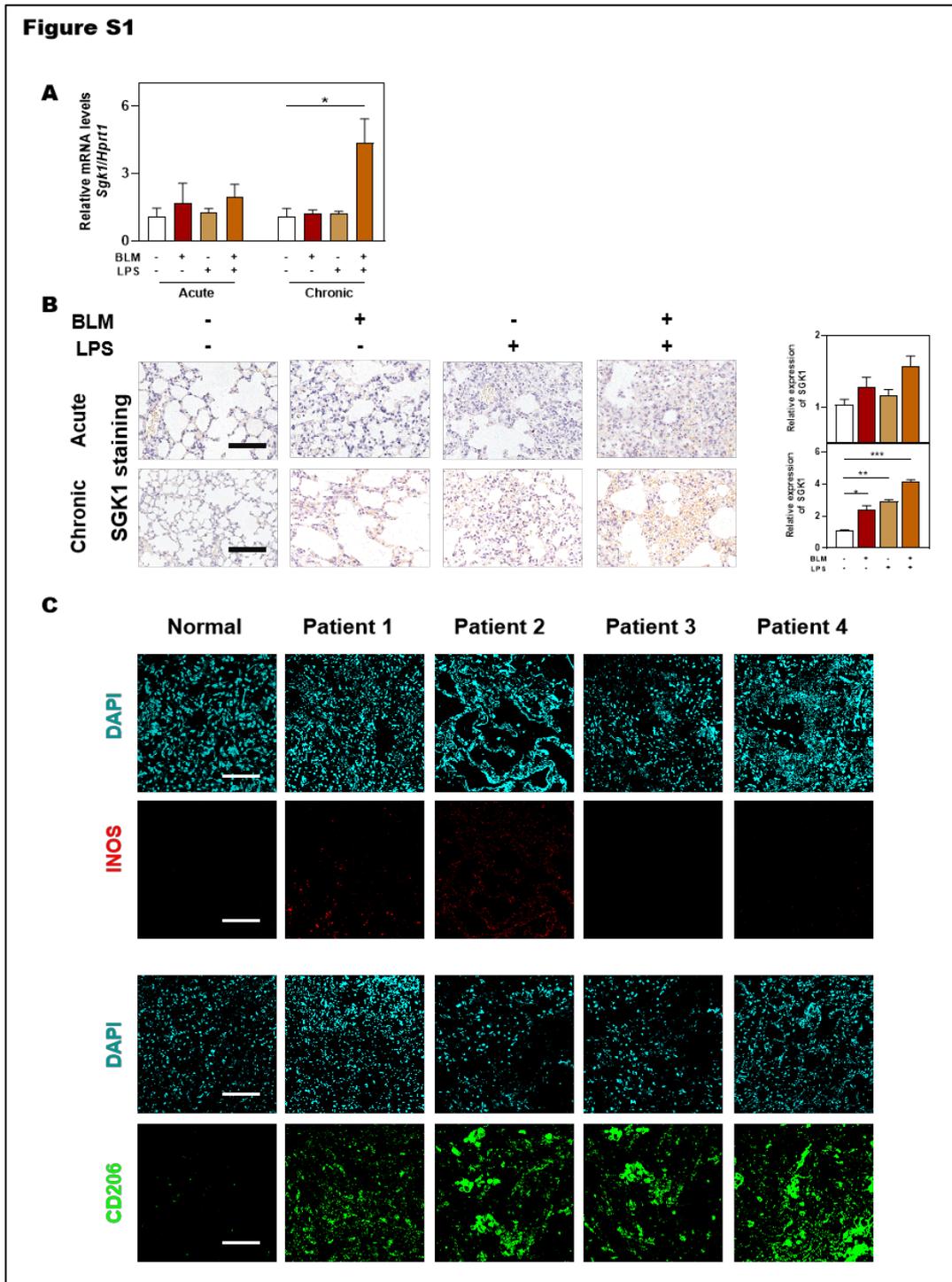


Figure S1. Expressions of SGK1 in BLM- and LPS- induced acute and chronic IPF mice models. (A) The mRNA levels of *Sgk1* in acute and chronic IPF mice. **(B)** Representative images of immunohistochemistry staining of SGK1 in acute and chronic IPF mice lungs (Scale bar = 100 μ m). **(C)** Representative images of

immunofluorescent staining of iNOS and CD206 in normal and IPF patient lungs. Scale bar = 200 μ m. Statistical significance: * P <0.05, ** P <0.01, *** P <0.001, compared with relative groups (n=6).

