

Supplementary Materials for

Towards a “Testis in a Dish”: Generation of Mouse Testicular Organoids that Recapitulate Testis Structure and Expression profiles

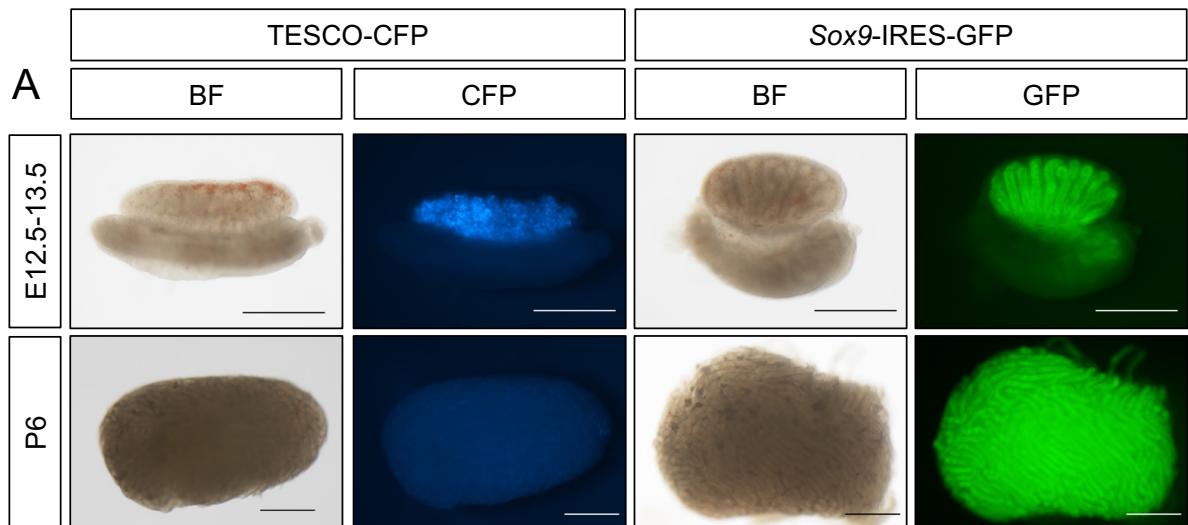
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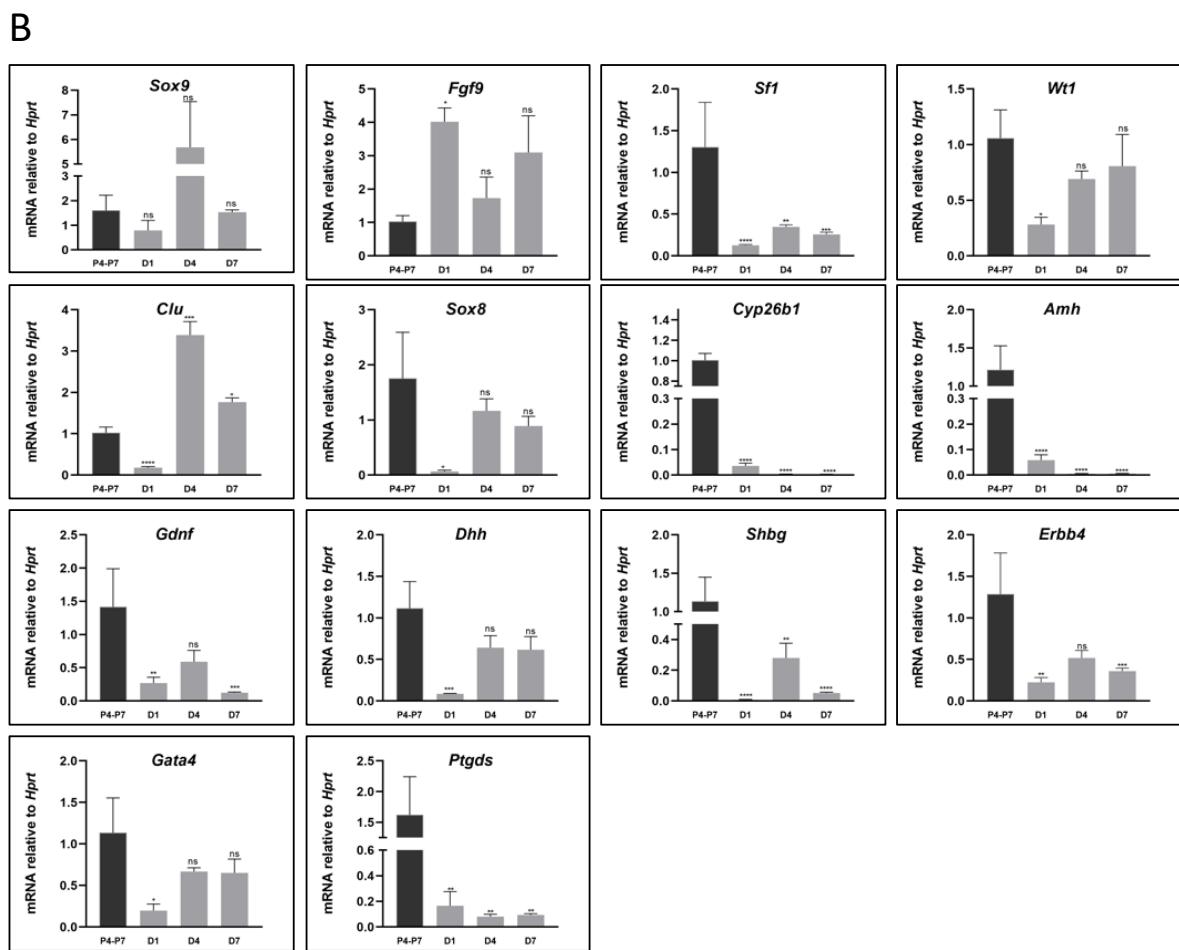
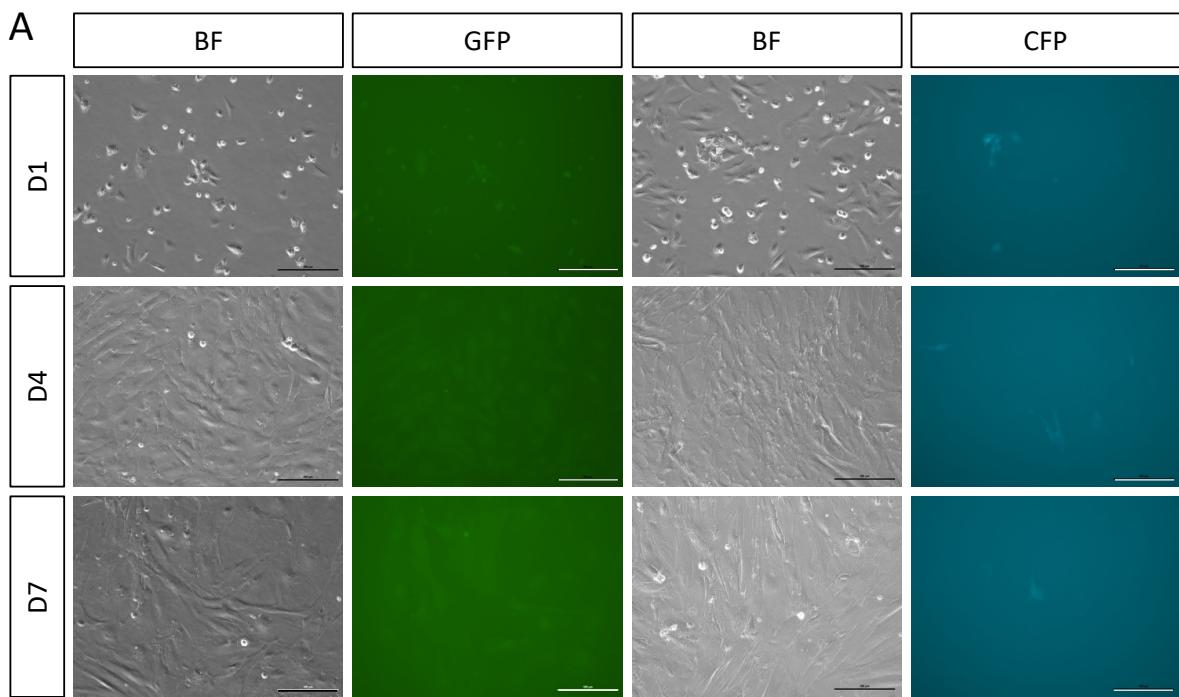
This PDF file includes:

