

Supplementary Data

Supplementary Figures

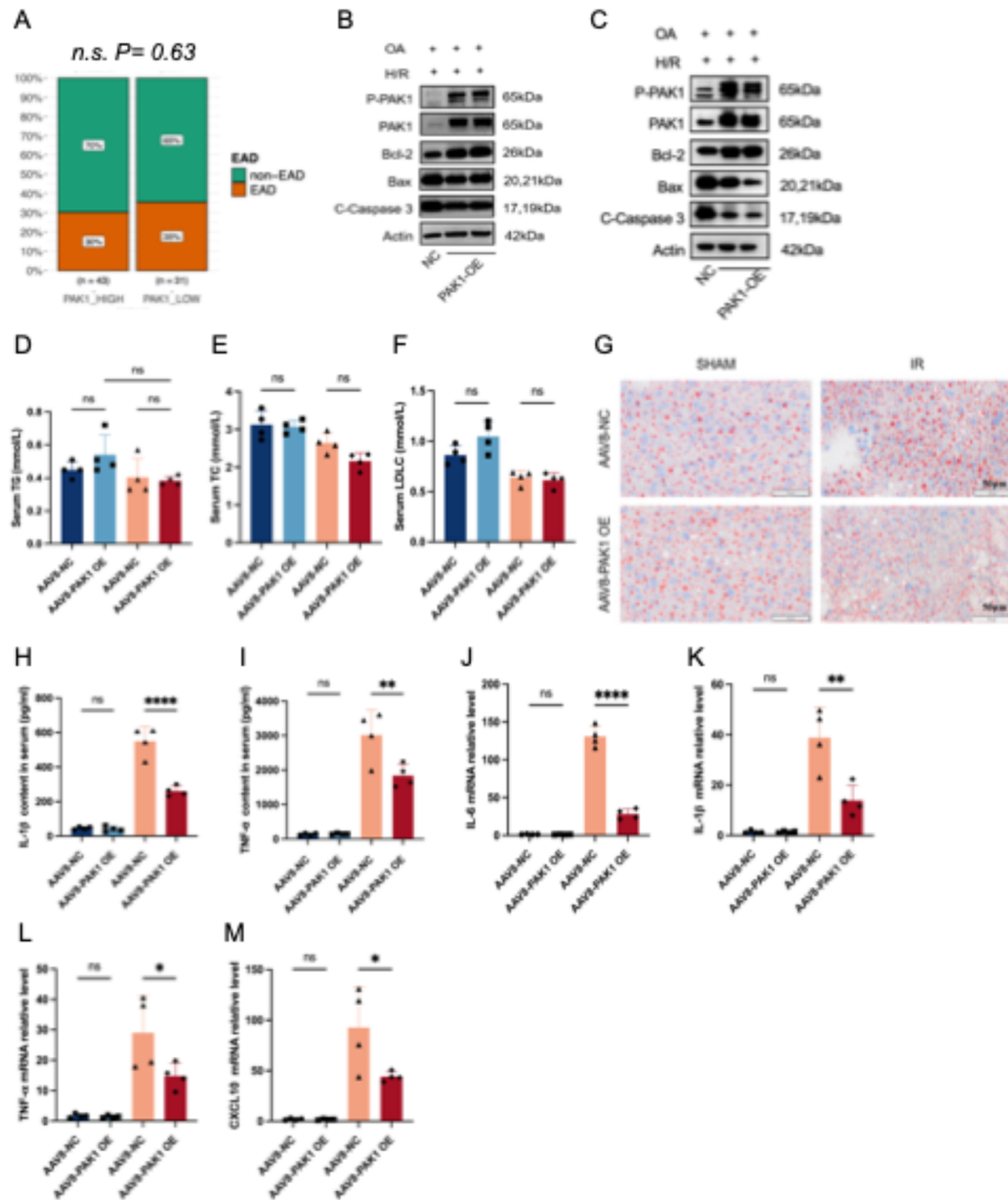


Figure S1 Clinical EAD analysis and complementary assessment of PAK1 overexpression in steatotic hepatic ischemia-reperfusion.