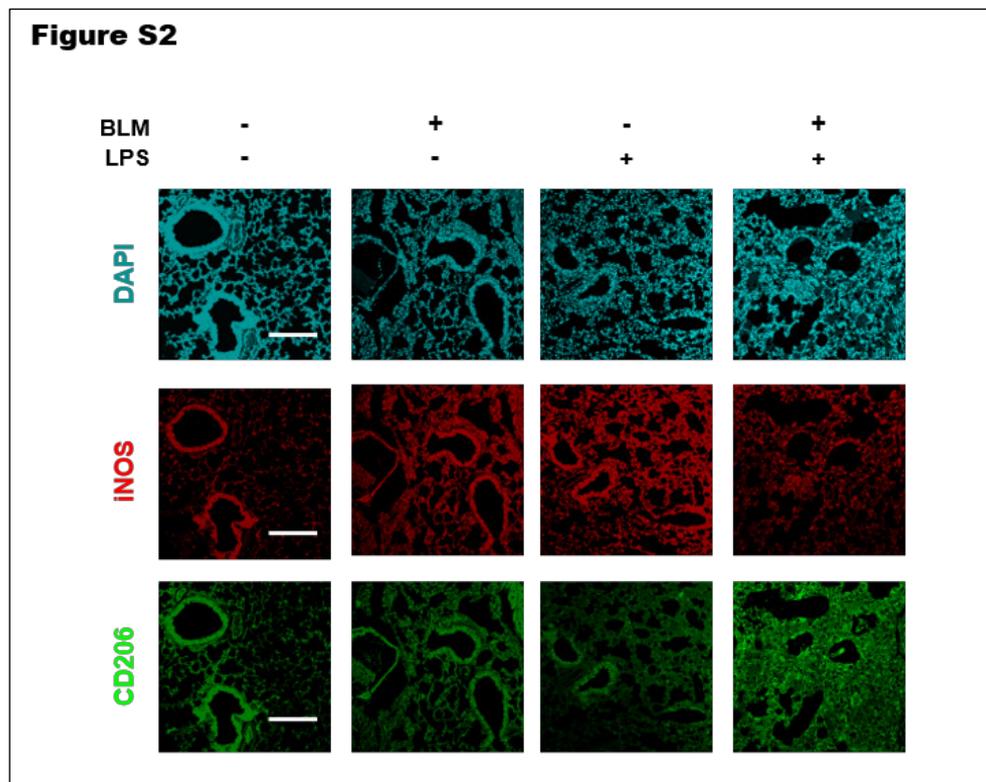


Figure S2. BLM + LPS promotes the reprogramming of macrophages from M1 to M2. Representative images of immunofluorescent staining of iNOS and CD206 in lungs. Scale bar = 200 μ m.

