

**Figure S1. Pyroptosis is involved in acute liver injury induced by TAA.** (A) H&E staining in liver sections (Scale bar: 50μm, 25μm) and serum ALT and AST levels of control group and TAA group (n = 6 per group). (B) IHC staining of GSDMD-N and Cleaved CASPASE1 in liver sections from indicated groups (Scale bar: 50μm, n = 6 per group). (C) Protein expression of GSDMD-N, Cleaved CASPASE1, mature IL-1β and mature IL-18 in mouse livers treated with PBS or TAA (n = 3 per group). (D) Ultrastructural features identified by TEM under control and TAA treatment (Scale bar: 5μm, 2μm, n = 6 per group). (E, F) TUNEL/GSDMD-N co-staining (Scale bar: 25μm) and IF staining of mature IL-1β and mature IL-18 (Scale bar: 50μm) in liver sections from indicated groups (n = 6 per group). (G) mRNA levels of *Il-1β*, *Il-18*, *Tnf-α*, *Tgf-β*, *Il-6* and *Mpo* in liver tissues were determined by RT-qPCR (n = 3 per group). (H) The levels of CASPASE1 activity, IL-1 and IL-18 in serum from indicated group were measured (n = 6 per group). (I) Phase contrast micrographs of primary hepatocytes from mice treated with TAA at 0, 6, 12, 24, and 48 hours (Scale bar: 50μm). (J) LDH activity in the cell culture supernatant at different timeline from indicated group (n = 3 per group). (K) PI/Cleaved CASPASE1 co-staining of primary hepatocytes at different timeline of TAA treatment (Scale bar: 50μm, n = 3 per group). All results are shown as mean ± SEM. \*p < 0.05.

**Figure S2. Hepatocyte pyroptosis is involved in acute liver injury.** (A) Signaling pathways related to inflammation and cell death were enriched by GO analysis in the primary hepatocytes from CCl<sub>4</sub>-treated mice. (B, C) Protein levels of GSDMD-N, Cleaved CASPASE1, mature IL-1β and mature IL-18 in primary hepatocytes under control and CCl<sub>4</sub> treatment (n = 3 per group). (D) mRNA levels of *Il-1β*, *Il-18*, *Tnf-α*, *Tgf-β*, *Il-6* and *Mpo* in primary hepatocytes from indicated groups (n = 3 per group). (E) CASPASE1 activity and the IL-1β and IL-18 level in the cell culture supernatant (n = 6 per group). All results are shown as mean ± SEM. \*p < 0.05.

**Figure S3. Hepatocyte pyroptosis is regulated by METTL3.** (A) DNA gel electrophoresis confirming successful construction of the *Mettl3*-Mut mice. (B) Protein expression of METTL3 in liver tissues and primary hepatocytes from *Mettl3*-WT and *Mettl3*-Mut control mice (n = 3 per group). (C) mRNA level of *Mettl3* in liver tissues and primary hepatocytes from *Mettl3*-WT and *Mettl3*-Mut control mice (n = 3 per group). (D) Protein expression of METTL3 and NLRP3 in primary hepatocytes. (E) Protein levels of METTL3, NLRP3, GSDMD-N, Cleaved CASPASE1, mature IL-1β and mature IL-18 in primary hepatocytes isolated from *Mettl3*-WT and *Mettl3*-Mut mice under CCl<sub>4</sub> treatment (n = 3 per group). (F) mRNA levels of *Mettl3*, *Il-1β*, *Il-18*, *Tnf-α*, *Tgf-β*, *Il-6* and *Mpo* in primary hepatocytes from indicated groups (n = 3 per group). (G)

Supernatant pyroptosis index of primary hepatocytes in indicated groups (n = 6 per group). **(H)** LDH activity in the culture supernatant of primary hepatocytes at different timeline (n = 6 per group). **(I)** Protein levels of METTL3, NLRP3, GSDMD-N and Cleaved CASPASE1 in primary hepatocytes transfected with si-*Mettl3* or control siRNA from control or CCl<sub>4</sub>-treated mice (n = 3 per group). All results are shown as mean ± SEM. \*p < 0.05. ns, not significant.