Figs. S1 to S10
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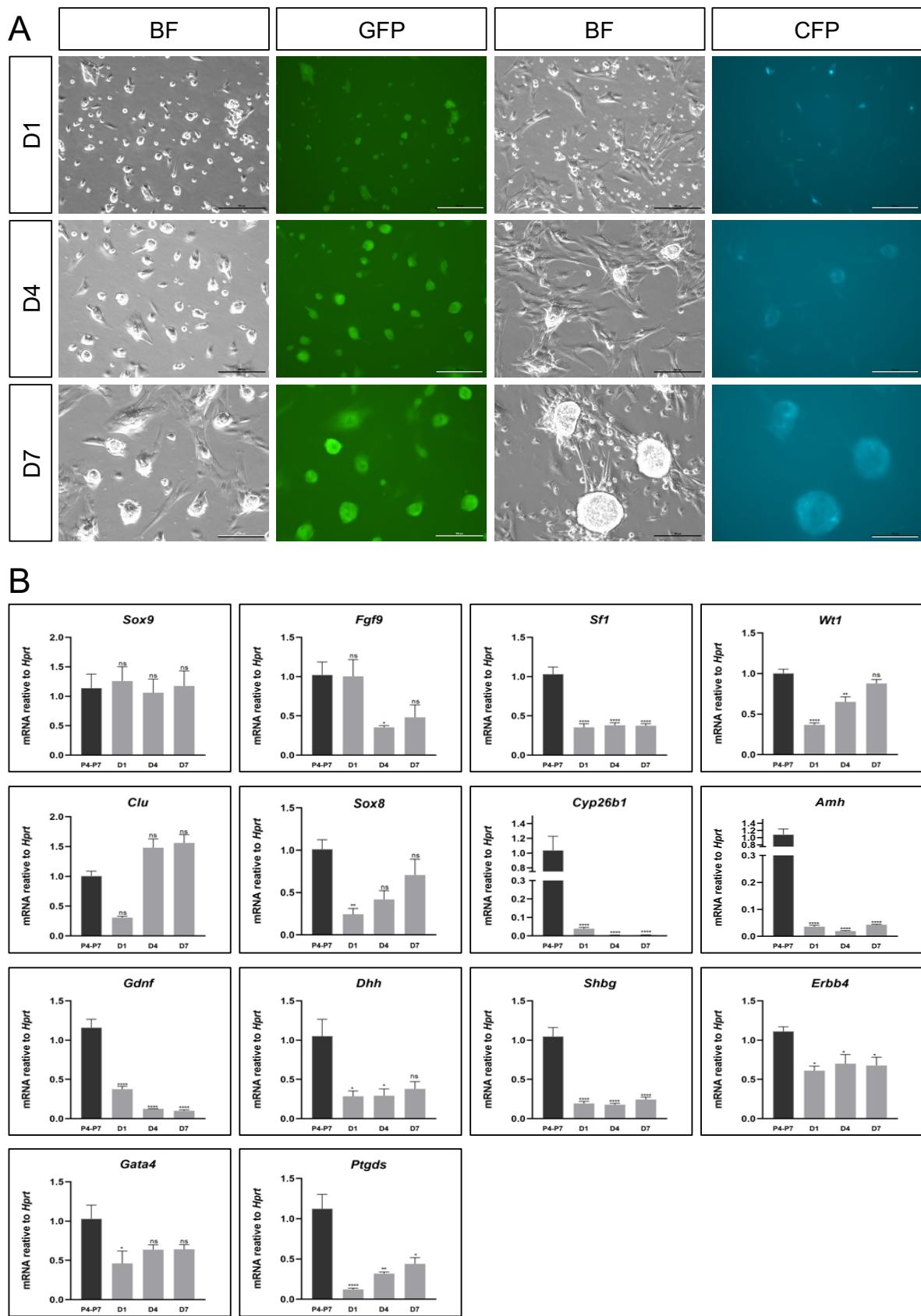
Other Supplementary Materials for this manuscript include the following: Movies S1 to S6



Supplementary Figure 1. Sertoli cell-specific reporter mouse lines used for testicular harvest. Bright-field (BF) and fluorescent images of testes from either E12.5-13.5 embryos or P6 pups harvested from *Sox9*-IRES-GFP or TESCO-CFP mice. Scale bars, 500 μ m.

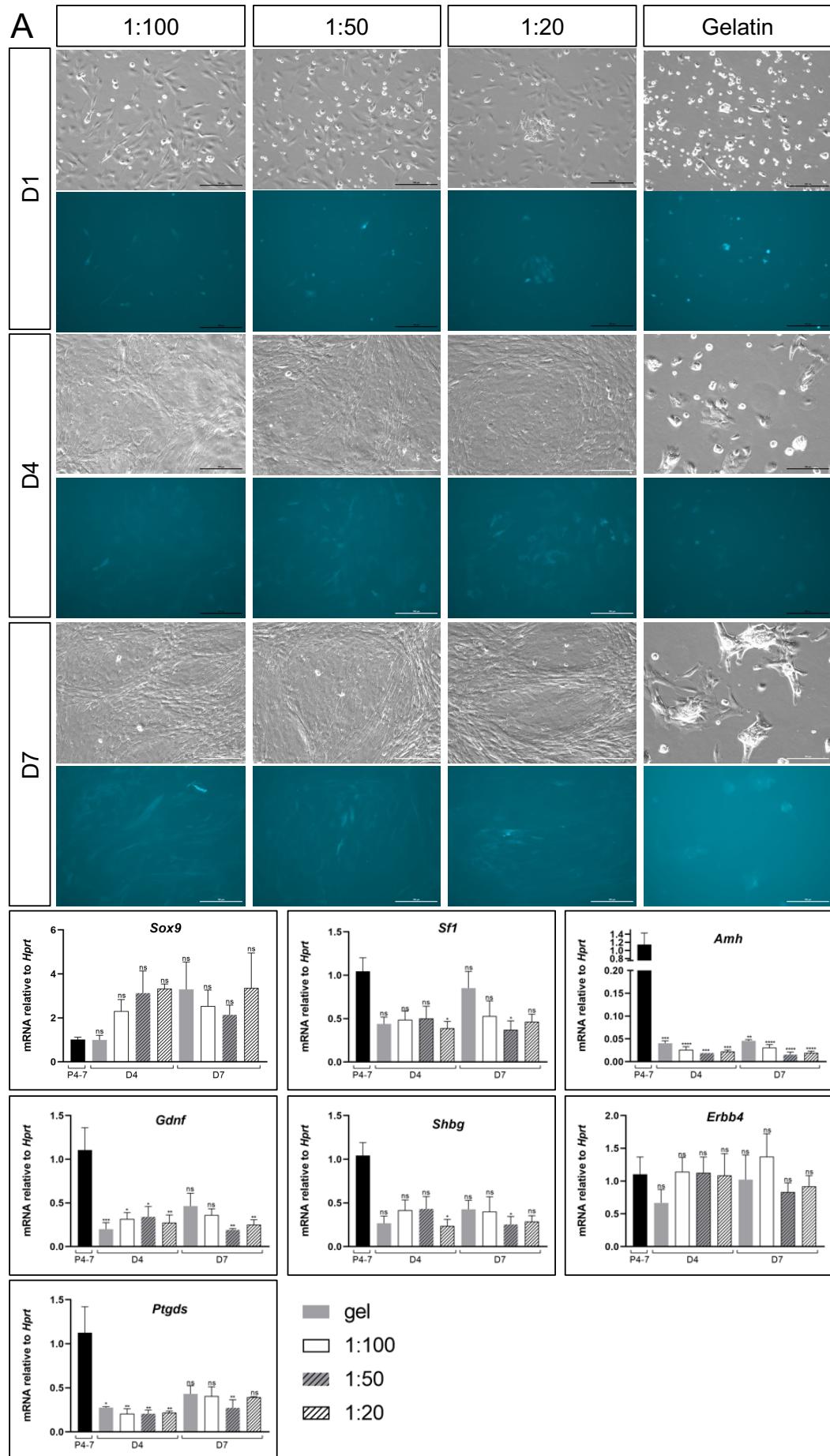


Supplementary Figure 2. Testicular cells cultured in 2D with serum-based media. (A) BF and fluorescent images of testicular cells harvested from either *Sox9*-IRES-GFP or TESCO-CFP pups. Cells were cultured for 7 days on 0.2% gelatin-coated dishes in Ad-DMEM/F12 media supplemented with 2% FBS. Scale bars, 100 μ m. (B) Quantitative PCR (qPCR) analysis of testicular cells from TESCO-CFP mice, cultured for 1-7 days in serum-based media. Data are presented as mean $2^{-\Delta\Delta C_t}$ values \pm SEM normalized to the housekeeping gene *Hprt*. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, and **** $P < 0.0001$, ns - not significant. $N=3$.