Figure S3

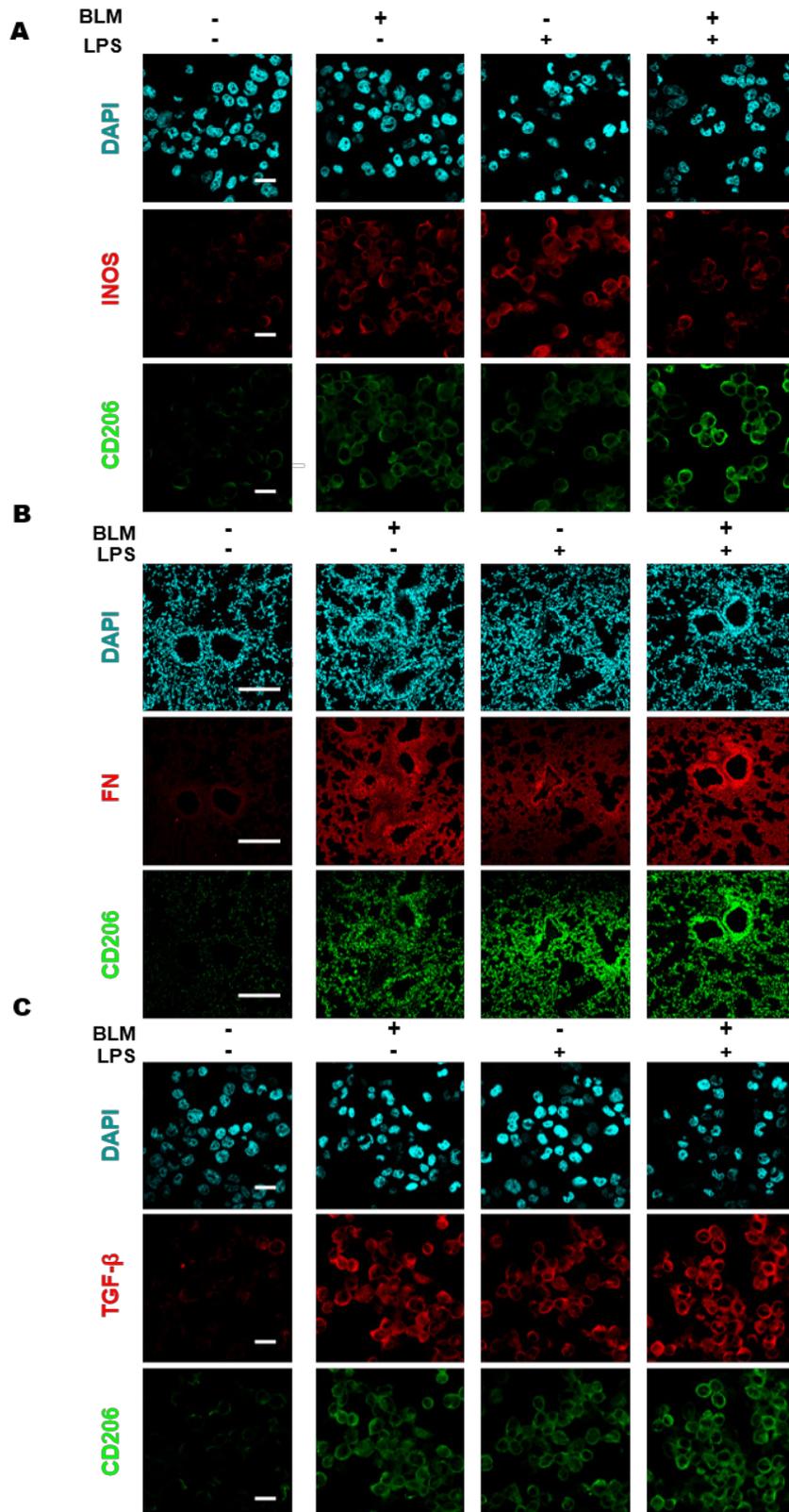


Figure S3. BLM plus LPS increases the number of M2 macrophages, aggravates

fibrotic injury along with increases TGF- β 1 secretion. Representative images of immunofluorescent co-staining of iNOS and CD206 **(A)** in THP-1 cells (Scale bar = 50 μ m), and co-staining of FN and CD206 **(B)** in lung tissues (Scale bar = 200 μ m). **(C)** Representative images of immunofluorescent result of TGF- β and CD206 in THP-1 cells treated with BLM and/or LPS. Scale bar = 50 μ m.