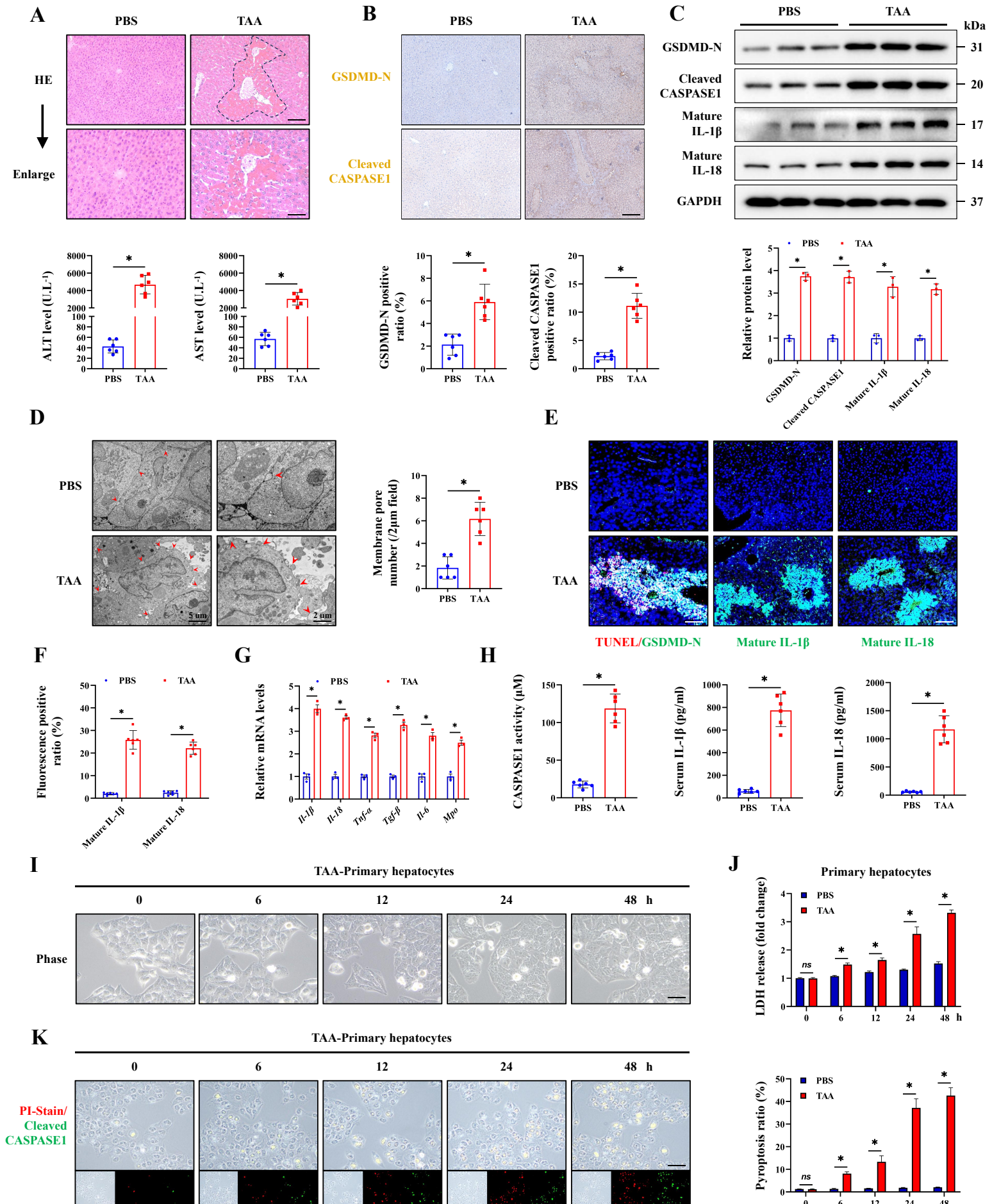
**Figure S4. Inhibition of DLGAP5 can effectively alleviate pyroptosis and acute liver injury.** **(A)** Molecular docking of AT9283 to DLGAP5 protein. **(B)** H&E staining in liver sections of ALI mice with or without AT9283 treatment (Scale bar: 50μm). **(C)** Serum ALT and AST levels from indicated groups (n = 6 per group). **(D)** mRNA levels of *Il-1β*, *Il-18*, *Tnf-α*, *Tgf-β*, *Il-6* and *Mpo* in liver tissues of above groups (n = 3 per group). **(E)** AT9283 treatment reduced the protein levels of DLGAP5, NLRP3, METTL3, GSDMD-N, Cleaved CASPASE1, mature IL-1β and mature IL-18 in ALI mouse liver (n = 6 per group). **(F, G)** IHC staining of GSDMD-N, Cleaved CASPASE1 and NLRP3 in liver sections from indicated groups (Scale bar: 50μm, n = 6 per group). **(H)** Serum pyroptosis index from indicated groups (n = 6 per group). All results are shown as mean ± SEM. \*p < 0.05. ns, not significant.

**Figure S5. CCl<sub>4</sub> treatment affects cellular communication in the liver.** **(A)** Signaling pathways related to cell communication were enriched according to the gene set enrichment analysis (GSEA). **(B)** Signaling pathways related to macrophage activation was screened by the GO analysis. **(C)** Cell clusters were identified and visualized by sn-RNA-seq. **(D)** The bar chart showed the proportion of cell types from healthy group and injury group.