(A) Incidence of early allograft dysfunction (EAD) in PAK1-high (n = 43 grafts) and PAK1-low (n = 31 grafts) groups. (B, C) Additional representative immunoblots of P-PAK1, PAK1, Bcl-2, Bax, and cleaved caspase-3 in OA-treated AML12-NC and AML12-PAK1-OE cells after H/R; actin was the loading control. (D-F) Serum triglyceride, total cholesterol, and low-density lipoprotein cholesterol in AAV8-NC and

AAV8-PAK1-OE mice after sham surgery or IR (n = 4 mice per group). (G) Representative Oil Red O staining after sham surgery or IR. Scale bar, 50 μ m. (H, I) Serum IL-1 β and TNF- α after sham surgery or IR (n = 4 mice per group). (J-M) Hepatic IL-6, IL-1 β , TNF- α , and CXCL10 mRNA after sham surgery or IR (n = 4 mice per group). Three independent cell experiments for Figure S1 B, C. One in vivo experiment using biologically independent mice. Data are mean $\pm$ SD. Fisher's exact test was used for A, and one-way ANOVA for D–F and H–M. ns, not significant; *P < 0.05; **P < 0.01; ****P < 0.0001. EAD, early allograft dysfunction; OA, oleic acid; H/R, hypoxia/reoxygenation; IR, ischemia-reperfusion; TG, triglyceride; TC, total cholesterol; LDL-C, low-density lipoprotein cholesterol.

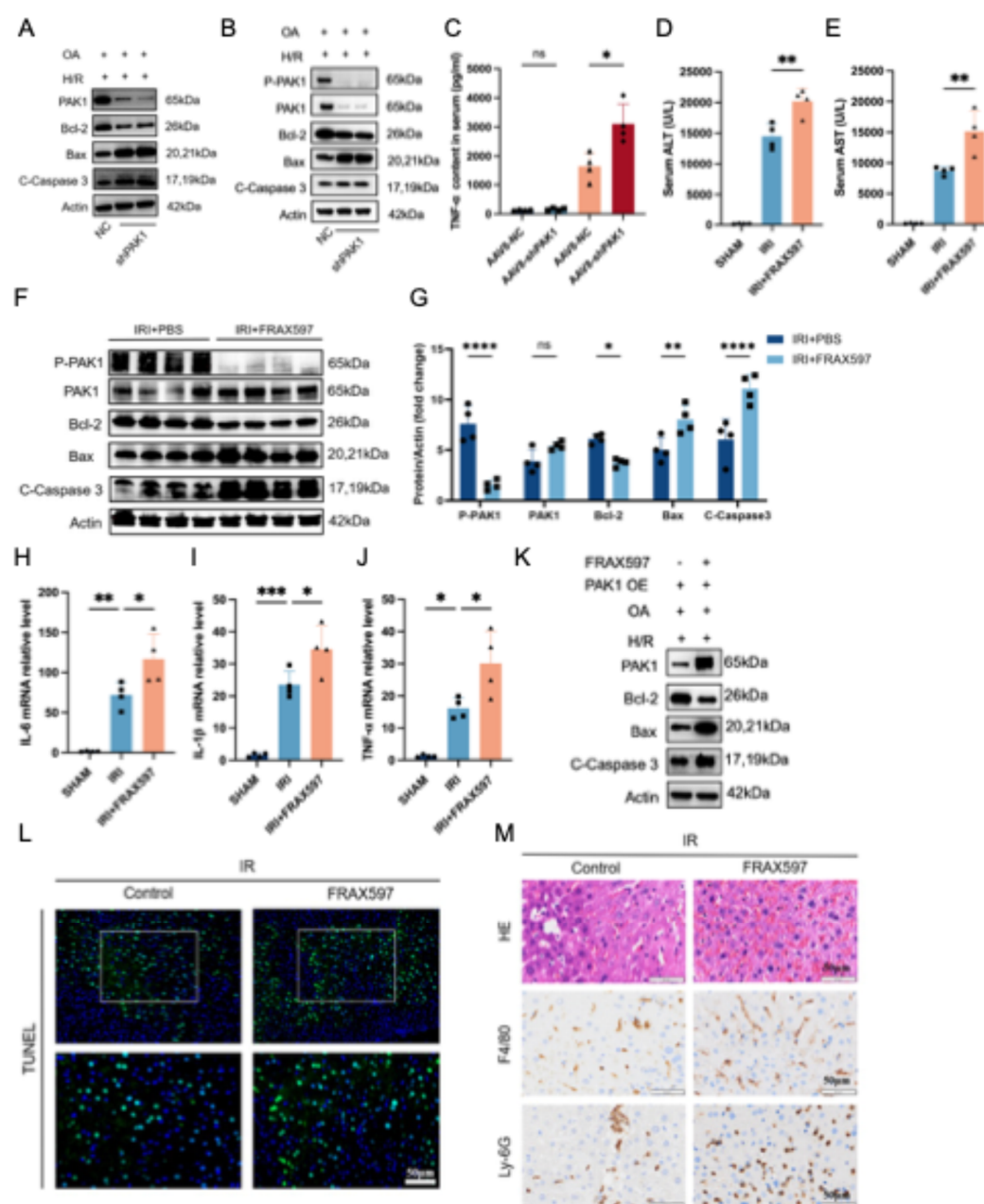


Figure S2 Complementary validation of PAK1 loss of function and pharmacological PAK inhibition during hepatic ischemia-reperfusion.