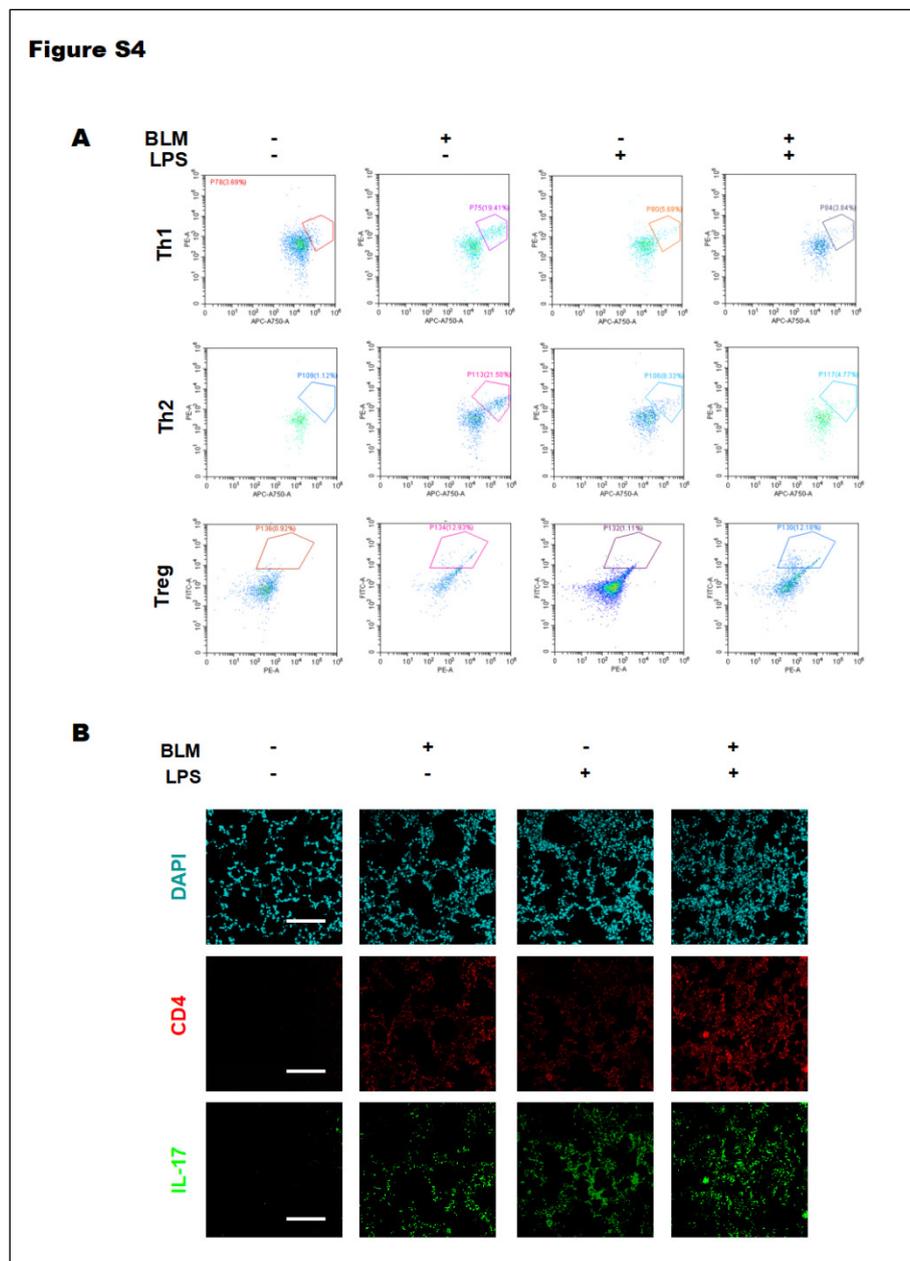


Figure S4. BLM plus LPS increases the number of Th17 cells. (A) Flow cytometry analyzes number of Th1, Th2 and Treg cells in the lung. **(B)** Representative images of

immunofluorescent result of CD4 and IL-17 in lungs. Scale bar = 200 μ m.