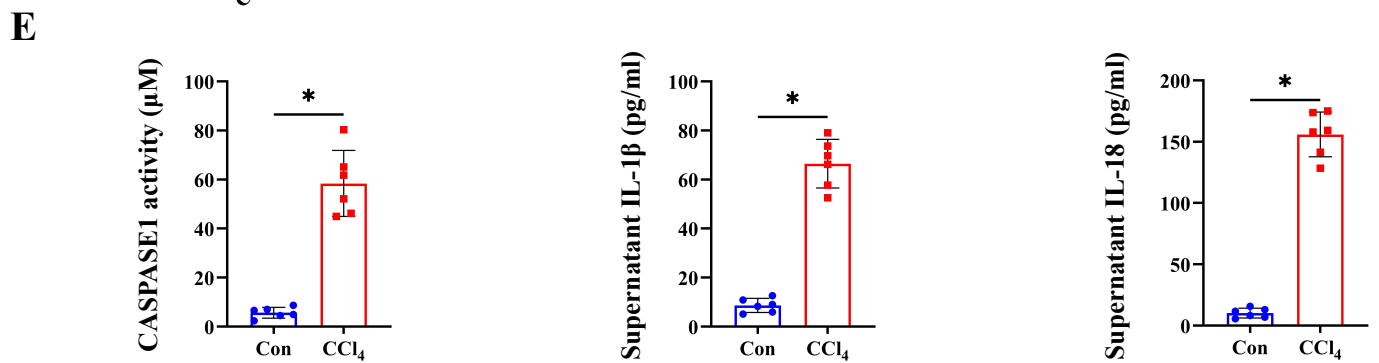
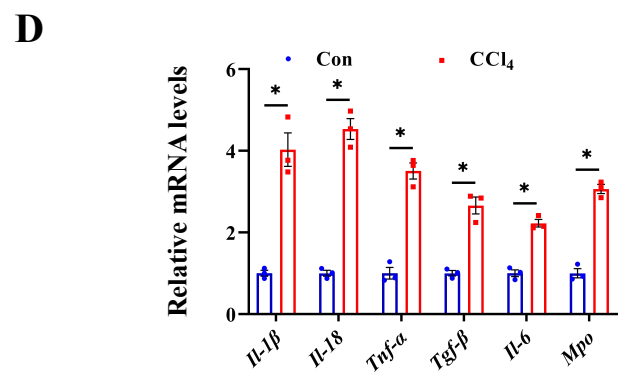
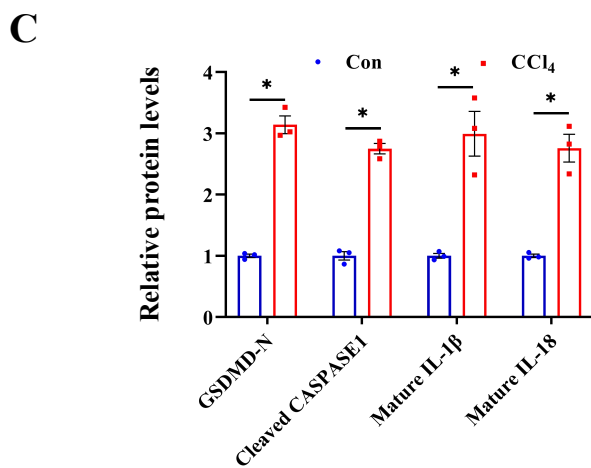
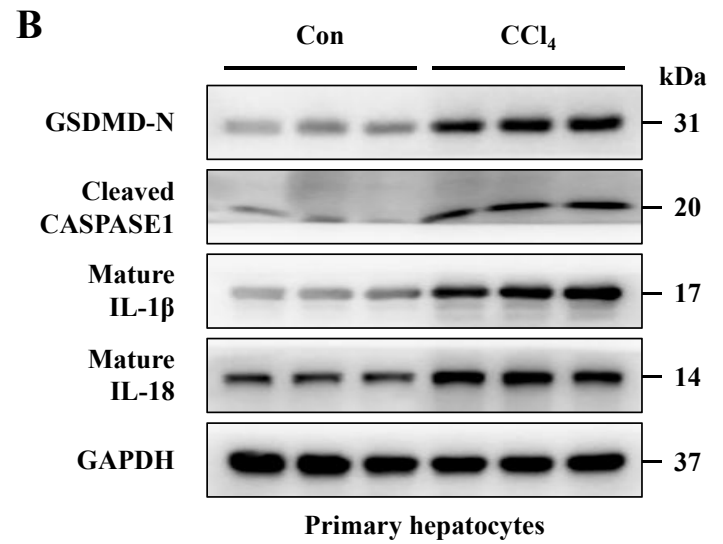
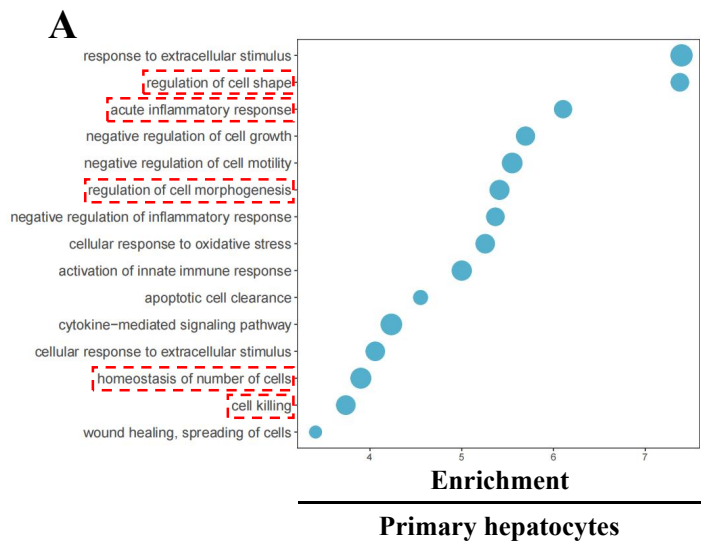
**Figure S6. Sufficient IL-1β or IL-18 signaling drives macrophage M1 polarization.** **(A)** Concentration of IL-1β and IL-18 in the supernatant of primary hepatocytes for co-culture (n = 6 per group) **(B)** iNOS fluorescence showing that sufficient supplement of either IL-1β (10 ng/ml) or IL-18 (20 ng/ml) further promoted macrophages M1 polarization by co-culture with primary hepatocytes from CCl<sub>4</sub>-treated mice (Scale bar: 25μm). **(C)** mRNA levels of *F4/80*, *Cd68* and *Nos2* in macrophages after PBS, IL-1β or IL-18 addition to the above co-culture system (n = 3 per group). **(D)** mRNA levels of *F4/80*, *Cd68* and *Nos2* in macrophages demonstrated that macrophage M1 polarization suppressed by hepatocyte *Nlrp3* knockdown in co-culture system was restored by adequate exogenous IL-1β (10 ng/ml) or IL-18 (20 ng/ml) addition (n = 3 per group) **(E)** iNOS fluorescence revealed macrophage M1 polarization suppressed by hepatocyte *Nlrp3*