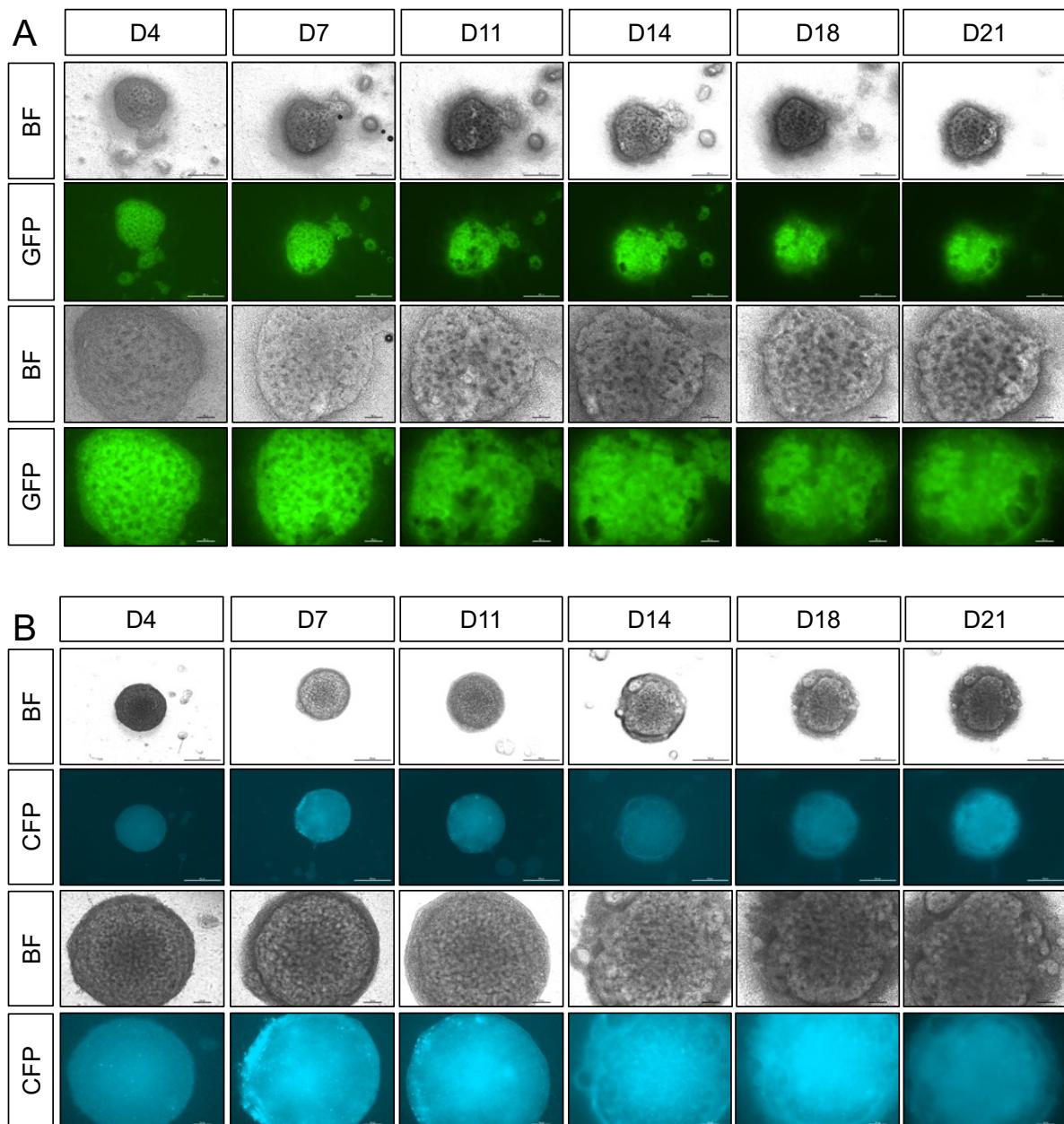
Supplementary Figure 3. Testicular cells cultured in 2D with testis-defined media. (A) BF and fluorescent images of testicular cells harvested from either *Sox9*-IRES-GFP or TESCO-

CFP pups. Cells were cultured for 7 days on 0.2% gelatin-coated dishes in defined media containing Ad-DMEM/F12 media supplemented with testicular growth factors as detailed in Supplementary Table 1. Scale bars, 100 μ m. (B) qPCR analysis of testicular cells from TESCO- CFP mice, cultured for 1-7 days with defined media. Data are presented as mean $2^{-\Delta\Delta C_t}$ values \pm SEM normalized to the housekeeping gene *Hprt*. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, and **** $P < 0.0001$, ns - not significant. $N=5$.

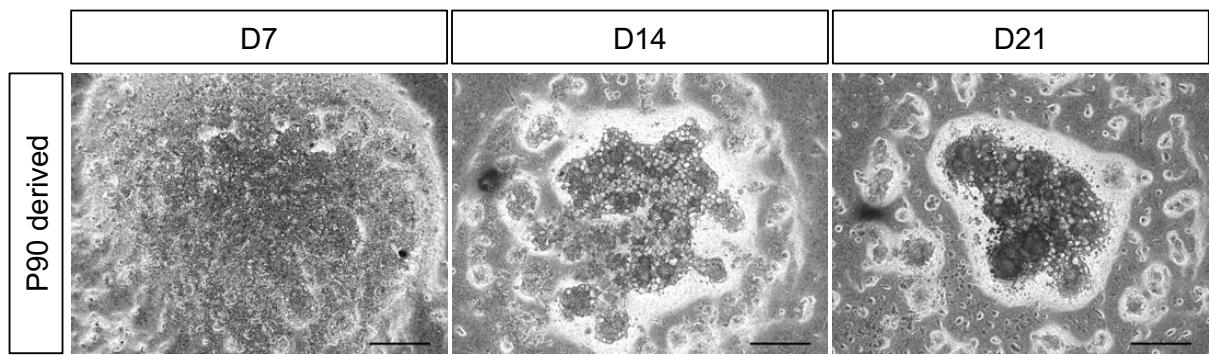


Supplementary Figure 4. Testicular cells cultured on Geltrex coating in defined media.