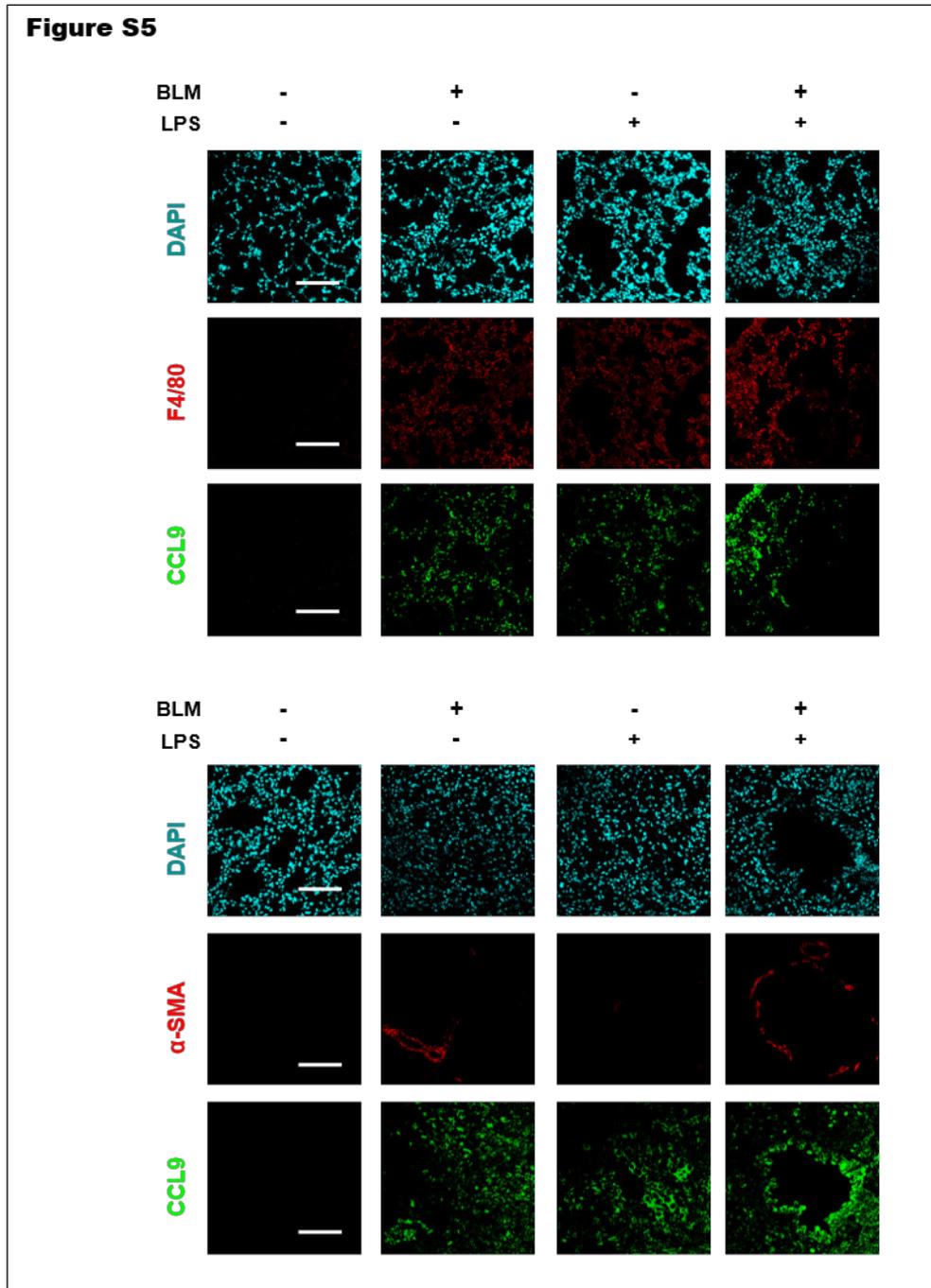


Figure S5. Macrophage rather than the myofibroblasts were the main producers of CCL9 in fibrotic lung tissues. Representative images of co-immunofluorescent staining of CCL9 with F4/80 and α -SMA. Scale bar = 200 μ m.