(A, B) Additional representative immunoblots of PAK1, P-PAK1, Bcl-2, Bax, and cleaved caspase-3 in OA-treated AML12-NC and AML12-shPAK1 cells after H/R; actin was the loading control. (C) Serum TNF- α in AAV8-NC and AAV8-shPAK1 mice after sham surgery or IR (n = 4 mice per group). (D, E) Serum ALT and AST in sham, IR, and IR+FRAX597 groups (n = 4 mice per group). (F, G) Representative liver immunoblots and quantification of P-PAK1, PAK1, Bcl-2, Bax, and cleaved caspase-3 after IR with vehicle or FRAX597 (n = 4 mice per group); actin was the loading control. (H-J) Hepatic IL-6, IL-1 β , and TNF- α mRNA in sham, IR, and IR+FRAX597 groups (n = 4 mice per group). (K) Representative immunoblots of PAK1, Bcl-2, Bax, and cleaved caspase-3 in PAK1-OE AML12 cells after H/R with or without FRAX597. (L) Representative TUNEL staining after IR with vehicle or FRAX597. Scale bar, 50 μ m. (M) Representative H&E, F4/80, and Ly6G staining after IR with vehicle or FRAX597. Scale bar, 50 μ m. Three independent cell experiments for Figure S2 A, B. One in vivo experiment using biologically independent mice. Data are mean $\pm$ SD. One-way ANOVA was used for C–E and H–J, and two-way ANOVA for G. ns, not significant; *P < 0.05; **P < 0.01; ***P < 0.001; ****P < 0.0001. FRAX597, PAK inhibitor; IR, ischemia-reperfusion; ALT, alanine aminotransferase; AST, aspartate aminotransferase.