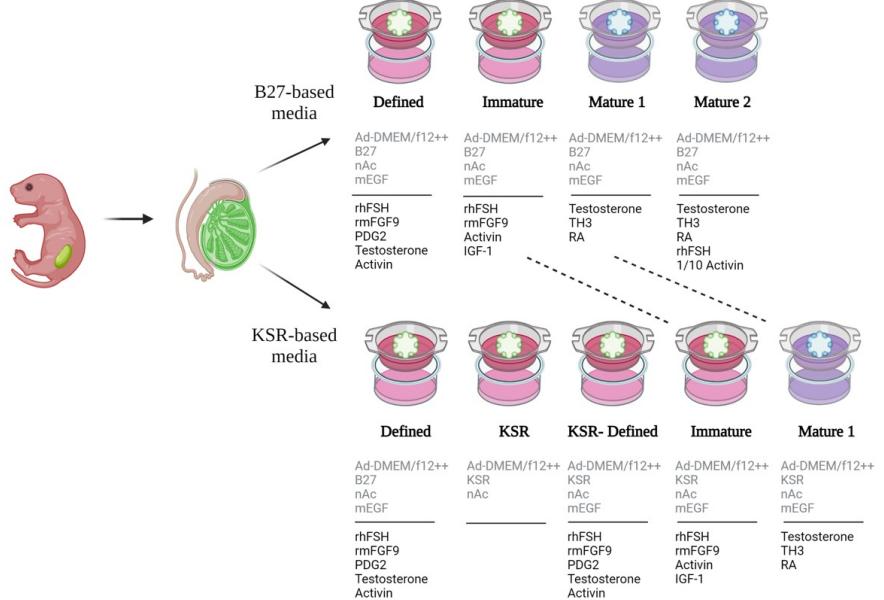
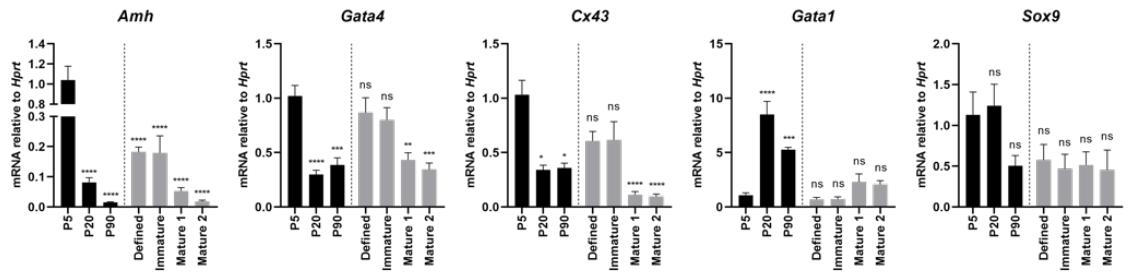
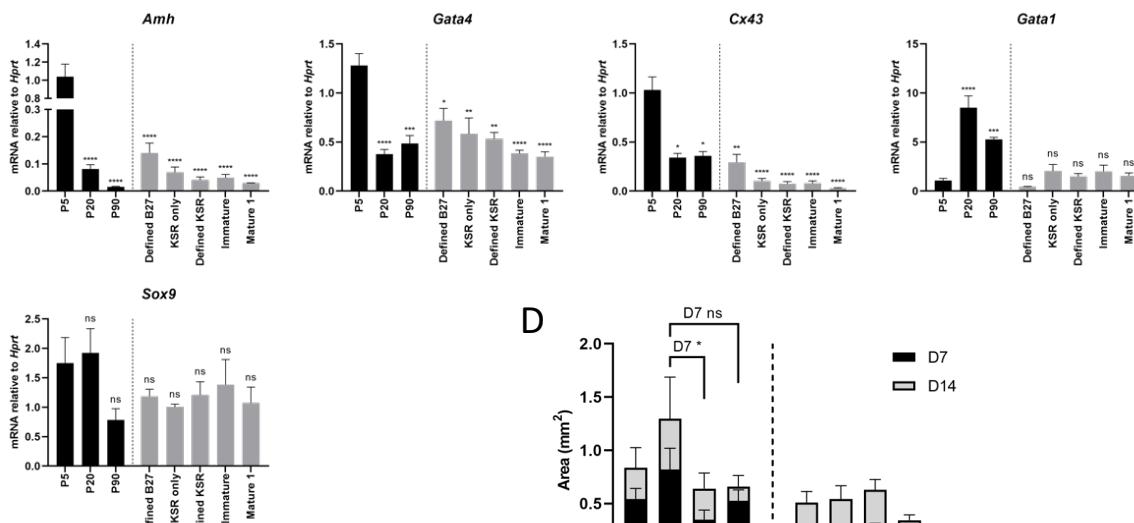
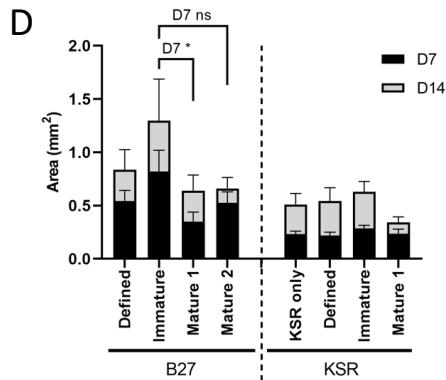
(A) BF and fluorescent images of testicular cells harvested from TESCO-CFP pups and cultured on various Geltrex dilutions (1:100, 1:50 and 1:20) or 0.2% gelatin (gel) coated wells. Scale bars, 100 μ m. (B) qPCR analysis of testicular cells from TESCO-CFP mice, cultured for 1-7 days on defined media and either Geltrex or Gelatin coating. Data are presented as mean $2^{-\Delta\Delta C_t}$ values \pm SEM normalized to the housekeeping gene *Hprt*. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, and **** $P < 0.0001$, ns - not significant. $N=5$.



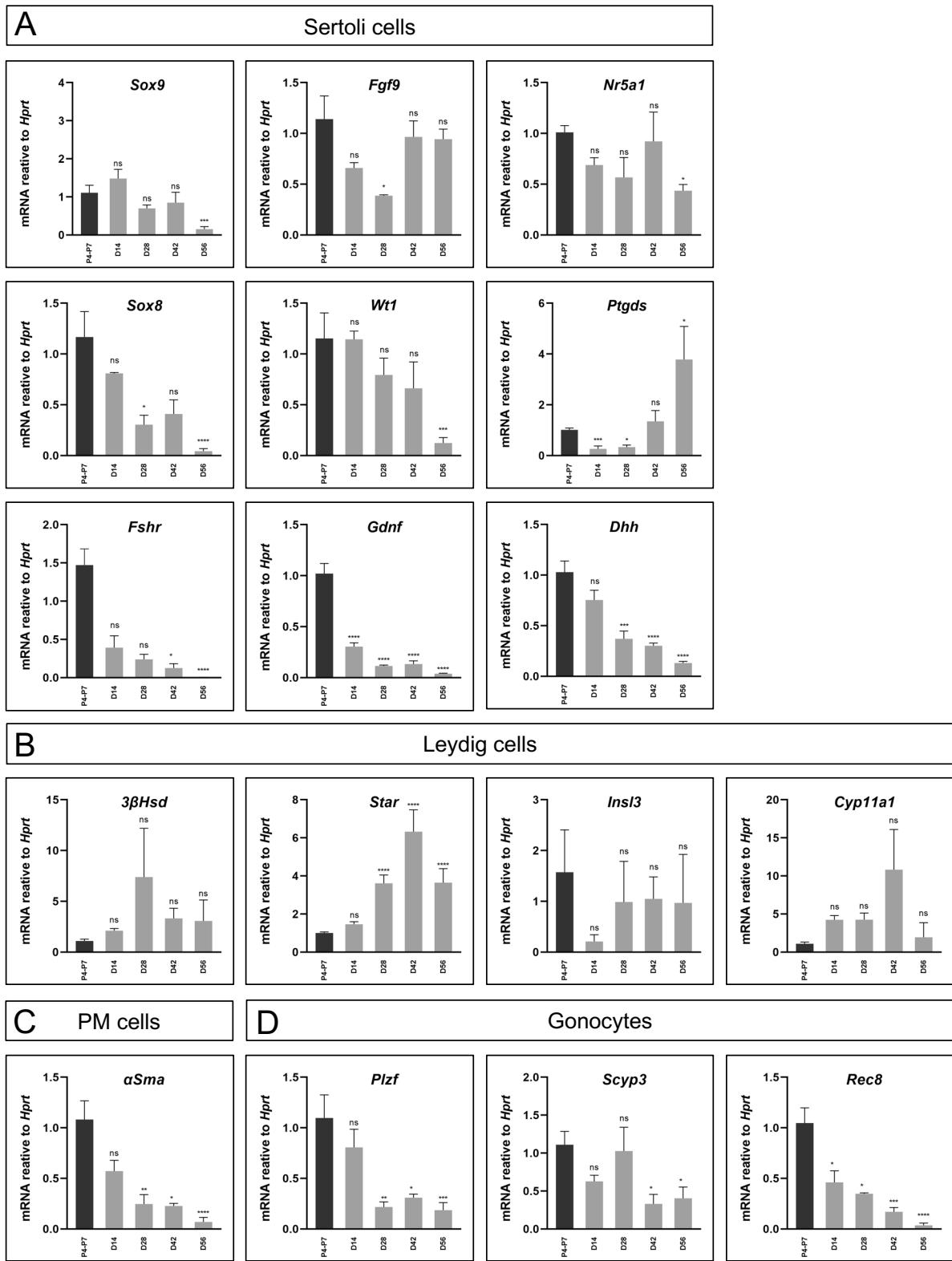
Supplementary Figure 5. Testicular neonatal organoids cultured on transwell inserts. (A) Representative BF and fluorescent images of organoids derived from *Sox9-IRES-GFP* testicular cells and cultured for up to 21 days on transwell inserts. (B) Representative BF and fluorescent images of organoids derived from TESCO-CFP testicular cells and cultured for up to 21 days on transwell inserts. Scale bars, 100 μ m.