Figure S6

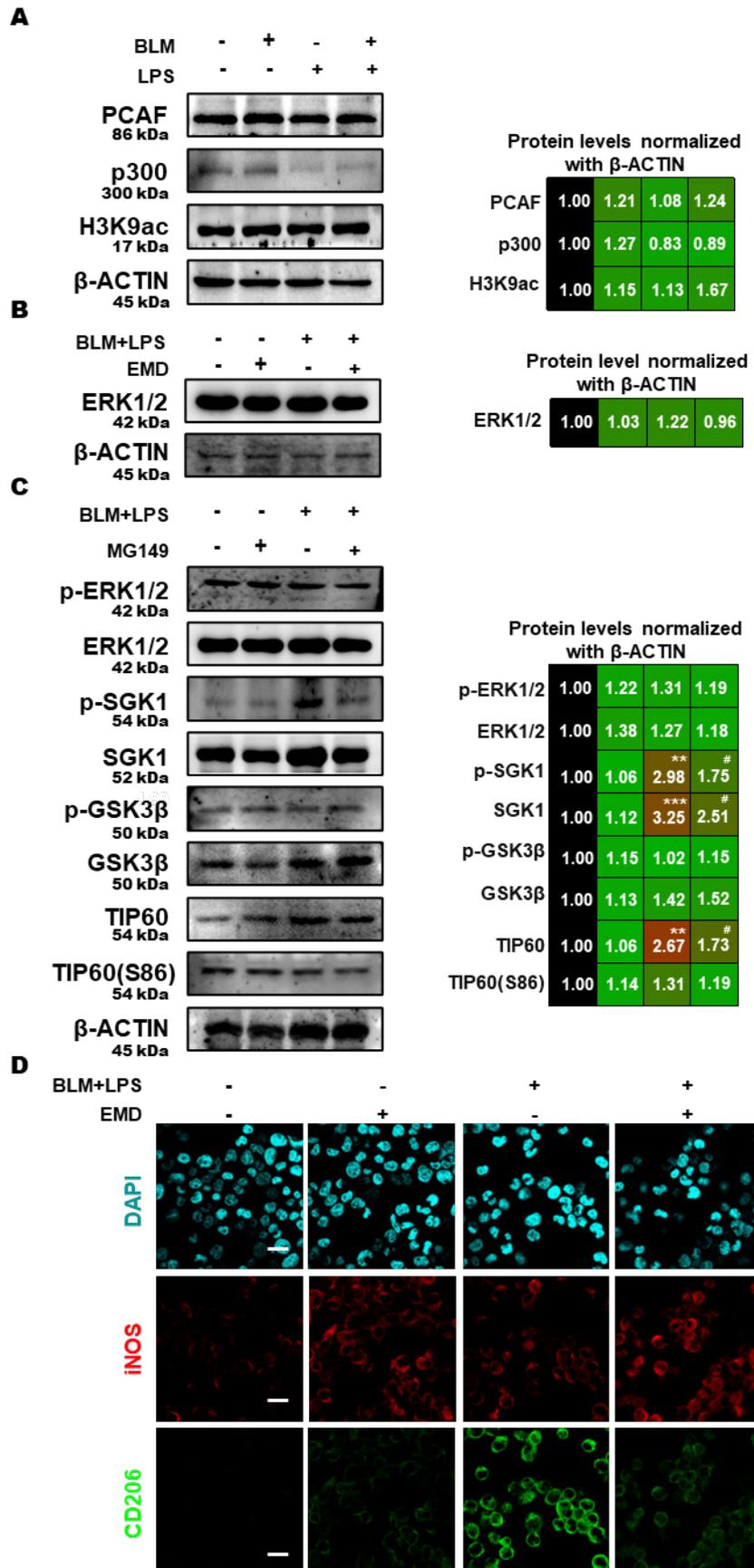


Figure S6. Macrophage reprogramming is regulated by SGK1-mediated GSK3 β -TIP60 pathway. Representative immunoblots against ERK1/2 **(A)**, PCAF, p300, K3K9ac **(B)**, p-ERK1/2, ERK1/2, p-SGK1, SGK1, p-GSK3 β , GSK3 β , TIP60, TIP60 (S86) and β -ACTIN **(C)** in THP-1 