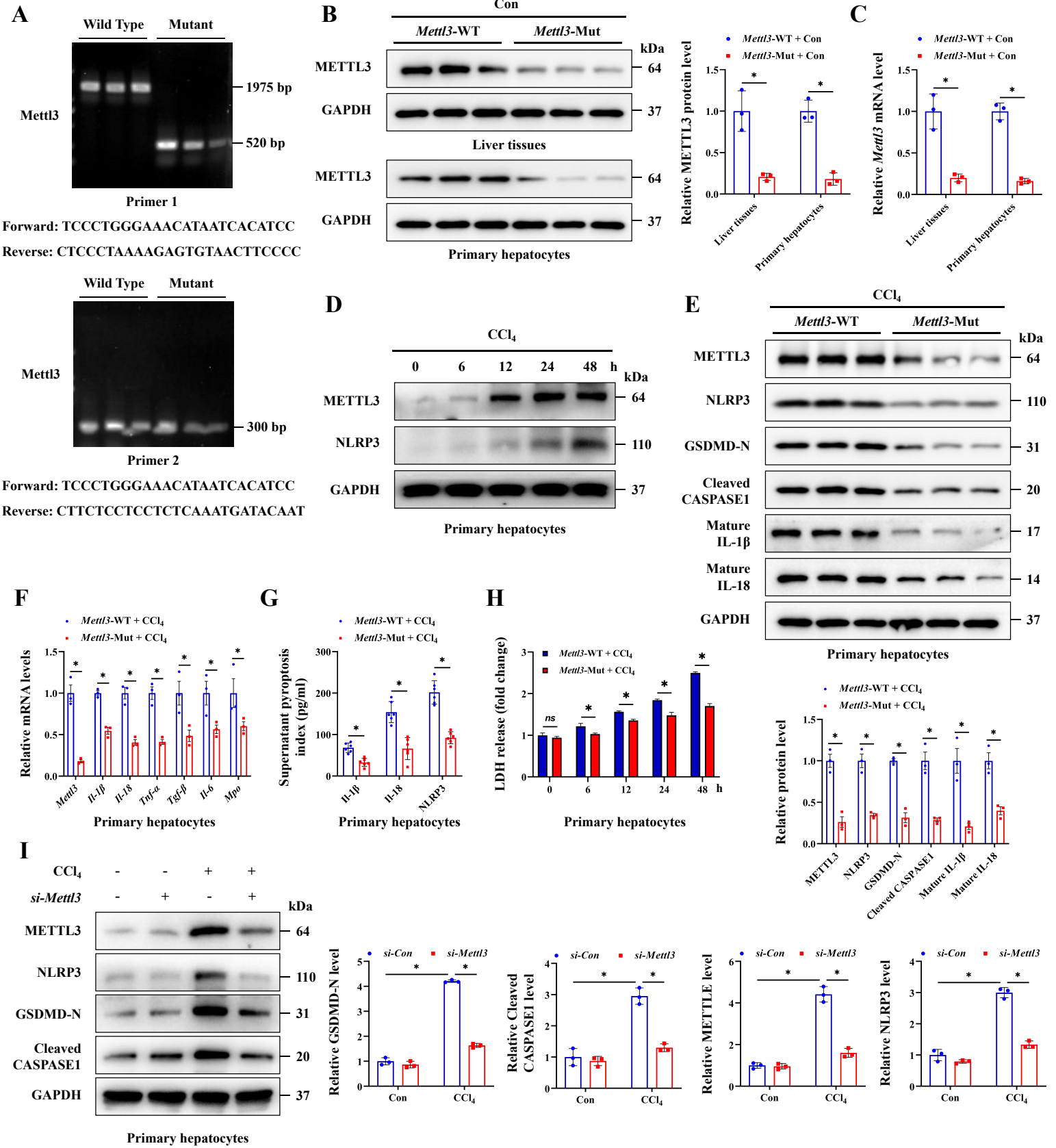
knockdown in co-culture system was restored by adequate exogenous IL-1 $\beta$  or IL-18 addition. All results are shown as mean  $\pm$  SEM. \* $p < 0.05$ .