Supplementary Figure 6. Organoids generated from mature testes. Representative BF images of organoids generated from P90 primary testicular cells, cultured for up to 21 days on transwell inserts in defined media. Scale bar, 500 μ m.

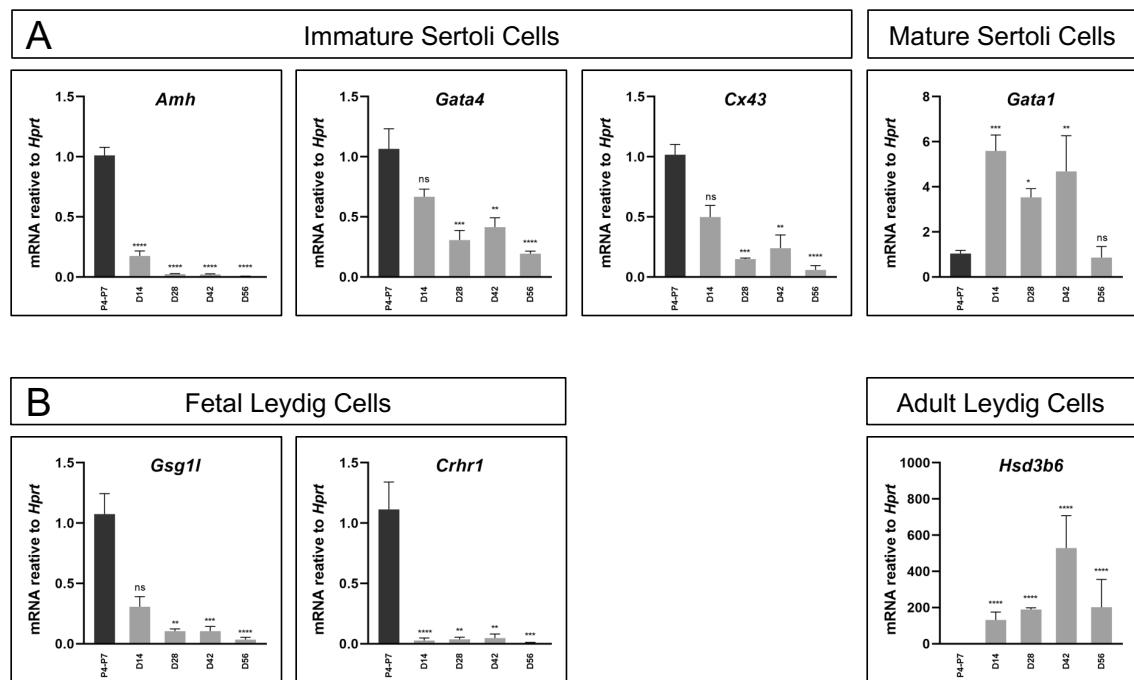
A**B B27-based media****C KSR-based media****D**

Supplementary Figure 7. Refinement of media composition for immature and mature state of testicular organoids. (A) Schematic representation of the surveyed media compositions on either B27-based or KSR-based media. The invariable additions are labelled in gray and the growth factors added to each media are labelled in black. For full media composition see Supplementary Table 1. Scheme created using BioRender. (B) qPCR analysis of Sertoli cell markers in organoids cultured in B27-based media for 14 days, compared to P5, P20 and P90 *in vivo* testis. Data are presented as mean $2^{-\Delta\Delta Ct}$ values \pm SEM normalized to the housekeeping gene *Hprt*. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, and **** $P < 0.0001$, ns - not significant. $N=4-5$. (C) qPCR analysis of Sertoli cell markers in organoids cultured on KSR-based media for 14 days, compared to P5, P20 and P90 *in vivo* testis. Data are presented as mean $2^{-\Delta\Delta Ct}$ values \pm SEM normalized to the housekeeping gene *Hprt*. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, and **** $P < 0.0001$, ns - not significant. $N=4-5$. (D) Organoid area on days 7 and 14 in B27- and KSR-based media. D7 bars are in black, D14 bars are cumulative and shaded grey. Data are presented as mean area \pm SEM. Statistical significance is shown for mature 1 and mature 2 vs. immature media at day 7 of culture. * $P < 0.05$, ns- not significant. $N=5-8$.