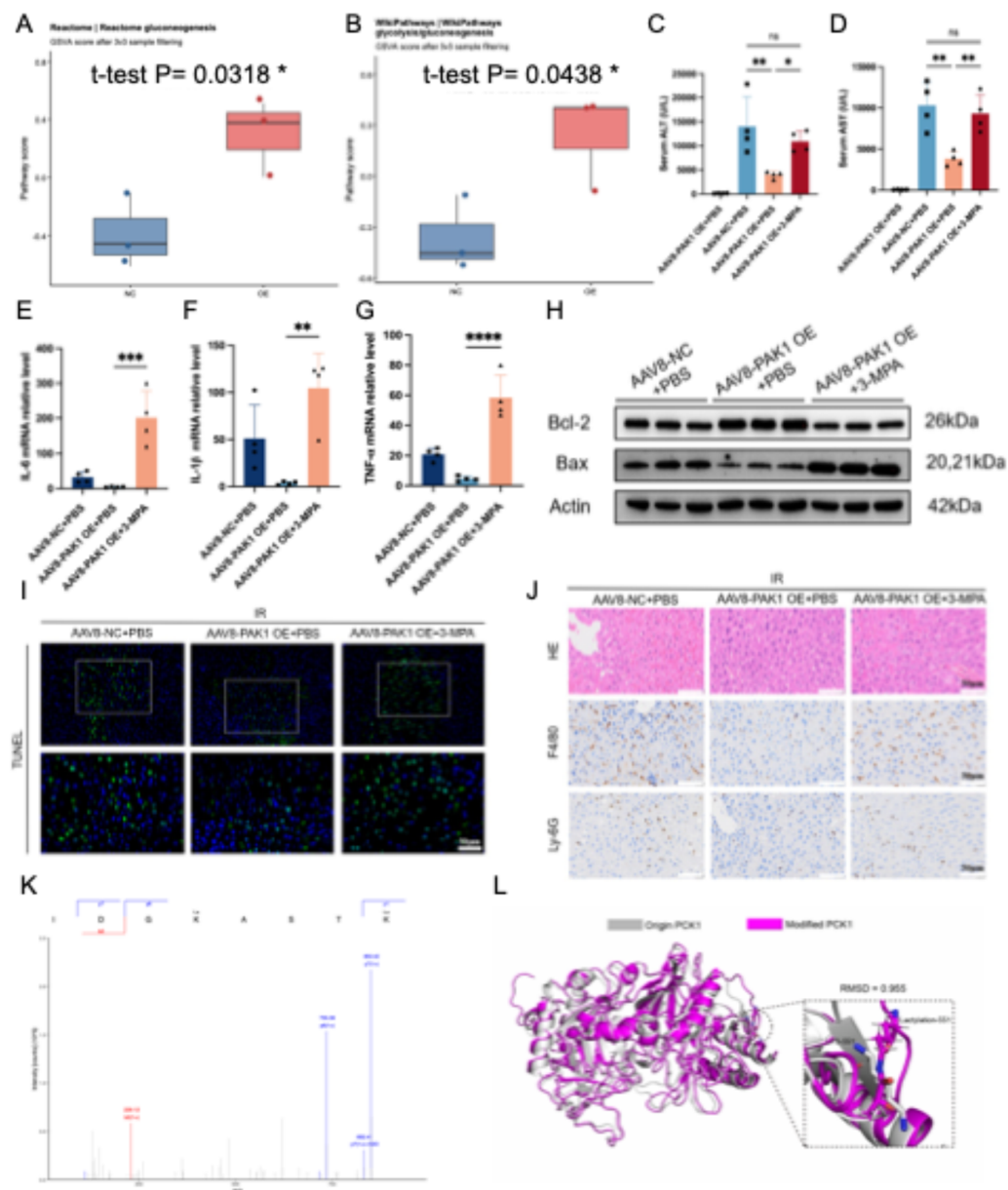


Figure S3 Pharmacological PCK1 inhibition attenuates the protective effects of hepatic PAK1 overexpression during steatotic hepatic ischemia-reperfusion.

(A, B) GSVA pathway scores for Reactome gluconeogenesis and WikiPathways glycolysis/gluconeogenesis in OA-treated control and PAK1-OE AML12 cells after H/R ($n = 3$ samples per group). (C, D) Serum ALT and AST levels in the four indicated mouse groups ($n = 4$ mice per group). (E-G) Hepatic IL-6, IL-1 β , and TNF- α mRNA levels in AAV8-NC+PBS, AAV8-PAK1-OE+PBS, and AAV8-PAK1-OE+3-MPA mice after IR ($n = 4$ mice per group). (H) Representative liver immunoblots of Bcl-2 and Bax in the same groups ($n = 3$ mice per group); actin was the loading control. (I) Representative TUNEL staining of liver sections from the indicated groups after IR, shown as overview images and enlargements of the boxed regions ($n = 4$ mice per group). Scale bar, 50 μ m. (J) Representative H&E, F4/80, and Ly6G staining of liver

sections from the indicated groups after IR (n = 4 mice per group). Scale bar, 50 μ m. (K) Representative LC-MS/MS spectrum of a PCK1 peptide carrying lactylation at K547 and K551. (L) Structural superposition of native and K551-lactylated PCK1, with the modified region enlarged (RMSD = 0.955). One RNA-seq/GSVA experiment with three samples per group. Data in C-G are mean $\pm$ SD, with individual data points shown. Unpaired two-sided Student's t tests were used for A and B (P = 0.0318 and P = 0.0438, respectively), and one-way ANOVA was used for C-G. ns, not significant; *P < 0.05; **P < 0.01; ***P < 0.001; ****P < 0.0001. GSVA, gene set variation analysis; OA, oleic acid; H/R, hypoxia/reoxygenation; IR, ischemia-reperfusion; AAV8, adeno-associated virus 8; NC, negative control; OE, overexpression; PBS, phosphate-buffered saline; 3-MPA, 3-mercaptopicolinic acid; ALT, alanine aminotransferase; AST, aspartate aminotransferase; TUNEL, terminal deoxynucleotidyl transferase dUTP nick-end labeling; H&E, hematoxylin and eosin; PCK1, phosphoenolpyruvate carboxykinase 1; LC-MS/MS, liquid chromatography-tandem mass spectrometry; RMSD, root-mean-square deviation.