cells. **(D)** Representative images of immunofluorescent result of iNOS and CD206 in THP-1 cells. Scale bar = 50 μ m. Statistical significance: ** P < 0.01, *** P < 0.001, compared with control group; # P < 0.05, compared with BLM + LPS group (n = 3).

Figure S7

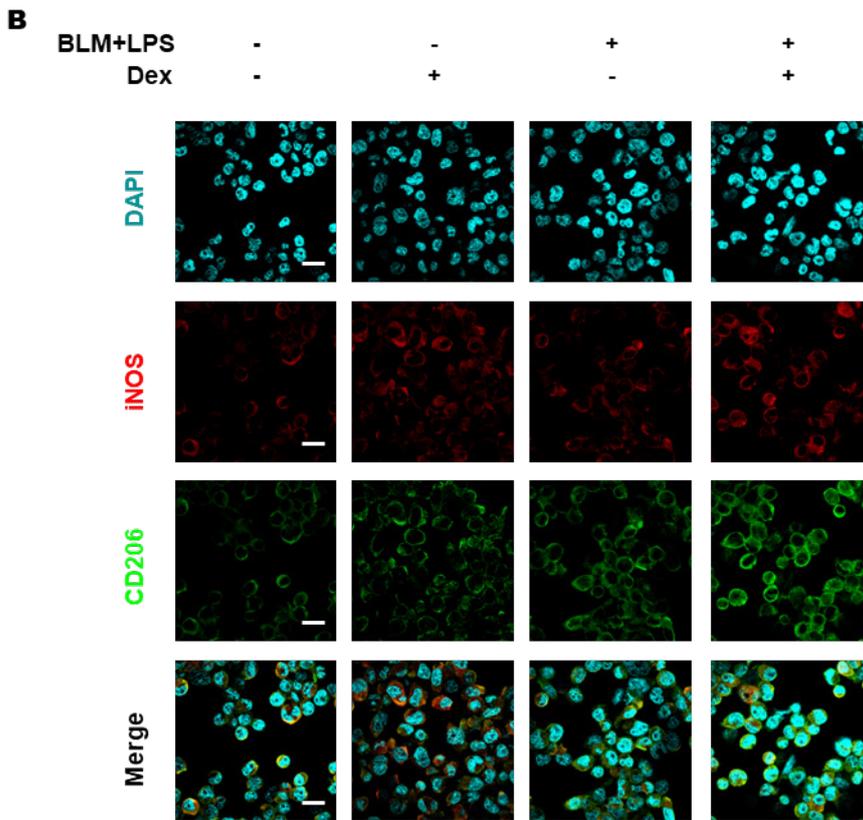
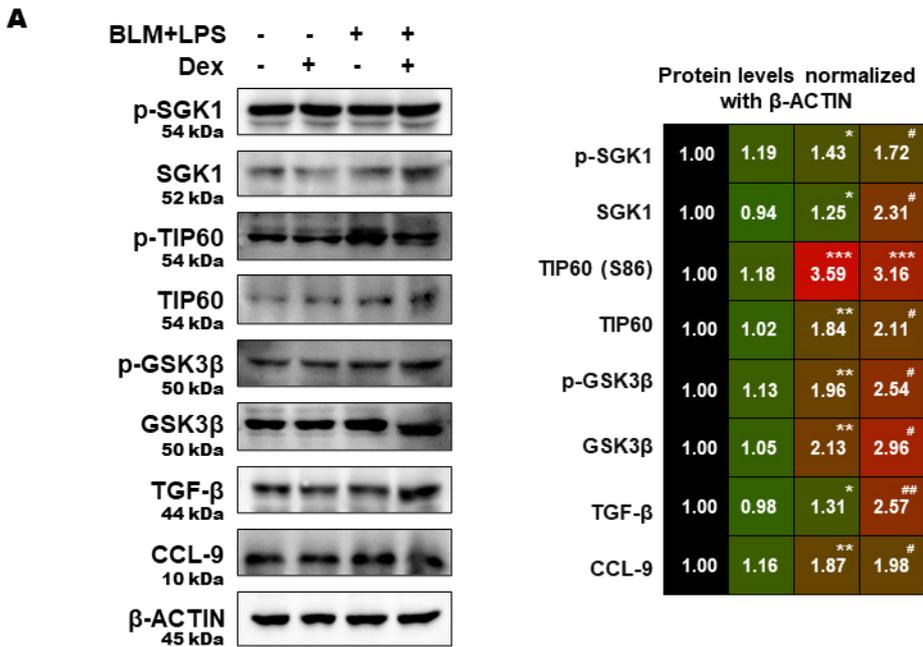