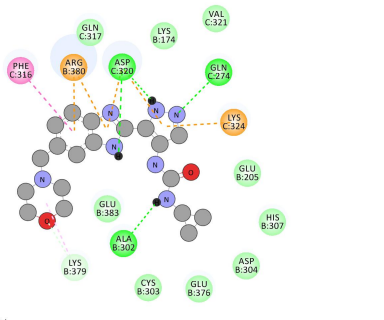
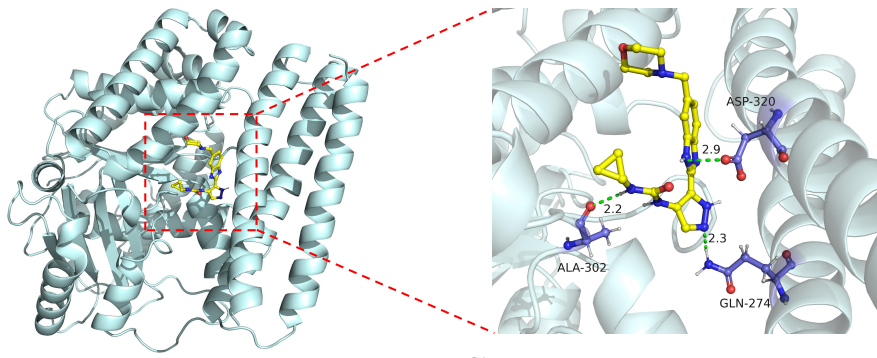
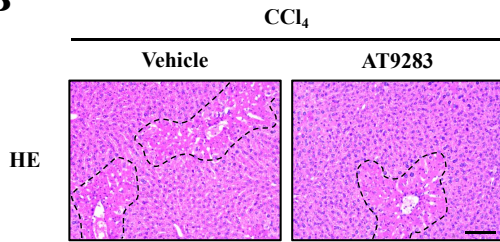
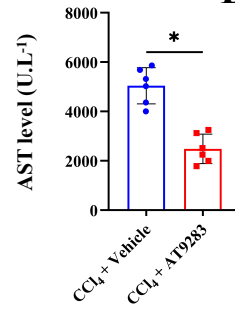
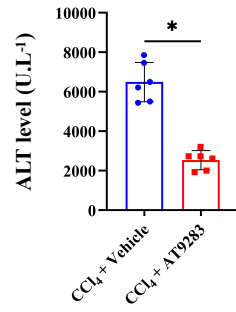
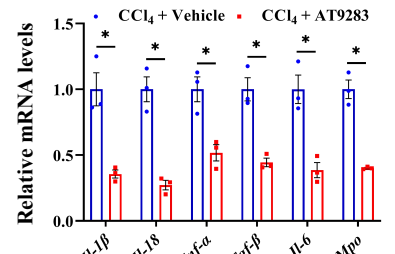
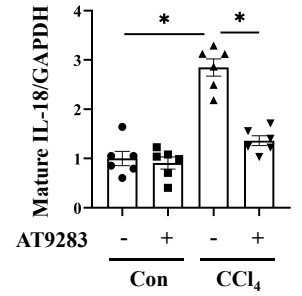
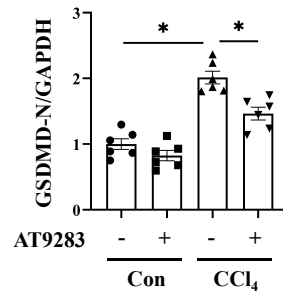
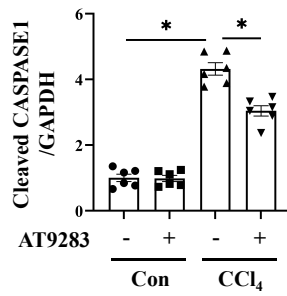
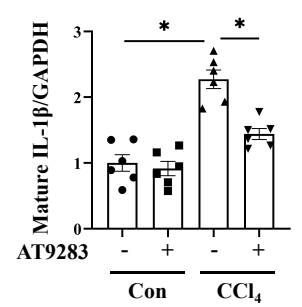
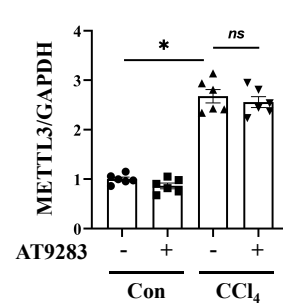
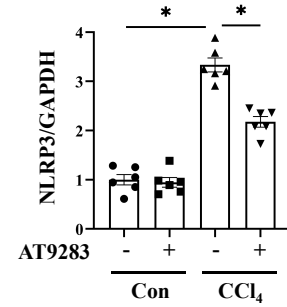
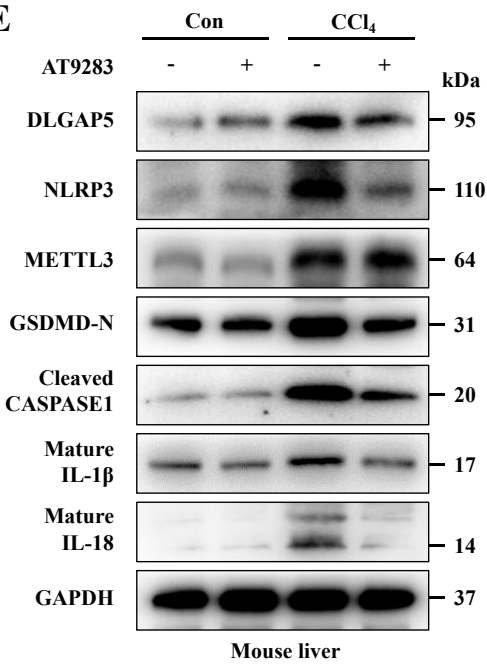
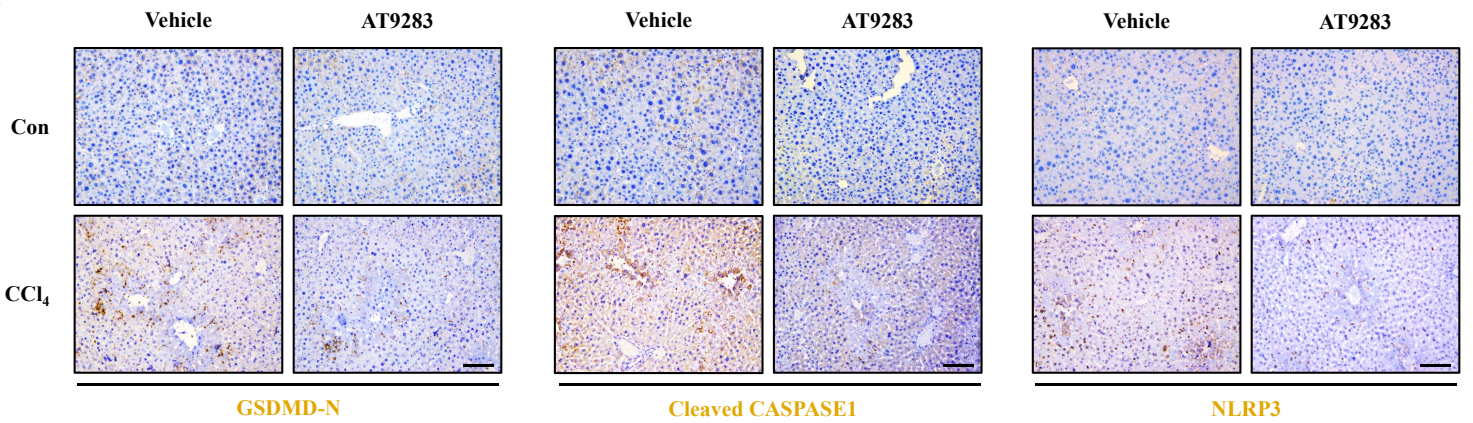
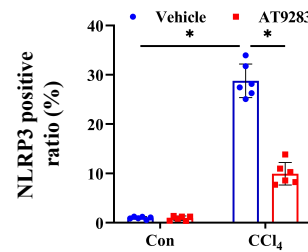
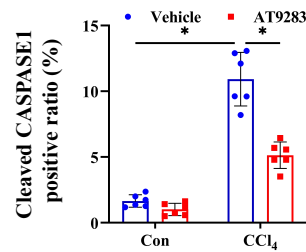
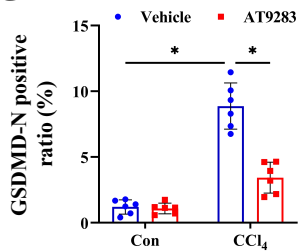
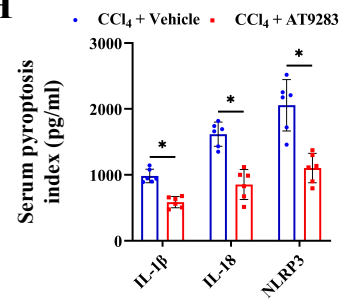
**Figure S7. Inhibition of hepatocyte pyroptosis alleviates macrophage M1 polarization and metabolic reprogramming.** (A) Heatmap of all genes involved in the top three up-regulated and down-regulated metabolism pathways according to Figure 9D. (B) iNOS fluorescence in primary macrophages showed that hepatocytes from CCl<sub>4</sub>-injured mice significantly enhanced M1 polarization of macrophages compared to hepatocytes from control mice, while the absence of *Nlrp3* in hepatocytes reversed that (Scale bar: 25 $\mu$ m). (C) mRNA levels of *F4/80*, *Cd68* and *Nos2* in primary macrophages from above groups under co-culture with hepatocytes with different treatments (n = 3 per group). (D, E) mRNA and protein levels of GLUT1, HK1, HK2 and LDHA in primary macrophages from indicated group (n = 3 per group). (F) Glucose uptake, lactate production and ATP production in primary macrophages from indicated group (n = 5 per group). (G) Measurement of ECAR and OCR in above macrophages after co-culture with hepatocytes from mice with different treatments. All results are shown as mean  $\pm$  SEM. \* $p < 0.05$ , # $p < 0.05$ .