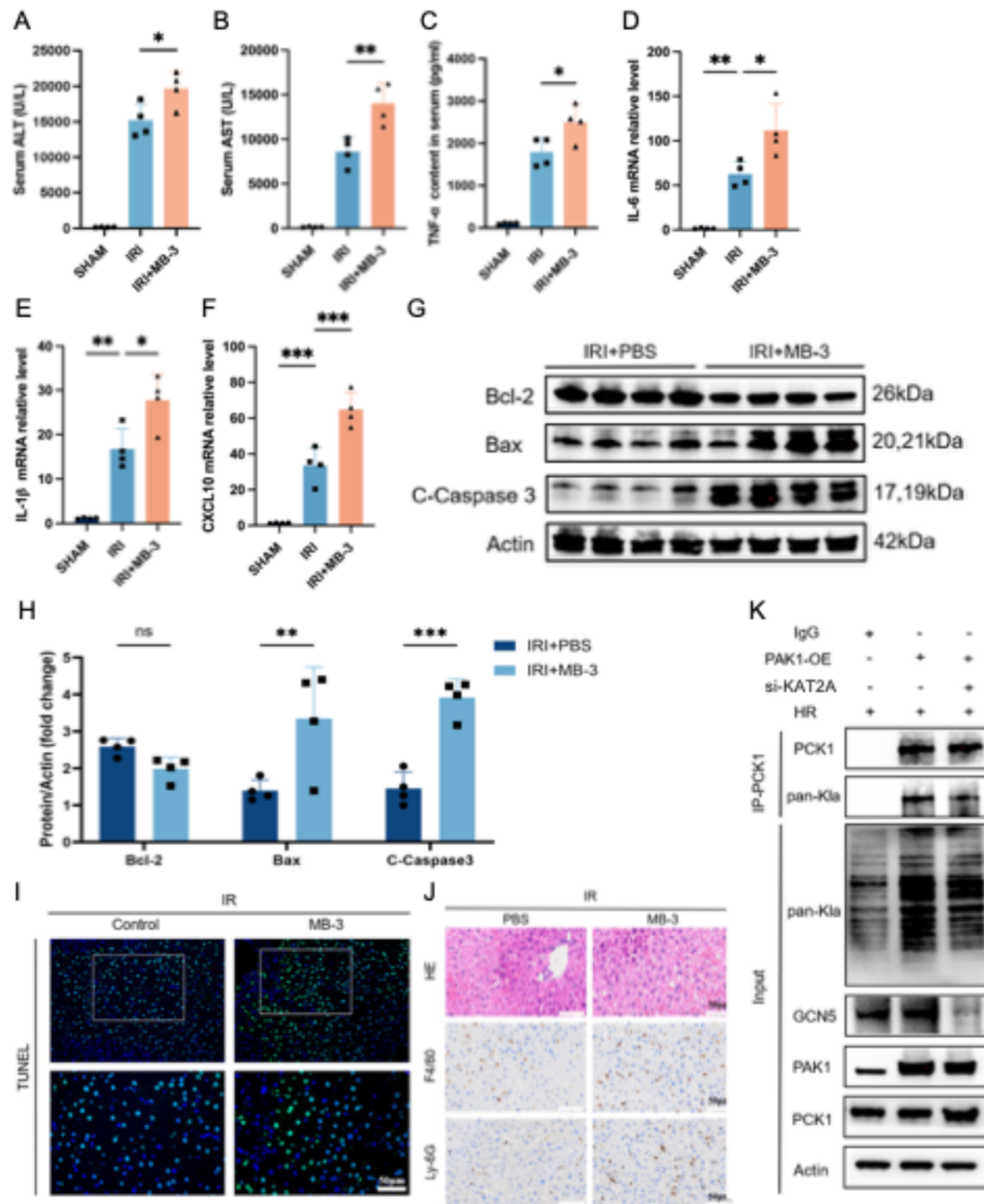


Figure S4 KAT2A inhibition exacerbates hepatic ischemia-reperfusion injury and reduces PAK1-associated PCK1 lactylation.

(A-C) Serum ALT, AST, and TNF- α in sham, IR, and IR+MB-3 groups (n = 4 mice per group). (D-F) Hepatic IL-6, IL-1 β , and CXCL10 mRNA in the same groups (n = 4 mice per group). (G, H) Representative liver immunoblots and quantification of Bcl-2, Bax, and cleaved caspase-3 after IR with vehicle or MB-3 (n = 4 mice per group); actin was the loading control. (I) Representative TUNEL staining after IR with vehicle or MB-3. Scale bar, 50 μ m. (J) Representative H&E, F4/80, and Ly6G staining after IR with vehicle or MB-3. Scale bar, 50 μ m. (K) PCK1 immunoprecipitation followed by pan-Kla immunoblotting in PAK1-OE AML12 cells after H/R with or without KAT2A knockdown; IgG and input controls are shown. One in vivo experiment using

biologically independent mice. Data are mean $\pm$ SD. One-way ANOVA was used for A-F, and two-way ANOVA for H. ns, not significant; * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$. MB-3, KAT2A inhibitor; KAT2A (also known as GCN5), lysine acetyltransferase 2A; PCK1, phosphoenolpyruvate carboxykinase 1; pan-Kla, pan-lysine lactylation.