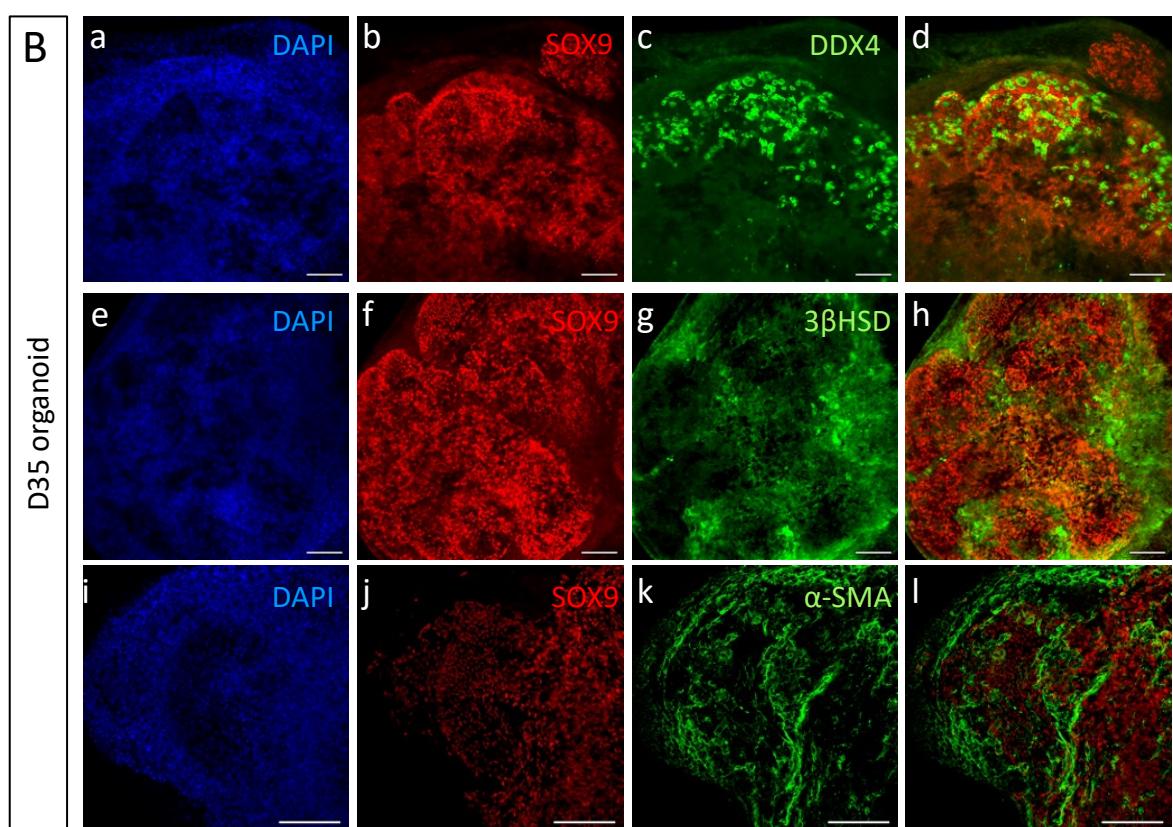
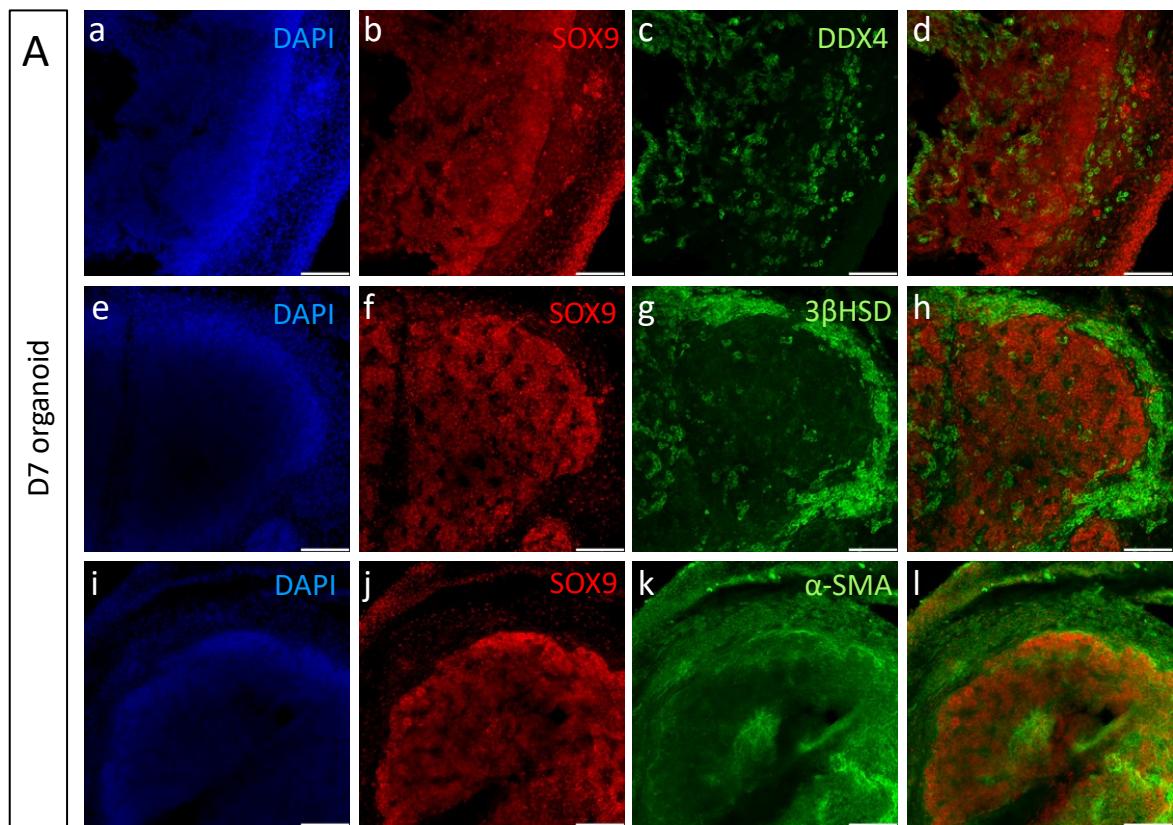


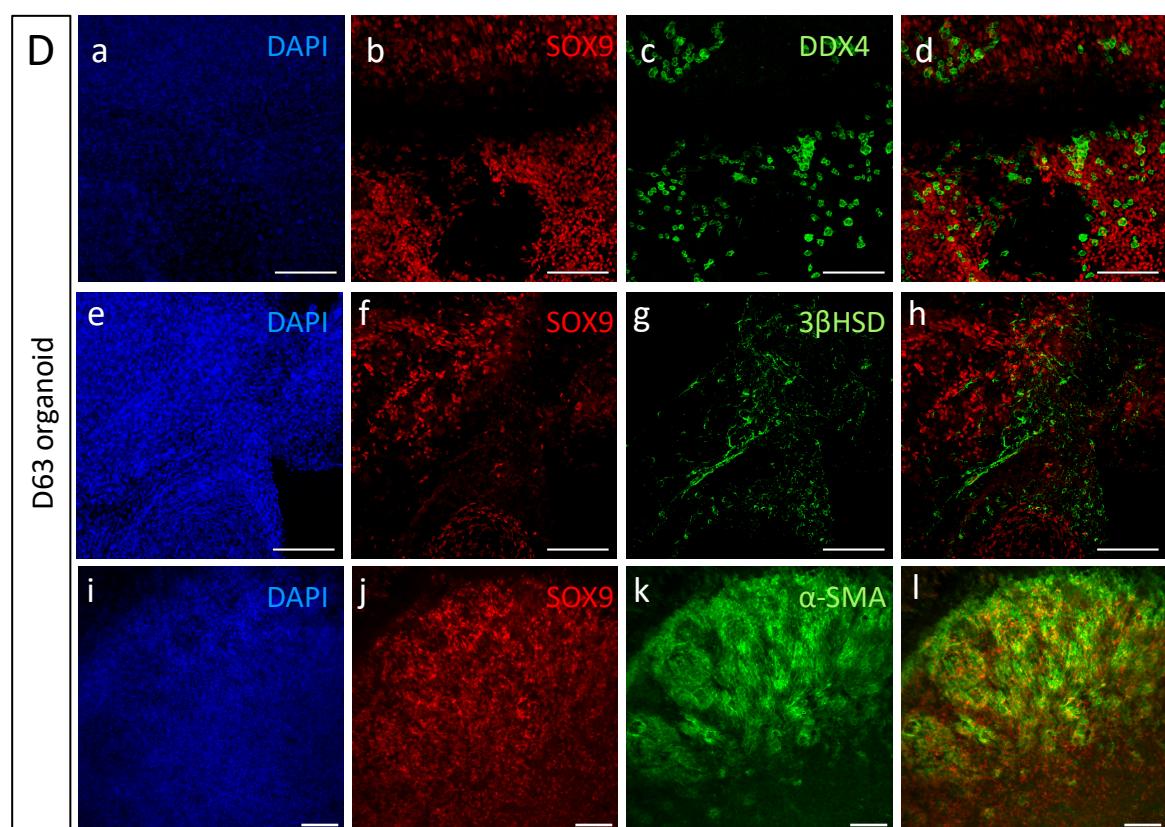
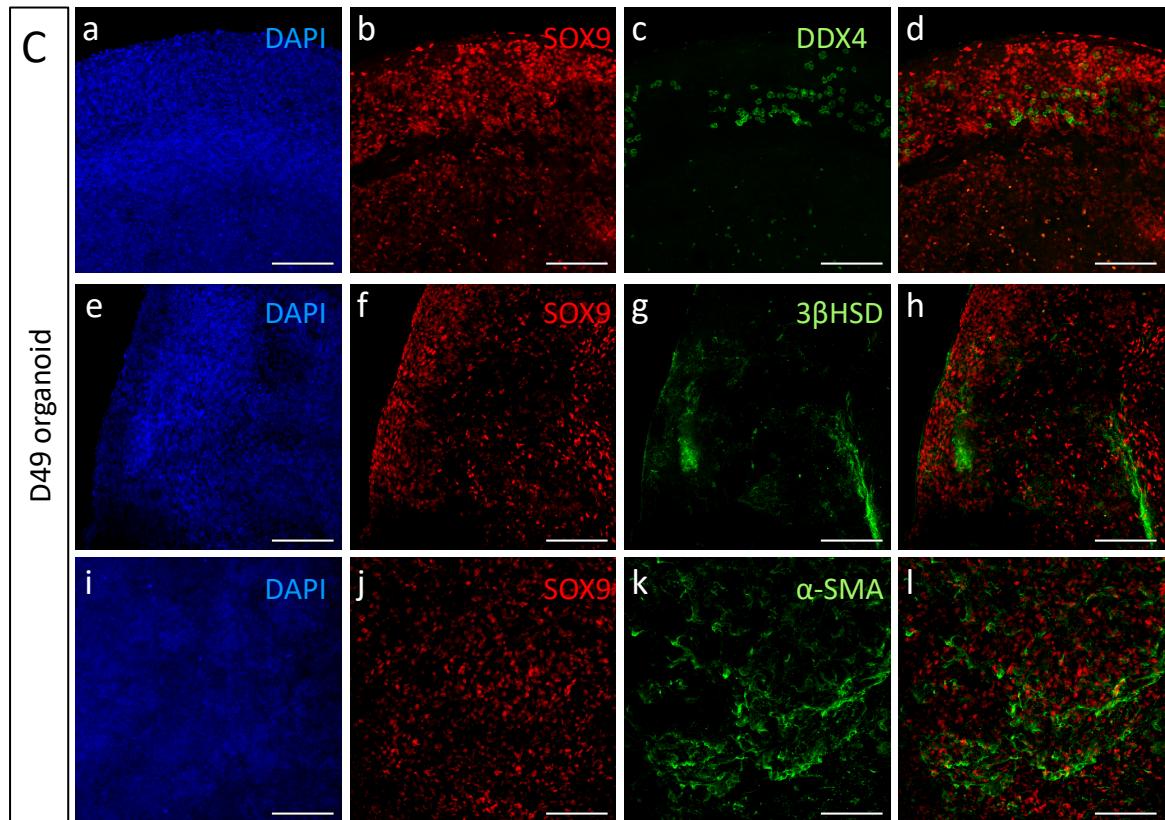
Supplementary Figure 8. Gene expression profiles of organoids cultured in transition media over time. (A-D) Quantitative RT-PCR was performed on mRNA extracted from neonatal testes, or TESCO-CFP organoids cultured for the indicated number of days on transition media assessing Sertoli cell markers (A), Leydig cell markers (B) or other testicular