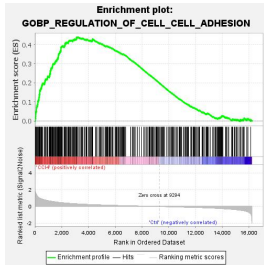




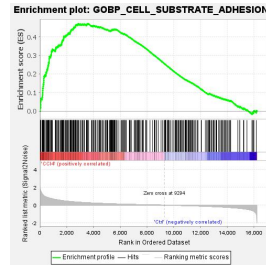
**A****B****C****D****E****F****G****H**

A

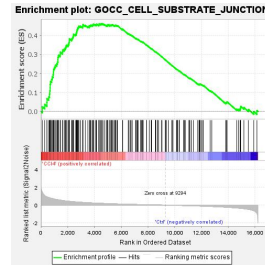
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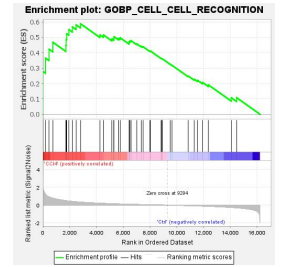
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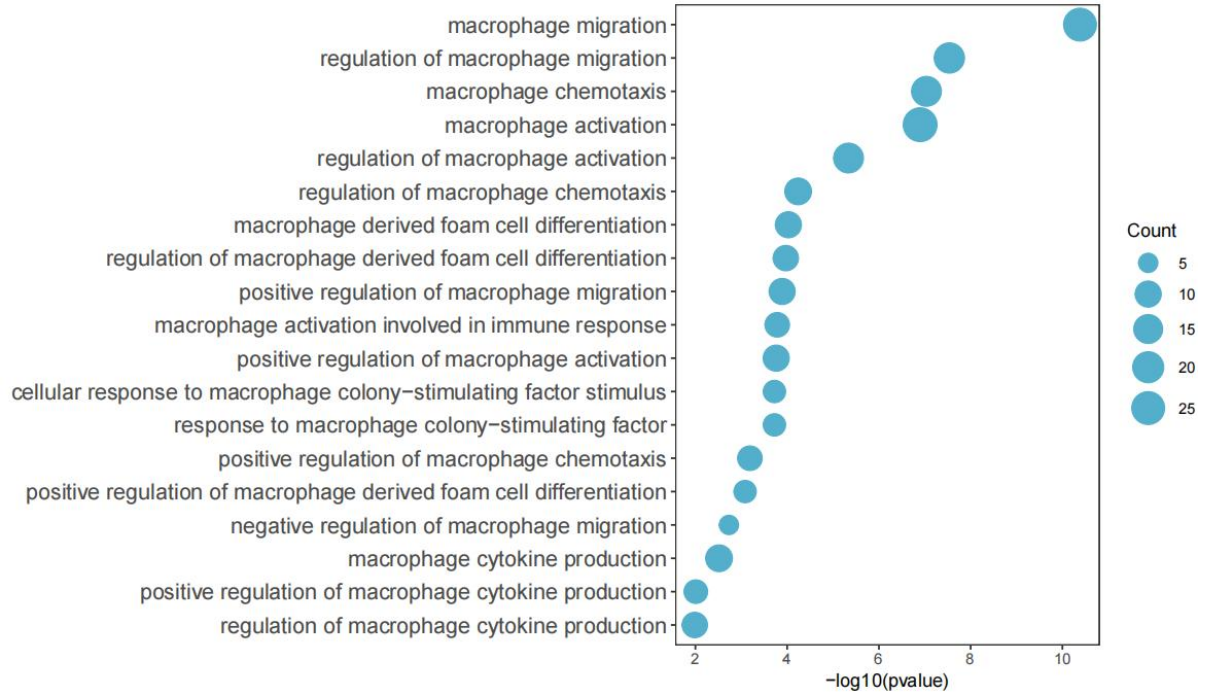
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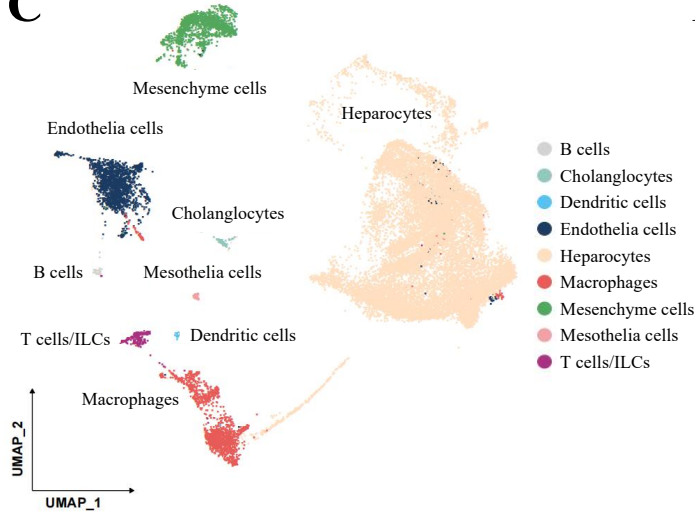
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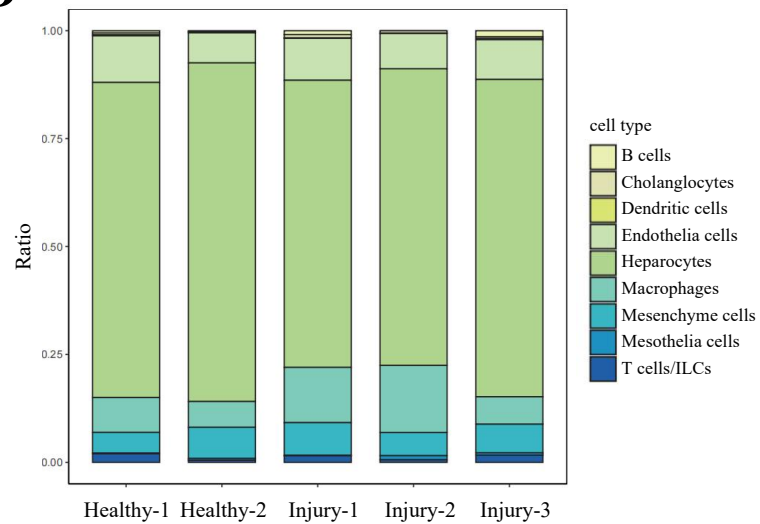
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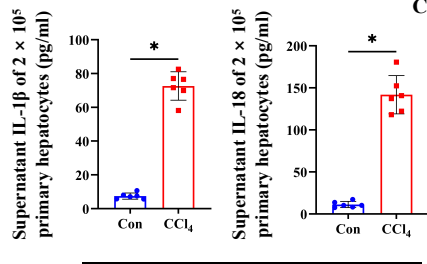