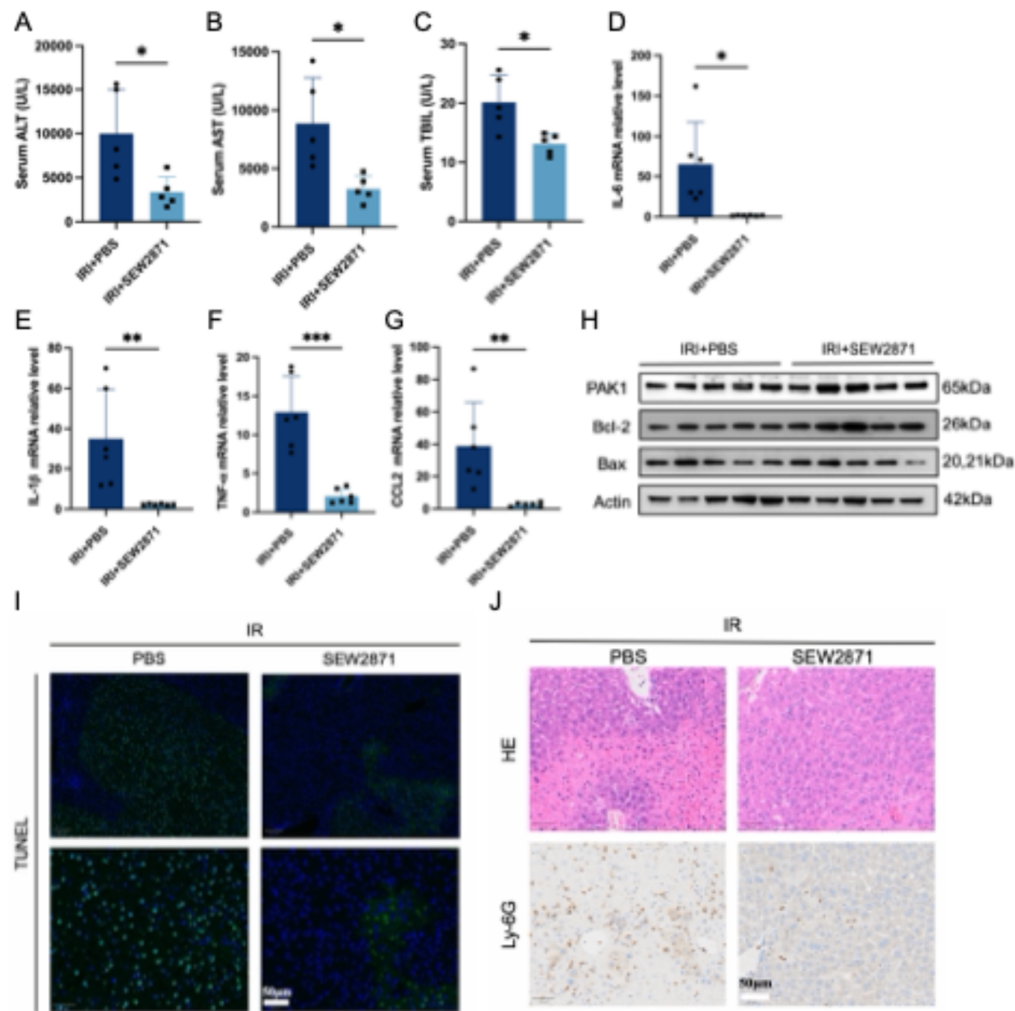


Figure S5 The S1PR1-selective agonist SEW2871 attenuates steatotic hepatic ischemia-reperfusion injury.