cells markers (C-D). Gene names are presented in the title. D denotes culture day of organoids, grey. P denotes days postpartum (dpp) of *in vivo* testis, black. Data are presented as mean $2^{-\Delta\Delta Ct}$ values \pm SEM normalized to the housekeeping gene *Hprt*. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, and **** $P < 0.0001$, ns- not significant. $N=3-6$.



Supplementary Figure 9. Transition media promotes maturation of Sertoli and Leydig cells. (A-B) Quantitative RT-PCR was performed on mRNA extracted from neonatal testes, or TESCO-CFP organoids cultured for the indicated number of days on transition media assessing Sertoli cell immature vs. mature markers (A), or fetal Leydig cell vs. adult Leydig cell markers (B). Gene names are presented in the title. D denotes culture day of organoids, grey. P denotes days postpartum (dpp) of *in vivo* testis, black. Data are presented as mean $2^{-\Delta\Delta Ct}$ values \pm SEM normalized to the housekeeping gene *Hprt*. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, and **** $P < 0.0001$, ns- not significant. N=3-6.





Supplementary Figure 10. Testicular organoids on transition media preserve testicular structures and gonadal cell types for up to 9 weeks. (A-D) Whole mount co-immunostaining of markers for the major types of testicular cells in organoids cultured for 7 days (A), 35 days (B), 49 days (C) and 63 days (D). SOX9 marks Sertoli cells, DDX4 labels gonocytes, 3 β HSD marks Leydig cells and α -SMA labels PM cells. Images on the right are merged views of all channels. Scale bars, 100 μ m.

Table S1. Media composition

	Media	Component	Final concentration	Manufacturer	Cat. number
B27-based media	Serum based medium	Ad-DMEM-F12	/	Thermo Fisher Scientific	12634010
		Penicillin-Streptomycin	1x	Thermo Fisher Scientific	15140122
		L-glutamine	2mM	Biological industries	03-020
		Fetal Bovine Serum (FBS)	2%	Thermo Fisher Scientific	10270106
B27-based media	Defined medium	Ad-DMEM-F12	/	Thermo Fisher Scientific	12634010
		Penicillin-Streptomycin	1x	Thermo Fisher Scientific	15140122
		L-glutamine	2mM	Biological industries	03-020
		B-27 supplement	1X	Thermo Fisher Scientific	17504044
		N-Acetyl-L-cysteine	1.25mM	Sigma	A9165
		Mouse EGF	50ng/ml	Thermo Fisher Scientific	PMG8043
		Recombinant human FSH	22ng/ml	R&D Systems	5925-FS
		Recombinant mouse FGF9	50ng/ml	R&D Systems	7399-F9
		Prostaglandin D2 (PGD2)	0.5µg/ml	Cayman	12010
		Testosterone	1µM	Sigma	T1500
	Immature	Recombinant Human/Mouse/Rat Activin A	50ng/ml	R&D Systems	338- AC
		Ad-DMEM-F12	/	Thermo Fisher Scientific	12634010
		Penicillin-Streptomycin	1x	Thermo Fisher Scientific	15140122
		L-glutamine	2mM	Biological industries	03-020-1A
		B-27 supplement	1X	Thermo Fisher Scientific	17504044
		N-Acetyl-L-cysteine	1.25mM	Sigma	A9165
	Mature1	Mouse EGF	50ng/ml	Thermo Fisher Scientific	PMG8043
		Recombinant human FSH	22ng/ml	R&D Systems	5925-FS
		Recombinant mouse FGF9	50µg/ml	R&D Systems	7399-F9
		Recombinant Human/Mouse/Rat Activin A	50ng/ml	R&D Systems	338- AC
		Recombinant human IGF-1	100ng/ml	R&D Systems	291-G1
		Ad-DMEM-F12	/	Thermo Fisher Scientific	12634010
		Penicillin-Streptomycin	1x	Thermo Fisher Scientific	15140122
		L-glutamine	2mM	Biological industries	03-020-1A
		B-27 supplement	1X	Thermo Fisher Scientific	17504044
		N-Acetyl-L-cysteine	1.25mM	Sigma	A9165
KSR-based media	Mature2	Mouse EGF	50ng/ml	Thermo Fisher Scientific	PMG8043
		Testosterone	1µM	Sigma	T1500
		Thyroid hormone (T3)	100nM	Tocris	6666
		Retinoic acid	1µM	Sigma	R2625
		Recombinant Human/Mouse/Rat Activin A	5ng/ml	R&D Systems	338- AC
	KSR	Ad-DMEM-F12	/	Thermo Fisher Scientific	12634010
		Penicillin-Streptomycin	1x	Thermo Fisher Scientific	15140122
		L-glutamine	2mM	Biological industries	03-020-1A
		KnockOut Serum Replacement (KSR)	10%	Thermo Fisher Scientific	10828028
		N-Acetyl-L-cysteine	1.25mM	Sigma	A9165
	KSR - Defined	Ad-DMEM-F12	/	Thermo Fisher Scientific	12634010
		Penicillin-Streptomycin	1x	Thermo Fisher Scientific	15140122
		L-glutamine	2mM	Biological industries	03-020-1A
		KnockOut Serum Replacement (KSR)	10%	Thermo Fisher Scientific	10828028
		N-Acetyl-L-cysteine	1.25mM	Sigma	A9165
		Mouse EGF	50ng/ml	Thermo Fisher Scientific	PMG8043
		Recombinant human FSH	22ng/ml	R&D Systems	5925-FS
		Recombinant mouse FGF9	50µg/ml	R&D Systems	7399-F9
		Prostaglandin D2 (PGD2)	0.5µg/ml	Cayman	12010
		Testosterone	1µM	Sigma	T1500
		Recombinant Human/Mouse/Rat Activin A	50ng/ml	R&D Systems	338- AC
KSR-based media	Immature	Ad-DMEM-F12	/	Thermo Fisher Scientific	12634010
		Penicillin-Streptomycin	1x	Thermo Fisher Scientific	15140122
		L-glutamine	2mM	Biological industries	03-020-1A
		KnockOut Serum Replacement (KSR)	10%	Thermo Fisher Scientific	10828028
		N-Acetyl-L-cysteine	1.25mM	Sigma	A9165
		Mouse EGF	50ng/ml	Thermo Fisher Scientific	PMG8043
		Recombinant human FSH	22ng/ml	R&D Systems	5925-FS
		Recombinant mouse FGF9	50µg/ml	R&D Systems	7399-F9
		Recombinant Human/Mouse/Rat Activin A	50ng/ml	R&D Systems	338- AC
		Recombinant human IGF-1	100ng/ml	R&D Systems	291-G1
	Mature1	Ad-DMEM-F12	/	Thermo Fisher Scientific	12634010
		Penicillin-Streptomycin	1x	Thermo Fisher Scientific	15140122
		L-glutamine	2mM	Biological industries	03-020-1A
		KnockOut Serum Replacement (KSR)	10%	Thermo Fisher Scientific	10828028
		N-Acetyl-L-cysteine	1.25mM	Sigma	A9165
		Mouse EGF	50ng/ml	Thermo Fisher Scientific	PMG8043