D





A



B

Co-culture system

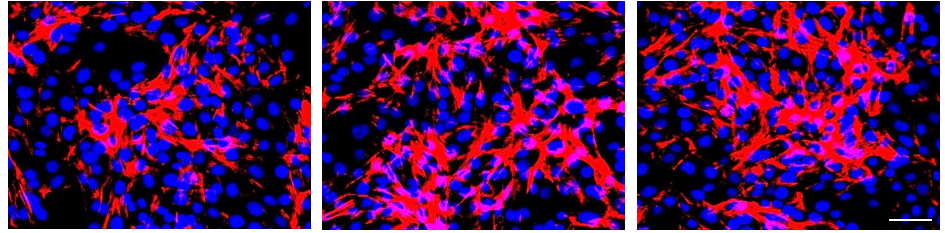
DAPI/iNOS

Hepatocytes from CCl<sub>4</sub>-treated mice

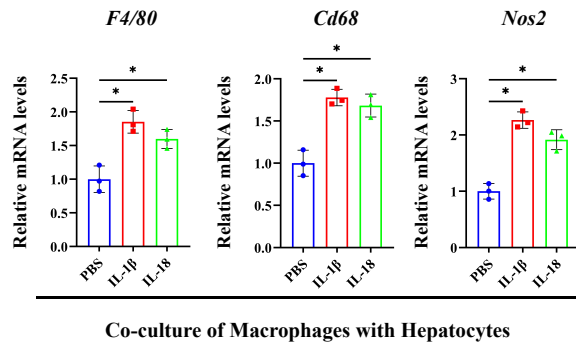
PBS

IL-1 $\beta$  (10 ng/ml)

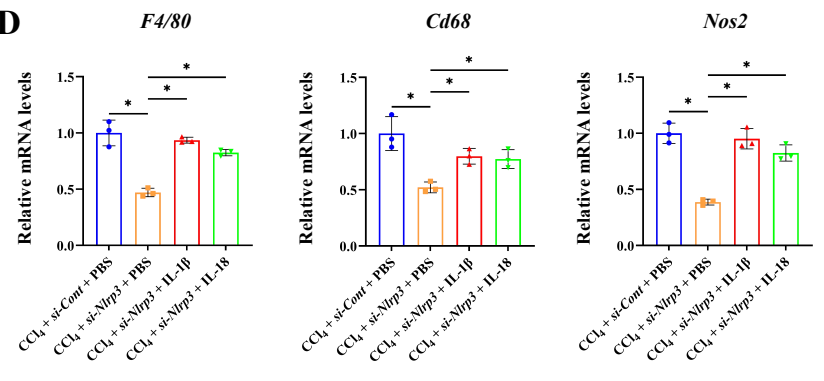
IL-18 (20 ng/ml)



C



D



E