(A-C) Serum ALT, AST, and total bilirubin after IR with vehicle or SEW2871 ($n = 5$ mice per group). (D) Hepatic IL-6 mRNA after IR with vehicle or SEW2871 ($n = 5$ mice per group). (E-G) Hepatic IL-1 β , TNF- α , and CCL2 mRNA after IR with vehicle or SEW2871 ($n = 6$ mice per group). (H) Representative liver immunoblots of PAK1, Bcl-2, and Bax after IR with vehicle or SEW2871 ($n = 5$ mice per group); actin was the loading control. (I) Representative TUNEL staining after IR with vehicle or SEW2871. Scale bars as indicated. (J) Representative H&E and Ly6G staining after IR with vehicle or SEW2871. Scale bar, 50 μ m. One in vivo experiment using the indicated numbers of biologically independent mice. Data are mean $\pm$ SD. Unpaired two-sided Student's t

tests were used for A-G. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$. S1PR1, sphingosine-1-phosphate receptor 1; ALT, alanine aminotransferase; AST, aspartate aminotransferase; TBIL, total bilirubin; IR, ischemia-reperfusion.

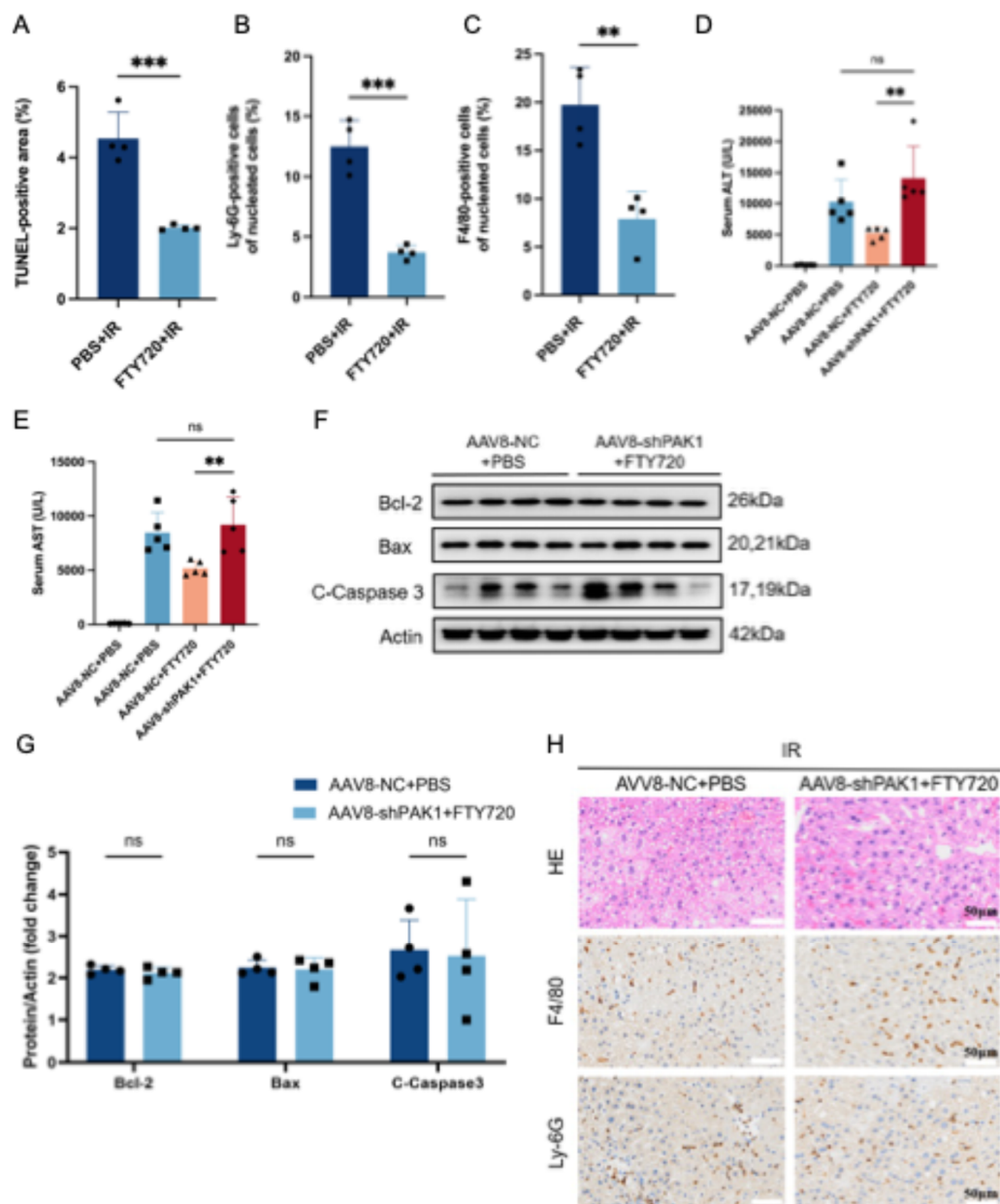


Figure S6 Hepatic PAK1 knockdown weakens the protective effects of FTY720 during steatotic hepatic ischemia-reperfusion.

