Supporting Information

Coupling of glucose metabolism with mitophagy via O-GlcNAcylation of PINK1

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Figure S1. Increased glucose uptake and O-GlcNAcylation in WT HeLa cells following O/A-induced mitophagy. (**A and B**) Flow cytometry assay of glucose uptake by WT HeLa cells in the 2-NBDG assay. (**C**) Flow cytometric analysis of intracellular O-GlcNAc levels in YFP-HeLa treated with CCCP (20 μ M) for 0.5 to 4 h. (**D**) YFP-HeLa cells were pretreated with or without Bafilomycin A1 (BafA1,100nM) for 1 h. Subsequently, the cells were treated with or without O/A (1 μ M and 1 μ M) for 0.5, 1, 2h

and subjected to western blotting analysis with the indicated antibodies. (**E**) Topology assay showing OGT localization at the OMM. Purified mitochondria were isolated from YFP-Parkin-HeLa cells and treated with the indicated reagents of proteinase K and digitonin. *p < 0.05.



Figure S2. OGA KO did not regulate mitophagy. Mcherry-Parkin expressing control and OGA KO cells HeLa cells treated with 1 μ M O/A, then harvested at the indicated time points for western blotting analysis with the indicated antibodies.



Figure S3. OGT KO reduced Parkin mitochondrial translocation. (**A and B**) Colocalizations between mCherry-Parkin and GFP-mitochondrial, YFP-Parkin and mitochondrial marker HSP60 as in (Fig 3D and 3F) were analyzed by Mander's overlap coefficient. (**C**) Control and OGT-KO HeLa stably expressing mCherry-Parkin upon treatment with or without O/A for 2h. Representative immunofluorescence images of WT and OGT-KO cells. Scale bar=10µm. (**D**) Colocalization between mCherry-Parkin

and GFP-mitochondrial as in (Fig S3C). (**E**) Control or OGT KD cells were lysed for western blotting analysis to confirm the knockdown effects of OGT. (**F**) JC-1 staining and quantification of Ctrl and OGT KD HEK293T treated with 1 μ M O/A or DMSO for 6h. **, p < 0.01, N.S., no significance.



Figure S4. Colocalization between mcherry-Parkin and GFP-mitochondrial as in (Fig 4B and 4D) was analyzed by Mander's overlap coefficient. (N.S., no significance; *, P < 0.1, **, P < 0.01; Two-way ANOVA with Sidak's multiple comparisons test).



Figure S5. PINK1 undergoes O-GlcNAcylation. (**A and B**) HEK293T and YFP-HeLa cells following O/A for the indicated time were subjected to immunoprecipitation with anti-O-GlcNAc antibody. Then the immunoprecipitation was blotted with PINK1 antibody and lysates were subjected to O-GlcNAc antibody respectively.

SeqName	Residue		0-GlcNAc result	Potential (o-glcnac)
NP 115785.1	22		+++	0.2932
NP 115785.1	73	s	+++	0.3181
NP 115785.1	118	S	+++	0.2733
NP 115785.1	123	s	+++	0.5152
NP 115785.1	133	т	+++	0.3832
NP 115785.1	136	Ś	+++	0.3192
NP 115785 1	145	т	+++	0.3192
NP_115705.1	161	ċ	+++	0.3103
NP_115705.1	167	2	+++	0.2017
NP_115705.1	174	э т	+++	0.2700
NP_115705.1	174	÷	+++	0.2371
NP_115/85.1	105	÷	+++	0.2923
NP_115/85.1	185	1	+++	0.4256
NP_115/85.1	187	5	+++	0.2909
NP_115/85.1	188	+	+++	0.2929
NP_115785.1	198	T	+++	0.4757
NP_115785.1	199	S	+++	0.3143
NP_115785.1	225	S	+++	0.3586
NP_115785.1	228	S	+++	0.3546
NP_115785.1	229	S	+++	0.3112
NP_115785.1	230	S	+++	0.3480
NP_115785.1	236	Т	+++	0.3678
NP_115785.1	238	S	+++	0.3788
NP_115785.1	245	S	+++	0.3743
NP_115785.1	257	Т	+++	0.4696
NP_115785.1	261	S	+++	0.2329
NP_115785.1	282	т	+++	0.5008
NP_115785.1	283	S	+++	0.5747
NP 115785.1	284	S	+++	0.3839
NP 115785.1	301	S	+++	0.2168
NP 115785.1	313	T	+++	0.2176
NP 115785.1	324	T	+++	0.3166
NP 115785.1	333	Ť	+++	0.3340
NP 115785.1	335	ŝ	+++	0.2933
NP 115785.1	365	ŝ	+++	0.2255
NP 115785 1	303	s	+++	0.2234
ND 115795 1	401	ç	+++	0.2955
NP_115705.1	401	5	+++	0.3334
NP_115/85.1	402	2	+++	0.3498
NP_115785.1	419	3	+++	0.7224
NP_115/85.1	420	1	+++	0.4/13
NP_115785.1	432	S	+++	0.3413
NP_115785.1	463	S	+++	0.2361
NP_115785.1	465	S	+++	0.2813
NP_115785.1	477	S	+++	0.4912
NP_115785.1	495	S	+++	0.5014
NP_115785.1	499	S	+++	0.2598
NP_115785.1	510	S	+++	0.1658
NP_115785.1	535	S	+++	0.3341
NP_115785.1	538	т	+++	0.2207
NP_115785.1	545	т	+++	0.3223
NP 115785.1	552	т	+++	0.2053
NP 115785.1	566	Ť	+++	0.2546

Figure S6. The predicted sites of PINK1 could be modified by O-GlcNAcylation. Computational O-GlcNAcylation sites of PINK1 predicted via YinOYang prediction program. This prediction program characterizes PINK1 O-GlcNAcylation over 50 different sites, where its prediction strengths are marked with high ["+++"], ranging from low to high ["+" to "++++"] in the program. The prediction strength depends on the sequence condition and conformation.



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Figure S7. S229A and S225A mutant suppress PINK1-Parkin-mediated mitophagy. (**A**) PINK1 KO cells transfected with WT PINK1 or S229A mutant were treated with or without O/A for 2h. (**B**) Colocalization between mCherry-Parkin and GFP-mitochondrial as in (Fig S7A) was analyzed. (**C and D**) TMRE staining and quantification of WT PINK1 and S229A mutant overexpressing YFP-HeLa PINK1 KO cells treated with 1 μ M O/A or DMSO for 2 and 4 h. (**E**) PINK1 WT and S225A were transfected in HeLa PINK1-/- stably expressing mCherry-Parkin cells, time course for mitochondrial proteins detection upon treatment with O/A. *p < 0.05. (**F**) Colocalization between mcherry-Parkin and GFP-mitochondrial as in (Fig 7C) was analyzed by Mander's overlap coefficient. (N.S., no significance; **, P < 0.01; Two-way ANOVA with Sidak's multiple comparisons test. GS, glucose starvation, Glc-re, glucose re-supplement). Scale bar=10 μ m.

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