Table S2. Primers for genotyping mice

Primer name	Description	Sequence 5' to 3'	Product size
CFP-F	CFP	CGTGACCACCTGACCTGG	327 bp product
CFP-R	CFP	GTGGCGGATCTTGAAGTTGG	
GFP-F	GFP	TGCAGTGCTTCAGCCGCTAC	462 bp product
GFP-R	GFP	CCAGCAGGACCATGTGATCC	

Table S3. Primers for Real-time quantitative RT-PCR

Primer name	Gene	Marker of	Sequence 5' to 3'
m3βHsd_F	3βHsd	Leydig cells	CTCAGTTCTTAGGCTTCAGCAATTAC
m3βHsd_R			CCAAAGGCAGATATGATTAGGA
mAcrosin_F	Acrosin	Mature sperm	TGTCCGTGGTGCAGGATAACA
mAcrosin_R			AATCCGGGTACCTGCTTGAGTT
mAmh_F	Amh	Immature Sertoli cells	CCACGGTTAGCACCAAATAGC
mAmh_R			CACACAGAACCTCTGCCCTACTC
mAr_F	Ar	Sertoli cells	CTGGGAAGGGTCTACCCAC
mAr_R			GGTGCTATGTTAGCGGCCTC
mClu_F	Clu	Sertoli cells	TGAAGGGCCAGTGTGAAAAGT
mClu_R			TTGAACAGTCCACAGACAAGATCTC
mCrhr1_F	Crhr1	Fetal Leydig cells	CCAGGATCAGCAGTGTGAGA
mCrhr1_R			TGTTTGTTGTTGAGCGG
mCx43_F	Cx43	Adult Sertoli cell	GGTCTGAGAGCCCCGAACTCTCCT
mCx43_R			CCCATGTCGGGCACCTCTCTT
mCyp11a1_F	Cyp11a1	Leydig cells	CACTTCTGGAGGGAGAGTGG
mCyp11a1_R			ATGCCTGGAAGAAAGACCGA
mCyp26b1_F	Cyp26b1	Sertoli cells/FLC	AAGGGCTCATGGGATTC
mCyp26b1_R			ACGACTGGAAGCCGGAAC
mDhh_F	Dhh	Sertoli cells	CCGCAACCACATCCACGTA
mDhh_R			CGGACCGCCAGTGAGTTATC
mErbb4_F	Erbb4	Sertoli cells	CAGCGTTCTCAGTCAGTGT
mErbb4_R			CTGCTGTTCCAGGTAGAGA
mFgf9_F	Fgf9	Sertoli cells	TCTTCCCACCGGTACTATCCA
mFgf9_R			CGAAGCGCTGTGGTCTTT
mFshr_F	Fshr	Sertoli cells	TTGCCTGATGATGTTTCCA
mFshr_R			CTGGCCCTCAACTTCTTCAG
mGata1_F	Gata1	Adult Sertoli cell	TGGGGACCTCAGAACCCCTG
mGata1_R			GGCTGCATTGGGAAAGTG
mGata4_F	Gata4	Sertoli cells	CCCCAATCTGATATGTTGATG
mGata4_R			TTGACACACTCTGCCTCTGA
mGdnf_F	Gdnf	Sertoli cells	GGGTGCGTTAACGACTGCCATA
mGdnf_R			GCCAAACCCAAGTCAGTGA
mGsg11_F	Gsg11	Fetal Leydig cells	GAGGAGCCGACGTTCATAGA
mGsg11_R			AGCACTGACGGTTAGCTCT
mHprt_F	Hprt	HK gene	GCAGTACAGCCCCAAATGG
mHprt_R			GGTCCTTTCAACCAGCAAGCT
mHsd3b6_F	Hsd3b6	Adult Leydig cells	GGAGGAGATCAGGGCTCTGG
mHsd3b6_R			CTAGGATGGTCTGCCTGG
mIns13_F	Ins13	Leydig cells	GCTGCTACTGATGCTCTGG
mIns13_R			CTAGACCCGCACTTCCTC
mNr5a1_F	Nr5a1	Sertoli cells	CCTCGATGTGAAATTCTGAACA
mNr5a1_R			TCCTGGCGTCCTTACG
mPlzf_F	Plzf	Spermatogonial stem cells (SSC)	CCAGTTGGAGACGACCTACA
mPlzf_R			GTGGCAGAGTTGCACTCAA
mPtgds_F	Ptgds	Sertoli cells	GGCTCCTGGACACTACACCT
mPtgds_R			CATAGTTGGCCTCCACCACT
mRec8_F	Rec8	Meiotic spermatids	GCCCTAGAAGGTGCTGGTTGG
mRec8_R			GTGGGGTCACCTCAGTGAGTAGG
mScyp3_F	Scyp3	Meiotic spermatids	GCAGTCTAGAATTGTTCAGAGCCAGA
mScyp3_R			TCCAAACTCTTATGAACTGCTCGTG
mShbg_F	Shbg	Sertoli cells	CCAAAATCAGCAAACCCATT
mShbg_R			CTGGATCCCAGGTTCGAAC
mSox8_F	Sox8	Sertoli cells	AGCGAGAAGAGGCCGTTG
mSox8_R			TCAGTACAGAGTCTGAGTCG
mSox9_F	Sox9	Sertoli cells	AAGAAAGACCACCCGATTACA
mSox9_R			CAGCGCCTTGAAGATAGCATT
mStar_F	Star	Leydig cells	GACGTGGAGCTCTGCTT
mStar_R			GCCTCTGCATAGCCACCTC
mWt1_F	Wt1	Sertoli cells	TTGAATGCATGACCTGGAATCA
mWt1_R			TTCCCTTAAGGTAGCTCCTAGGTT
moSma_F	aSma	Peritubular myoid cells (PMC)	CCCTGAAGAGCATCCGACA
moSma_R			TGGCGGGACATTGAAGGT

Table S4. Antibody list

Protein	Cat. No.	Raised in	Company	Dilution
SOX9	AB5535	Rabbit	Merck Millipore	1:5000
DDX4	ab27591	Mouse	Abcam	1:300
α SMA	A2547	Mouse	Sigma	1:200
3β HSD	sc-515120	Mouse	Santa Cruz	1:200
γ H2AX	ab11174	Rabbit	Abcam	1:1000
TRA98	ab82527	Rat	Abcam	1:200
AMH	sc-6886	Goat	Santa Cruz	1:200
CLAUDIN-11	Zy-364500	Rabbit	Invitrogen	1:100
REC8	ab192241	Rabbit	Abcam	1:200

Movie S1. Supplementary movie 1_D21 organoid_SOX9_DDX4 b_d

Movie S2. Supplementary movie 2_D21 organoid_SOX9_3bHSD f_h

Movie S3. Supplementary movie 3_D21 organoid_SOX9_SMA j_1

Movie S4. Supplementary movie 4_D21 organoid_AMH_CLD11_DDX4 m_p

Movie S5. Supplementary movie 5_D21 organoid_SOX9

Movie S6. Supplementary movie 6_D21 organoid_DAPI