(A-C) Quantification of TUNEL-positive area, Ly6G-positive cells, and F4/80-positive cells after IR with vehicle or FTY720 ($n = 4$ mice per group). (D, E) Serum ALT and AST in sham AAV8-NC+vehicle mice and in IR mice receiving AAV8-NC+vehicle, AAV8-NC+FTY720, or AAV8-shPAK1+FTY720 ($n = 5$ mice per group). (F, G) Representative liver immunoblots and quantification of Bcl-2, Bax, and cleaved

caspase-3 in IR AAV8-NC+vehicle and AAV8-shPAK1+FTY720 mice (n = 4 mice per group); actin was the loading control. (H) Representative H&E, F4/80, and Ly6G staining in the same groups. Scale bar, 50 μ m. One in vivo experiment using the indicated numbers of biologically independent mice. Data are mean $\pm$ SD. Unpaired two-sided Student's t tests were used for A-C, one-way ANOVA for D and E, and two-way ANOVA for G. ns, not significant; **P < 0.01; ***P < 0.001. FTY720, fingolimod; AAV8, adeno-associated virus 8; shPAK1, short hairpin RNA targeting PAK1; ALT, alanine aminotransferase; AST, aspartate aminotransferase.

Supplementary Tables

Table S1. Plasmids and expression vectors used in this study

Plasmids and Expression Vectors	Cat No.	Manufacture
pLV3-CMV-PAK1(mouse)-3 $\times$ FLAG-CopGFP-Puro	P85525	Miaoling Bio
pLV3-CMV-PAK1(human)-3 $\times$ FLAG-CopGFP-Puro	P47371	Miaoling Bio
pLV3-CMV-ACSS2(human)-3 $\times$ FLAG-EF1a-mCherry-Neo	P89668	Miaoling Bio
pCMV-ACSS2(human)-3 $\times$ HA-Neo	P50020	Miaoling Bio
pCMV-KAT2A(human)-3 $\times$ HA-WPRE-Neo	P92970	Miaoling Bio
pLV3-CMV-KAT2A(human)-3 $\times$ FLAG-CopGFP-Puro	P81889	Miaoling Bio
pLV3-CMV-PCK1(human)-3 $\times$ HA-Puro	P87757	Miaoling Bio
pLV3-CMV-PCK1(mouse)-3 $\times$ FLAG-CopGFP-Puro	P57261	Miaoling Bio
pLV3-CMV-ACSS2 S267A(human)-3 $\times$ FLAG-Neo	-	Miaoling Bio
pLV3-CMV-ACSS2 S659A(human)-3 $\times$ FLAG-Neo	-	Miaoling Bio

Table S2. Primers for real-time qPCR detection

Gene		Sequence5'---3'
Mouse IL-6	F	CTGCAAGAGACTTCCATCCAG
	R	AGTGGTATAGACAGGTCTGTTGG
Mouse IL-1 β	F	TTCAGGCAGGCAGTATCACTC
	R	GAAGGTCCACGGGAAAGACAC
Mouse TNF- α	F	CAGGCGGTGCCTATGTCTC
	R	CGATCACCCCGAAGTTCAGTAG
Mouse β -actin	F	GTGACGTTGACATCCGTAAAGA
	R	GCCGGACTCATCGTACTCC
Mouse CXCL10	F	AATGAGGGCCATAGGGAAGC

	R	AGCCATCCACTGGGTAAAGG
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Table S3. Antibodies for immunoblot analyses

Antibody	Cat No.	Manufacture
Cleaved caspase 3	9661S	CST
Bax	2772S	CST
Bcl2	ab182858	Abcam
PAK1	21401-1-AP	Proteintech
P-PAK1	2601S	CST
β-actin	66009-1-Ig	Proteintech
Flag	66008-4-Ig	Proteintech
HA	51064-2-AP	Proteintech
ACSS2	16087-1-AP	Proteintech
Pan Phospho-Serine/Threonine	T91067	Abmart
Anti-L-Lactyl Lysine Rabbit mAb	1401RM	PTM