Figure S7. Macrophage reprogramming is promoted by activating SGK1. (A)

Representative immunoblots against p-SGK1, SGK1, p-GSK3β, GSK3β, TIP60, TIP60

(S86), TGF- β , CCL-9 and β -ACTIN in THP-1 cells. **(B)** Representative images of immunofluorescent result of iNOS and CD206 in THP-1 cells. Scale bar = 50 μ m. Statistical significance: * P < 0.05, ** P < 0.01, *** P < 0.001, compared with control group; # P < 0.05, ## P < 0.01, compared with BLM + LPS group (n = 3).

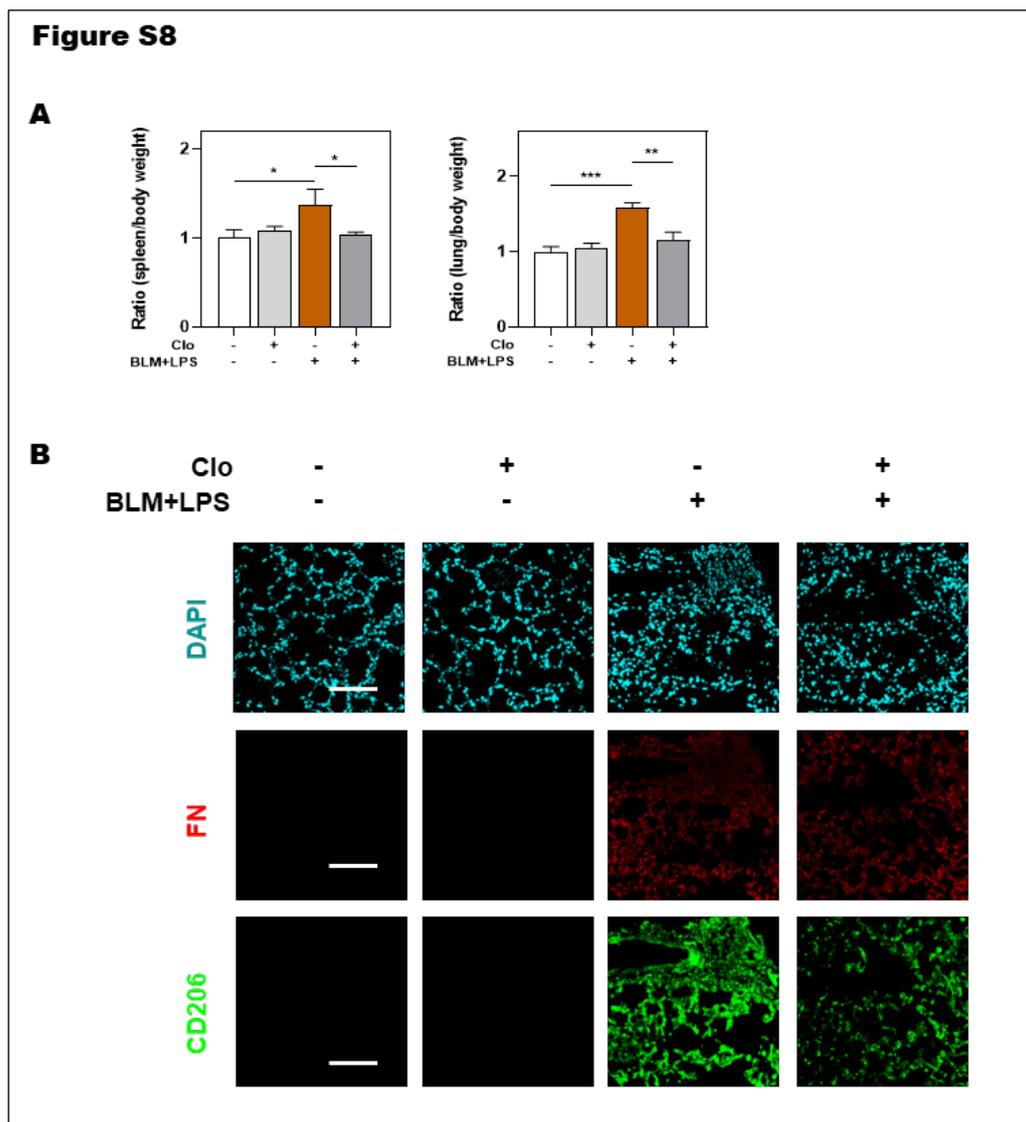


Figure S8. Lung fibrotic injury and M2 reprogramming process are largely improved after Clo intervention in BLM + LPS induced IPF mice. **(A)** Ratio of spleen and lung to body weight. **(B)** Immunofluorescent staining of FN and CD206 in

mouse lungs. Scale bar = 200 μm .

Table S1 The baseline characteristics of the IPF patients.

Characteristics	Values (<i>n</i> = 27)
Age (y)	56.44±8.71
Sex (male/female)	14/13
BMI (kg/m ²)	25.67±3.54
Smoking status	
Current smoker	3
Exsmoker	1
Nonsmoker	23
Exposure history (house dust,birds, cotton, chemical materials)	
None	27
Yes	0
System disease (dermatomyositis, rheumatoid arthritis, sarcoidosis, lupus & scleroderma)	
None	27
Yes	0
Pulmonary function tests	
FVC	3.15±0.89
% FVC	95.04±18.07
FEV ₁	2.51±0.81
% FEV ₁	91.94±20.22
FEV ₁ /FVC (%)	78.98±10.00
% DL _{CO}	88.74±16.83
% DL _{CO} /V _A	102.11±18.98
Echocardiography	
% Ejection fraction	63.63±3.28

Notes: Data presented as *n* or mean ± standard deviation, unless otherwise indicated.

Abbreviations: DL_{CO} = diffusing capacity of the lung for carbon dioxide, FEV₁ = forced expiratory volume in 1 s, FVC = forced vital capacity, V_A = alveolar volume.

Table S2 The antibodies used in our study.

Antibody	Catalog Number	Vendor
FIBRONECTIN	15613-1-AP	Proteintech Group (Rosemont, USA)
E-Cadherin	20874-1-AP	Proteintech Group
SGK1	28454-1-AP	Proteintech Group
iNOS	18985-1-AP	Proteintech Group
TGF beta1	21898-1-AP	Proteintech Group
CD206	60143-1-Ig	Proteintech Group
Histone-H3	17168-1-AP	Proteintech Group
F4/80	28463-1-AP	Proteintech Group
Phospho-GSK3 β	29125-1-AP	Proteintech Group
GSK-3 β	sc-377213	Santa Cruz Biotechnology (Texas, USA)
PCAF	sc-13124	Santa Cruz Biotechnology
TIP60	sc-32244	Santa Cruz Biotechnology
p300	sc-32244	Santa Cruz Biotechnology
MIP-1 γ	sc-74228	Santa Cruz Biotechnology
IL-17	sc-374218	Santa Cruz Biotechnology
APC anti-mouse cluster of differentiation 3 antibody	100236	BIOLEGEND (Beijing, China)
APC/Cyanine7 anti-mouse CD4 antibody	100413	BIOLEGEND
FITC anti-mouse IL-17A antibody	506907	BIOLEGEND
alpha-smooth muscle actin	19245S	Cell Signaling Technology (Danvers, USA)
Phospho-SGK1	5599S	Cell Signaling Technology
Acetyl-Histone H3	4353S	Cell Signaling Technology
β -ACTIN	4970S	Cell Signaling Technology
Tip60	ab73207	Abcam (Melbourne, VIC, Australia)
Collagen 1	ab34710	Abcam
Goat anti-mouse IgG (H+L) Highly Cross-Adsorbed secondary antibody (Alexa Fluor Plus 488)	UI287767	Thermo Fisher Scientific
Goat anti-rabbit IgG (H+L), F(ab') ₂ Fragment (Alexa Fluor 594 Conjugate)	8889S	Cell Signaling Technology